

Research article

Open Access

In vitro antimicrobial activity of natural toxins and animal venoms tested against *Burkholderia pseudomallei*

R Perumal Samy¹, A Pachiappan¹, P Gopalakrishnakone*¹,
Maung M Thwin¹, Yap E Hian², Vincent TK Chow², Ho Bow² and
Joseph T Weng²

Address: ¹Venom and Toxin Research Programme, Department of Anatomy, Yong Loo Lin School of Medicine, National University of Singapore, Singapore - 117597 and ²Department of Microbiology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore - 117597

Email: R Perumal Samy - antrps@nus.edu.sg; A Pachiappan - antap@nus.edu.sg; P Gopalakrishnakone* - antgopal@nus.edu.sg; Maung M Thwin - antmmt@nus.edu.sg; Yap E Hian - mcye@nus.edu.sg; Vincent TK Chow - mchb@nus.edu.sg; Ho Bow - mctch@nus.edu.sg; Joseph T Weng - mctw@nus.edu.sg

* Corresponding author

Published: 20 June 2006

Received: 13 January 2006

BMC Infectious Diseases 2006, 6:100 doi:10.1186/1471-2334-6-100

Accepted: 20 June 2006

This article is available from: <http://www.biomedcentral.com/1471-2334/6/100>

© 2006 Perumal Samy et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: *Burkholderia pseudomallei* are the causative agent of melioidosis. Increasing resistance of the disease to antibiotics is a severe problem in treatment regime and has led to intensification of the search for new drugs. Antimicrobial peptides are the most ubiquitous in nature as part of the innate immune system and host defense mechanism.

Methods: Here, we investigated a group of venoms (snakes, scorpions and honey bee venoms) for antimicrobial properties against two strains of Gram-negative bacteria *Burkholderia pseudomallei* by using disc-diffusion assay for *in vitro* susceptibility testing. The antibacterial activities of the venoms were compared with that of the isolated L-amino acid oxidase (LAO) and phospholipase A₂ (PLA₂s) enzymes. MICs were determined using broth dilution method. Bacterial growth was assessed by measurement of optical density at the lowest dilutions (MIC 0.25 mg/ml). The cell viability was measured using tetrazolium salts (XTT) based cytotoxic assay.

Results: The studied venoms showed high antimicrobial activity. The venoms of *C. adamantus*, *Daboia russelli russelli*, *A. halys*, *P. australis*, *B. candidus* and *P. guttata* were equally as effective as Chloramphenicol and Ceftazidime (30 µg/disc). Among those tested, phospholipase A₂ enzymes (crotoxin B and daboitoxin) showed the most potent antibacterial activity against Gram-negative (TES) bacteria. Naturally occurring venom peptides and phospholipase A₂ proved to possess highly potent antimicrobial activity against *Burkholderia pseudomallei*. The XTT-assay results showed that the cell survival decreased with increasing concentrations (0.05–10 mg/mL) of *Crotalus adamantus* venom, with no effect on the cell viability evident at 0.5 mg/mL.

Conclusion: This antibacterial profile of snake venoms reported herein will be useful in the search for potential antibacterial agents against drug resistant microorganisms like *B. pseudomallei*.

Background

Gram-negative *Burkholderia pseudomallei*, the causative agent of melioidosis, are found widely in soil and surface water throughout the tropics. High incidence of melioidosis has been found particularly in Southeast Asia and Northern Australia [1,2]. A number of cases have been reported in Malaysia [3], Thailand [4], Northern Australia [5], South China [6], Taiwan [7], South India, Africa and America. The majority of adult patients develop acute pulmonary or septicaemic illness with high mortality rates [8], or subacute melioidosis, characterized by multiple-abscess formation. In cases of septicaemic melioidosis, which is associated with a vigorous inflammatory cytokine response [9], septic shock continues to be a major cause of morbidity and mortality in patients. Tumor necrosis factor (TNF) is involved in the acquisition of melioidosis, and is also related to disease severity [10].

Antibiotic resistance has been of great concern during the last decades due to the extensive clinical use of classical antibiotics [11]. Currently available antimicrobials fail to lower the mortality rate of melioidosis [12,13]. *B. pseudomallei* demonstrate high levels of resistance to the action of cationic antimicrobial peptides such as polylysine, protamine sulfate, human neutrophil peptides (HNP-1), and polymyxins [14,15]. Therefore, it is of considerable interest to develop antimicrobials with a new mechanism(s) of action which can potentially evade the emergence of drug resistance. In the search for such new agents, we are looking for effective bactericidal peptides from multi-cellular animals that have potential activity against *B. pseudomallei*. The antimicrobial peptides are ubiquitous in nature as part of the innate immune system and host defence mechanisms. They have been increasingly recognized as a critical first line of defence against many pathogens isolated from various sources [16].

Proteinacious components such as neurotoxin, cytotoxin, myotoxin, protease and nuclease are found in varying quantities in snake venoms depending on the species. It has been reported that the basic myotoxic PLA₂ enzymes, (myotoxin II) of *Bothrops asper* [17] and BnpTx I of *Bothrops neuwiedi pauloensis* [18], possess bactericidal activity against *E. coli* and *S. aureus*. The crude venom and enzymes (LAAO1 and LAAO2) from *Pseudechis australis* are also known to have antibacterial effects against gram (+) and gram (-) bacteria [19]. Besides the strong haemolytic activity shown against sheep erythrocytes, the magainin-type polycationic antimicrobial peptides (pandinin 1 & 2) from the venom of African scorpion *Pandinus imperator* demonstrate high antimicrobial activity against a range of gram (+) and gram (-) bacteria [20]. Moreover, anti-bacterial peptides called the cecropins and defensins have been identified in the hemolymph of scorpions [21]. They are active against bacteria through lysis of

their cells thus causing death of the microorganisms. Bactericidal peptides (lycotoxins) have also been identified in the venom of the spider *Lycosa singoriensis* [22], while another family of peptides (cupiennius) with bactericidal activity has been isolated from the venom of a different species of *Cupiennius salei* [23]. Melittin is an important venom component of *Apis mellifera* (common honey bee) which is more active against gram (+) and gram (-) bacteria [24,25]. The highly cationic and salt insensitive cathelicidins of mammalian origin are also reported to have a broad-spectrum of antimicrobial activities [26]. The peptides differ widely in sequence and structure, yet kill bacteria by thinning and disrupting the bacterial membrane [27], and as a result these peptides are important as unique source leads for the development of new therapeutic agents [28]. The above examples thus emphasize the fact that animal toxins or peptides can be rich sources of antimicrobials which may be useful as therapeutics in human.

Although a large number of bioactive components have previously been isolated and characterized from snake venoms, no systematic search for antibacterial components from animal venoms has yet been reported. In our previous studies, majority of the venoms exerted a broad spectrum of antimicrobial activity against various gram-negative and gram-positive bacteria (Journal of Applied Microbiology – under revision). We describe here the antimicrobial properties of 34 different venoms of scorpions, snakes and honey bee, along with those of nine purified venom phospholipase A₂ enzymes and two L-amino acid oxidase, evaluated against multi-drug resistant pathogen obtained from the patient with melioidosis (*B. pseudomallei* strains KHW and TES).

Methods

Venoms

Lyophilized venoms – *Acanthophis augtra*, *Acanthophis antarcticus*, *Acanthophis praelongus*, *Acanthophis Pyrrhus*, *Androctonus australis*, *Bungarus candidus*, *Hyrophis cyanocinctus*, *Naja naja naja*, *Notechis aterater*, *Naja sumatrana*, *Naja kaouthia*, *Pseudechis australis*, *Pseudechis guttata*, *Pseudechis porphyriacus*, *Pseudechis colletti*, *Pseudonaja inframagula*, *Pseudonaja nuchalis*, *Pseudonaja textilis*, *Pseudechis affinis*, *Rhabdophis tigrinus*, *Oxyuranus scutellatus*, *Agkistrodon halys*, *Bitis gabonica rhinoceros*, *Crotalus adamanteus*, *Echis carinatus*, *Daboia russelli russelli*, *Daboia russelli siamensis*, *Trimeresurus wagleri*, *Apis mellifera*, *Bothotus hottentota*, *Buthotus hottentota hottentota*, *Buthus martensii* and *Naja naja naja* venoms were extracted from long-term captive specimens. Venoms from captive specimens were collected manually by milking. Each sample of freeze-dried venom was stored at 4°C. The solid venoms were obtained from commercial sources (Venom supplies Pte Ltd, Tanunda, South Australia). L-amino acid oxidase

purified from the venom of *B. atrox* and *C. adamanteus* were obtained commercially (Sigma Aldrich, St Louis, MO, USA).

Antibacterial effects of venoms

Each sample of freeze-dried venom was dissolved in 1 mL of 50 mM Tris-HCl buffer (pH 7.4), vortexed (Labnet VX100), and filtered using 0.22 µm syringe filter (Millipore, NY, USA) before storage at 4 °C. All the venoms were tested for their antibacterial activity against gram-negative bacteria *Burkholderia pseudomallei* (KHW) and *Burkholderia pseudomallei* (TES) by disc-diffusion susceptibility tests were performed following standards recommended by the NCCLS [29] with some modifications. Bacterial inoculums (200 µL of a 0.1 A₆₀₀ culture containing 3.2 × 10⁸ colony forming units cfu/mL) were spread by using a sterile cotton swab onto 20 mL of sterile TS agar plates (90 mm diameter). The surface of the medium was allowed to dry for about 3 min. Sterile paper discs (7 mm diameter) were then placed onto the TS agar surface and 20 µL (0.1 mg/mL) of venom sample was added per disc in 5 replicates. Disc containing 20 µL of Tris-HCl buffer served as a normal control and discs containing antibiotics were used as drug controls. The plates were incubated at 37 °C for 24 h, following which the diameter of inhibition zones were measured and calculated, and the resultant activities compared with purified PLA₂ enzymes.

Purification of phospholipase A₂ (PLA₂) enzymes

Phospholipase A₂ enzymes were purified from their corresponding crude venoms as described [30] with minor modification. In brief, the venom (0.4 g/4 ml) was fractionated on a Superdex-G75 column with Tris-HCl buffer (50 mM, pH 7.4) as an eluant, followed by further purification on a RP-HPLC Jupiter C18 column (AKTA explorer Workstation, Amersham Pharmacia Biotech, Sweden), eluted with a linear gradient of 80% acetonitrile in 0.1% trifluoroacetic acid. Elution of proteins was monitored at 280 nm and 215 nm. The homogeneity of purified PLA₂s was determined by using MALDI-TOF on a Voyager DE-STR Biospectrometry workstation (Applied Biosystem CA, USA).

Antibacterial effects of purified PLA₂

All the venom enzymes used in the experiment have been purified by successive chromatographic steps with the final purity of at least 95% as assessed by Reversed-phase HPLC. The activities of the following phospholipase A₂ enzymes purified from the venoms of different snake species (in parenthesis) – crotoxin A (*Crotalus durissus terrificus*), crotoxin B (*Crotalus durissus terrificus*), ammodytoxin A (*Vipera ammodytes ammodytes*), mojave toxin (*Crotalus scutulatus scutulatus*), β-bungarotoxin (*Bungarus multicinctus*), taipoxin (*Oxyuranus scutellatus scutellatus*), mulgatoxin (*Pseudechis australis*), *Daboia* toxin (*Daboia russelli*

russelli), honey bee (*Apis mellifera*) venom phospholipase A₂ – and two L-amino acid oxidase – LAAO (*B. atrox*), LAAO (*C. adamanteus*) were evaluated. 0.5 µmoles of each of the enzyme dissolved in 500 µL of 50 mM Tris-HCl (pH 7.4) buffer was examined. *In vitro* antimicrobial activity was determined by the previously described disc-diffusion method [29] with some modifications.

MIC determinations

In preliminary experiments, the disc-diffusion assay for determining antibacterial effects of crude venoms was compared to the activity of isolated phospholipase A₂ (PLA₂) enzymes using broth-microdilution method. Differences in data between these two assays were observed. Thus, we used the broth-microdilution technique to test the activities of the phospholipase A₂ against pathogen from patients by comparing their activities to the activity of the antibiotics. Bacteria from frozen suspensions were sub-cultured onto Tryptic soya (TS) agar plates and passaged twice prior to susceptibility testing. The bacteria were grown in Tryptic soy broth (TSB) for 5 – 7 h (exponential phase) before adjusting their concentration to a 0.5 McFarland turbidity standard. The adjusted bacterial cultures were diluted to approximately (A₆₀₀ of 0.8) 3.2 × 10⁸ colony forming units (cfu/mL). The enzymes (PLA₂s) to be examined were dissolved in 1 M Tris-HCl buffer (pH 7.4) for determination of the activities (MIC). The bacteria were washed and incubated with the enzymes in appropriate buffer. MICs were determined by the broth microdilution method recommended by the NCCLS [31,32], for which serial dilutions of each enzyme solutions were prepared (final concentration 0.5, 0.25, 0.125, 0.0625 and 0.03125 mg/mL) in microtiter trays with appropriate medium (TSB). Each dilution series included control wells containing bacteria without enzymes. A total of 200 µL of the adjusted inoculum of 10⁵ cfu/mL was added to each well (96-well plates). The culture trays were incubated at 37 °C for 24 h with shaking at 230 rpm. The inhibition of bacterial growth was determined by measuring the absorbance at 560 nm (Molecular Devices E precision microplate reader). The MIC was taken as the lowest concentration of phospholipase A₂ enzyme that inhibited visible growth of the organism. The results given are mean values of three independent determinations.

Biochemical characterization

Phospholipase A₂ enzyme activity

Phospholipase A₂ (PLA₂) catalyzes the hydrolysis of phospholipids at the *sn*-2 position yielding a free fatty acid and a lysophospholipid. The Cayman Chemical secretory PLA₂ (sPLA₂) assay kit was used for the measurement of enzyme activity. This assay uses the 1, 2-dithio analog of diheptanoyl phosphatidylcholine which serves as a substrate for most PLA₂s [33]. sPLA₂ specific activity is expressed as µmole/min/mg protein.

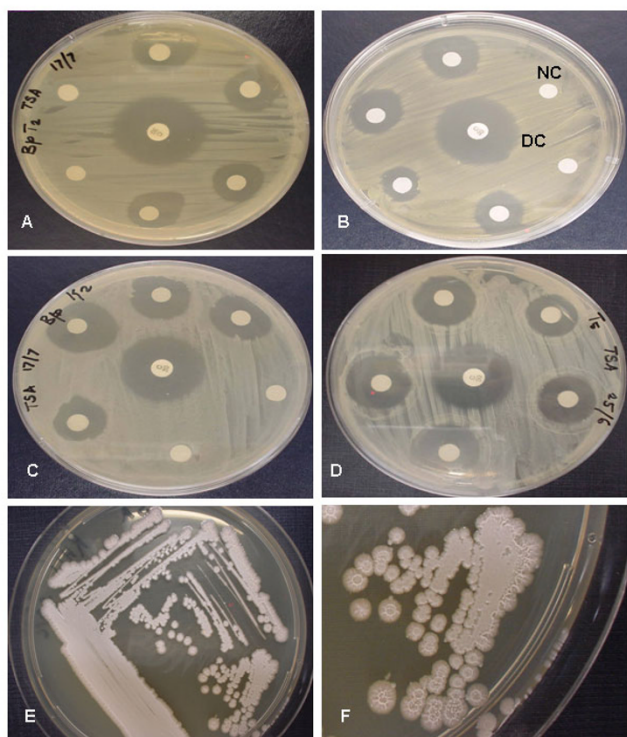


Figure 1

Antimicrobial activities of crude venoms (each disc contained 20 μ l of 100 μ g/ml) of different snake species tested against gram-negative *Burkholderia pseudomallei*. Following 24 h incubation at 37°C, zone of inhibition given by each venom was compared with that of the standard drug chloramphenicol (30 μ g/disc). Inhibition zones against *Burkholderia pseudomallei* given by venoms of: (A) *Daboia russelli russelli*, (B) *Agkistrodon halys*, (C) *Crotalus adamanteus*, (D) *Bitis gaboniarhinoceros*. Rough wrinkled morphological features of gram-negative *Burkholderia pseudomallei* bacteria were grown on Tryptic Soy agar plates at 37°C. (E) bacilli after 36 h incubation, (F) prominent bacilli after 72 h incubation.

Protein assay

The protein concentration of the crude venom solutions was determined using the Bio-Rad protein assay reagent [34] and bovine serum albumin as a standard.

Cell Proliferation assay (XTT-based cytotoxicity assay)

The human macrophage cell line (U-937) was purchased from ATCC (Virginia, USA) and cell Proliferation Kit II was from Roche Applied Sciences (Singapore). Sterile Roswell Park Memorial Institute (RPMI) medium, Fetal Bovine Serum (FBS), 1 M Tris-HCl buffer (pH 7.4), and 10 mM HEPES were purchased from National University Medical Institute (NUMI), Singapore. All chemicals were of analytical and cell culture grade. Human macrophage (U-937) cell line was cultured in 72 cm² flasks at a density of 1×10^7 cells/12 mL in RPMI culture medium supple-

mented with 10% fetal bovine serum (FBS), and 1 ml of HEPES. The cell viability was measured using tetrazolium salts (XTT) as described [35]. Briefly, the cells were allowed to adhere to the bottom of the flask for overnight at 37°C in a humidified atmosphere of 5% CO₂ and 95% air. The culture medium was changed three times a week. To analyze the initial events of venom-mediated cell viability, five different venoms were applied to cultured macrophage cell lines with each venom tested at different concentrations (0.05 – 10 mg/mL) and varied time intervals (12, 24, and 48 h). Cell proliferation was spectrophotometrically quantified using an ELISA plate reader at 490 nm. All assays were prepared in triplicates and repeated thrice. The cytotoxicity of purified PLA₂ enzymes was also tested at different concentrations (0.05–10 μ g/mL) using XTT assay.

Statistical analysis

The results (mean \pm S.D, n = 5) were analysed by one way-ANOVA with repeated measures to analyze factors influencing the zone size of growth inhibition, and for comparison with standard drugs.

Results and discussion

In the present study, both the strains of *B. pseudomallei* (KHW & TES) proved to be highly or intermediately susceptible to various snake venoms at the tested concentration (Table 1). The venom from crotalid (*C. adamanteus*) species showed the most potent antibacterial activity and exhibited larger zone of inhibition on *B. pseudomallei* (Fig. 1) than the venoms of a viperid, *D. russelli russelli* and an Australian elapid, *Pseudechis australis*. Crotalid venoms have previously been reported to have broad activity against aerobic gram-positive and negative bacteria [36], but this is the first report showing the potent antibacterial property of *C. adamanteus* venom against the drug resistant *B. pseudomallei*. The antibacterial effects of crotalid, viperid and elapid venoms established in the present study may likely be due to the cyto-toxins (direct lytic factors) and phospholipase A₂ enzymes contained in those venoms [37]. Association between L-amino acid oxidase activity (LAAO1 and LAAO2) and antibacterial property of *Pseudechis australis* venoms has previously been suggested [19]. However, according to our results, the 5 different venoms of *C. adamanteus*, *Daboia russelli russelli*, *P. australis*, *P. guttata* and *A. halys* exhibited stronger antimicrobial activity against both strains of *B. pseudomallei* than that shown by the L-amino acid oxidase enzymes of *C. adamanteus* and *B. atrox* venoms, thus suggesting that LAAO activity alone may not be solely responsible for the antibacterial activity of these venoms.

In the present study, we have therefore tested a variety of phospholipase A₂ enzymes purified from different snake venoms against *Burkholderia pseudomallei*, and have found

Table 1: Antibacterial properties of animal venoms tested against *Burkholderia pseudomallei* at 0.1 mg/mL concentration.

Common name	Scientific name	Phospholipase A ₂ enzymatic activity		Yield of protein (mg/mL)	Micro-organisms	
		Total activity (μmoles/min)	Specific activity (μmoles/min/mg protein)		<i>B. pseudomallei</i> (KHW)	<i>B. pseudomallei</i> (TES)
Elapidae						
Death adder	<i>Acanthophis augtra</i>	3.74	74.5 ± 0.5	0.1	-	-
Common death adder	<i>Acanthophis antarcticus</i>	190	487.5 ± 0.96	0.78	16.6 ± 0.43	14.3 ± 0.34
Northern death adder	<i>Acanthophis praelongus</i>	241.2	1416 ± 3.84	0.34	8.90 ± 0.23	8.47 ± 0.21
Desert death adder	<i>Acanthophis pyrrhus</i>	138	1150 ± 0.98	0.24	14.7 ± 0.23	16.5 ± 0.32
Hector	<i>Androctonus australis</i>	22.2	85.2 ± 0.27	0.52	-	8.13 ± 0.14
Malayan krait	<i>Bungarus candidus</i>	34.9	166.3 ± 0.56	0.42	-	-
Sea snake	<i>Hydrophis cyanocinctus</i>	3.3	40.5 ± 0.56	0.16	-	-
Indian cobra	<i>Naja naja naja</i>	293.4	1333 ± 1.7	0.44	12.2 ± 0.16	10.2 ± 0.09
Kreft's tiger snake	<i>Notechis aterater</i>	41.96	183.1 ± 0.59	0.46	-	-
Spitting cobra	<i>Naja sumatrana</i>	406.5	903.5 ± 0.6	0.90	-	-
Cobra	<i>Naja kaouthia</i>	228.4	1904 ± 3.8	0.24	8.17 ± 0.15	-
King brown snake	<i>Pseudechis australis</i>	434.5	3949 ± 3.2	0.22	27.7 ± 0.13	29.8 ± 0.105
Speckled brown snake	<i>Pseudechis guttata</i>	308.5	791 ± 2.0	0.78	25.4 ± 0.19	26.8 ± 0.109
Red-bellied black snake	<i>Pseudechis porphyriacus</i>	726.7	3303 ± 1.7	0.44	16.8 ± 0.15	14.2 ± 0.17
Collett's snake	<i>Pseudechis colletti</i>	15.7	111.8 ± 0.89	0.28	-	-
-	<i>Pseudonaja inframaggula</i>	1995	5945 ± 26.6	0.66	7.7 ± 0.16	8.32 ± 0.11
Western brown snake	<i>Pseudonaja nuchalis</i>	162.8	361.8 ± 0.6	0.90	8.2 ± 0.08	9.30 ± 0.11
Eastern brown snake	<i>Pseudonaja textilis</i>	416.8	832.3 ± 0.6	1	14.2 ± 0.09	8.10 ± 0.14
Dugite	<i>Pseudechis affinis</i>	218.7	376.8 ± 0.9	1.16	-	-
Tiger keelback	<i>Rhabdophis tigrinus</i>	36.3	259.1 ± 0.42	0.28	-	-
The coastal taipan	<i>Oxyuranus scutellatus</i>	1275.7	6075 ± 0.98	0.63	-	-
Viperidae						
Pallas	<i>Agkistrodon halys</i>	86.5	157.4 ± 0.20	1.1	20.4 ± 0.14	26.4 ± 0.08
Puff adder	<i>Bitis arietans</i>	124.1	248.2 ± 0.27	0.54	18.2 ± 0.16	16.2 ± 0.17
West African gaboon viper	<i>Bitis gabonica rhinoceros</i>	126.5	452.4 ± 0.57	0.56	16.0 ± 0.19	14.5 ± 0.26
Common lancehead snake	<i>Bothrops atrox</i> (L-amino acid oxidase)	3.8	6.36 ± 0.06	1.2	24.4 ± 0.19	26.2 ± 0.19
South American rattlesnake	<i>Crotalus adamanteus</i> (L-amino acid oxidase)	34.2	201.3 ± 0.21	0.34	25.4 ± 0.208	23.4 ± 0.18
South American rattlesnake	<i>Crotalus adamanteus</i>	236.4	619.4 ± 0.46	1.56	30.0 ± 0.23	28.3 ± 0.23
Saw-scaled viper	<i>Echis carinatus</i>	53.4	106.5 ± 0.49	1.4	8.0 ± 0.14	7.5 ± 0.15
Indian Russell's viper	<i>Daboia russelli russelli</i>	392.8	785.2 ± 0.40	1.36	29.9 ± 0.12	28.6 ± 0.16
Burmese Russell's viper	<i>Daboia russelli siamensis</i>	262.4	524.4 ± 0.44	1.24	16.2 ± 0.15	15.6 ± 0.18
Wagler's pit viper	<i>Trimeresurus wagleri</i>	4.6	38.2 ± 0.26	0.24	15.1 ± 0.14	16.3 ± 0.14
Apiidae						
Honeybee venom	<i>Apis mellifera</i>	3.7	20.5 ± 0.1	0.36	7.22 ± 0.23	12.3 ± 0.21
Scorpionidae						
Black scorpion	<i>Androctonus crasicuda</i>	3.6	69.5 ± 0.3	0.104	-	-
Scorpion	<i>Buthotus hottentota</i>	4.4	10.4 ± 0.2	0.86	-	-
Scorpion	<i>Buthotus hottentota hottentota</i>	5.1	38.8 ± 0.1	0.26	-	-
Chinese red scorpion	<i>Buthus martensii</i> Karsch.	4.6	90.4 ± 0.2	0.102	-	-
Ceftazidime (CF)	30 μg/disc	-	-	-	28.9 ± 0.56	31.7 ± 0.78
Chloramphenicol (C)	30 μg/disc	-	-	-	28.5 ± 0.37	30.0 ± 0.12
Streptomycin (S)	10 μg/disc	-	-	-	27.3 ± 0.25	26.4 ± 0.29

Values of Phospholipase A₂ enzymatic activity (μmoles/min/mg protein) are presented as mean ± S.D. (n = 10) of ten replicates.

*The values given as mean ± S.D. (n = 5) represent a venom inhibition zone in mm, including the 7 mm diameter of the disc, after 24 h incubation. The bacterial inoculum per plate contained 3.2×10^8 colony forming units (cfu/mL) which were spread onto the TS agar surface with sterile cotton swap. Sterile paper discs (7 mm diameter) were placed onto the TS agar surface and 20 μL of venom solution (0.1 mg/mL) added. Micro-organisms: *B.p* (KHW)-*Burkholderia pseudomallei* (KHW) and *B.p* (TES) *Burkholderia pseudomallei* (TES), Control (0); No activity (-), Drug controls: Chloramphenicol (30 μg/disc); Ceftazidime (30 μg/disc); Streptomycin (10 μg/disc).

Table 2: In vitro antibacterial activity of purified phospholipase A₂ enzymes from snake venoms.

Phospholipase A ₂ enzymes (PLA ₂ s)	Snake species (Scientific name)	Mol. wt.(kDa)	Conc. (mg/mL)	Micro-organisms	
				<i>B. pseudomallei</i> (Strain KHW)	<i>B. pseudomallei</i> (Strain TES)
Crotoxin A (CA)	<i>Crotalus durissus terrificus</i>	23.5	0.5 mg/mL	-	-
Crotoxin B (CB)	<i>Crotalus durissus terrificus</i>	23.5	0.5 mg/mL	24.8 ± 0.089	27.6 ± 0.133
Ammodytoxin A	<i>Vipera ammodytes ammodytes</i>	13.8	0.5 mg/mL	-	-
Mojave toxin	<i>Crotalus scutulatus scutulatus</i>	23.5	0.5 mg/mL	-	-
β-Bungarotoxin	<i>Bungarus multicinctus</i>	20.5	0.5 mg/mL	-	-
Taipoxin	<i>Oxyuranus scutellatus scutellatus</i>	45.6	0.5 mg/mL	-	-
Mulgatoxin	<i>Pseudechis australis</i>	13.2	0.5 mg/mL	20.5 ± 0.075	22.7 ± 0.117
Daboiatoxin (DbTx)	<i>Daboia russelli siamensis</i>	15.0	0.5 mg/mL	24.8 ± 0.103	26.2 ± 0.121
Bee venom PLA ₂	<i>Apis mellifera</i>	19.0	0.5 µg/mL	18.3 ± 0.089	20.4 ± 0.075

*The values given represent a venom inhibition zone in mm, including the 7 mm diameter of the disc, after 24 h incubation. The bacterial inoculum per plate contained 3.2×10^8 cfu/mL forming units which were spread onto the TS agar surface with sterile cotton swap. Sterile paper discs (7 mm diameter) were placed onto the TS agar surface and 20 µL of enzymes (0.5 mg/ml concentration) added, Control (0); No activity (-).

that crotoxin B and daboiatoxin display the strongest antibacterial activity against both the strains of *B. pseudomallei* (KHW & TES) (Table 2). Crotoxin B is a basic neurotoxic phospholipase A₂ (*Crotalus durissus terrificus*) containing three chain protein that enhances the lethal potency of crotoxin [38], while daboiatoxin is a monomeric PLA₂ (*Daboia russelli siamensis*) with strong neurotoxic and myotoxic activities [30]. Moreover, mulgatoxin (myotoxic PLA₂ of *P. australis*) and bee (*Apis mellifera*) venom PLA₂ have also been found to exert significant antibacterial activity against both strains (KHW & TES) of *B. pseudomallei*.

The PLA₂ enzymes showing potent inhibitory activity were further studied for minimum inhibitory concentrations (MICs) (Fig. 2, 3). Among those examined, the two purified PLA₂ enzymes (crotoxin B and daboiatoxin) showed stronger inhibitory activity at a lower dilution (MICs 0.25 mg/mL) against *B. pseudomallei* (TES) than melittin and mulgatoxin. The two isolates (TES and KHW) of *B. pseudomallei* showed various levels of sensitivity to the two PLA₂ toxins with MICs ranging between 0.5 and 0.03125 mg/ml. The susceptibility of crotoxin B (69%) and daboiatoxin (63%) against TES was more or less equal to that of chloramphenicol and ceftazidime against TES (Table 3). The antibiotics chloramphenicol (70%) and ceftazidime (80%) were highly susceptible against *B. pseudomallei* TES & KHW, but MIC breaking points of antibiotics were recorded at the lowest dilution (MICs 0.125 mg/mL) than the PLA₂ toxins. Our results corroborate with the previous report on the conventional four-drug regimen which has been replaced by ceftazidime for acute treatment. However, the chloramphenicol has also been given for the first 8 weeks of oral treatment [39]. In the present study, the antibiotic activity was gradually declined at the dilu-

tion 0.03125 mg/mL against *B. pseudomallei* (KHW) and (TES). The rate of resistance of ceftazidime and chloramphenicol against KHW was 20% and 30%, respectively. In a randomised trial previously reported, 161 severe melioidosis patients were treated with ceftazidime (120 mg/kg) + chloramphenicol (100 mg/kg) + doxycycline (4 mg/kg) + TMP/SMX for less than 7 days. The mortality was 37% for ceftazidime and 74% for chloramphenicol [40]. However, in our study, melittin was slightly more active than mulgatoxin against *B. pseudomallei* (strain KHW) at MIC 0.25 mg/mL. It showed very weak MICs (Fig. 2, 3) when compared with chloramphenicol and ceftazidime. Other PLA₂s (ammodytoxin A, Mojave toxin, β-bungarotoxin and taipoxin) however, lacked any activity against both isolates of *B. pseudomallei* at all tested dilutions (0.5 – 0.03125 mg/mL). Based on previous studies using cationic antibacterial peptides, cathelicidin-derived peptides had modest MIC against *S. maltophilia* and *A. xylosoxidans* (1.0 to > 32 mg/mL), but none inhibited *Burkholderia cepacia* [41]. Moreover, another cationic peptide (hBD-3) that was proven highly or intermediately effective (MBCs >100 µg/mL) against 23 tested strains did not show any effect against *Burkholderia cepacia* at 50 µg/mL [42]. When the MIC of peptide D2A21 was compared with that of the tracheal antimicrobial peptide (TAP), the former peptide displayed greater potency than TAP against *P. aeruginosa* at 0.125 – 4 mg/mL, *S. aureus* at 0.25 – 4 mg/mL, and *Burkholderia cepacia* at 32 to >64 mg/mL, respectively [43]. Relative to the MIC of these cationic antimicrobial peptides, crotoxin B appears to have a much lower dose (MIC 0.25 mg/mL) against *B. pseudomallei*. MIC value of PLA₂s are more or less equally comparable to that of ceftazidime (MICs 0.125 mg/mL), a drug of choice for melioidosis infections in humans. The rat model of lung infection (14 days) showed that the *B. cenocepacia* LPS inner core oli-

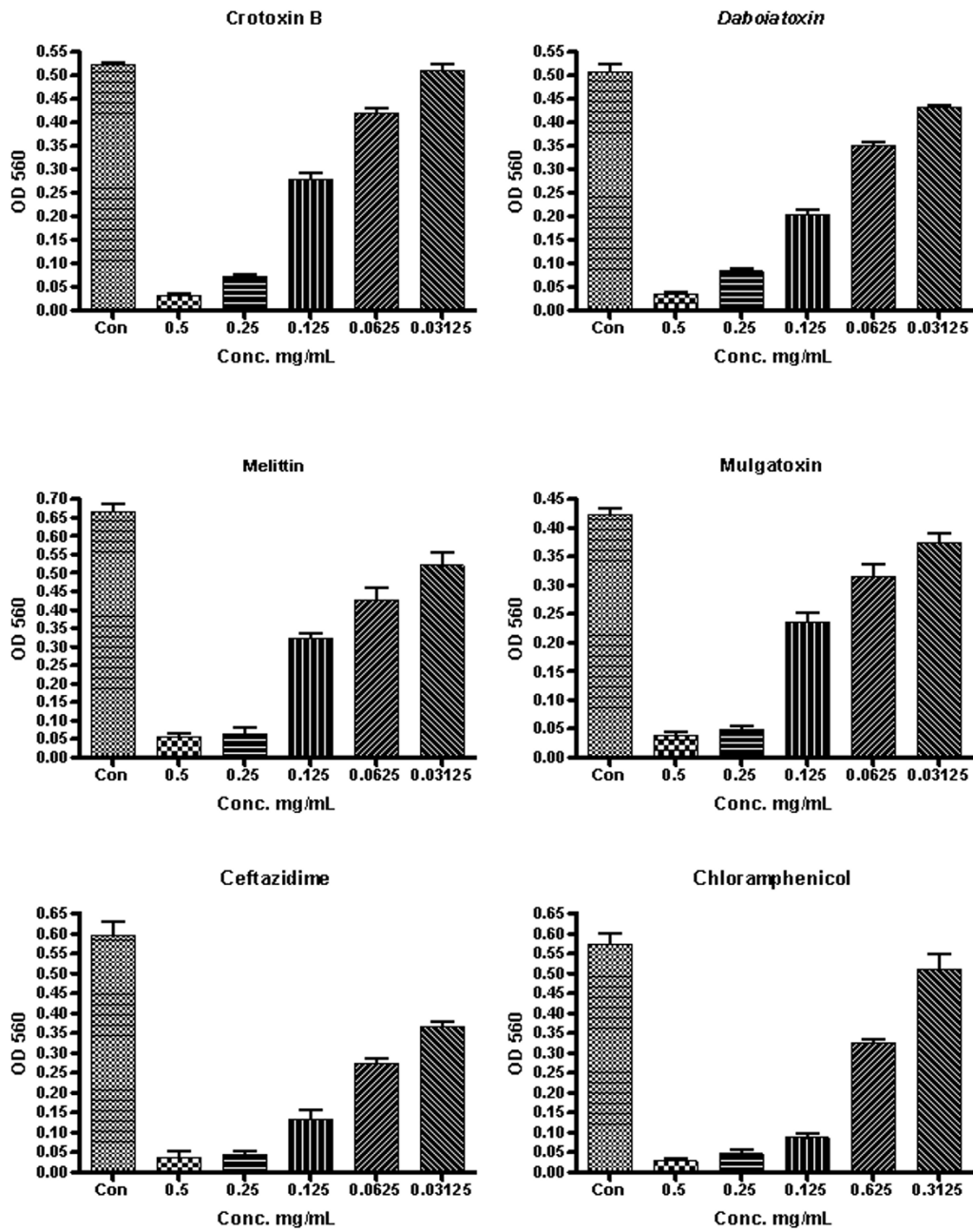


Figure 2

Phospholipase A₂ activity against pathogen from patients with KHW. Micro-dilution technique was used to test the MICs of PLA₂s as compared to that of the antibiotics. The values are the optical density read at 560 nm (means ± S.D.) from a single experiment performed in triplicates.

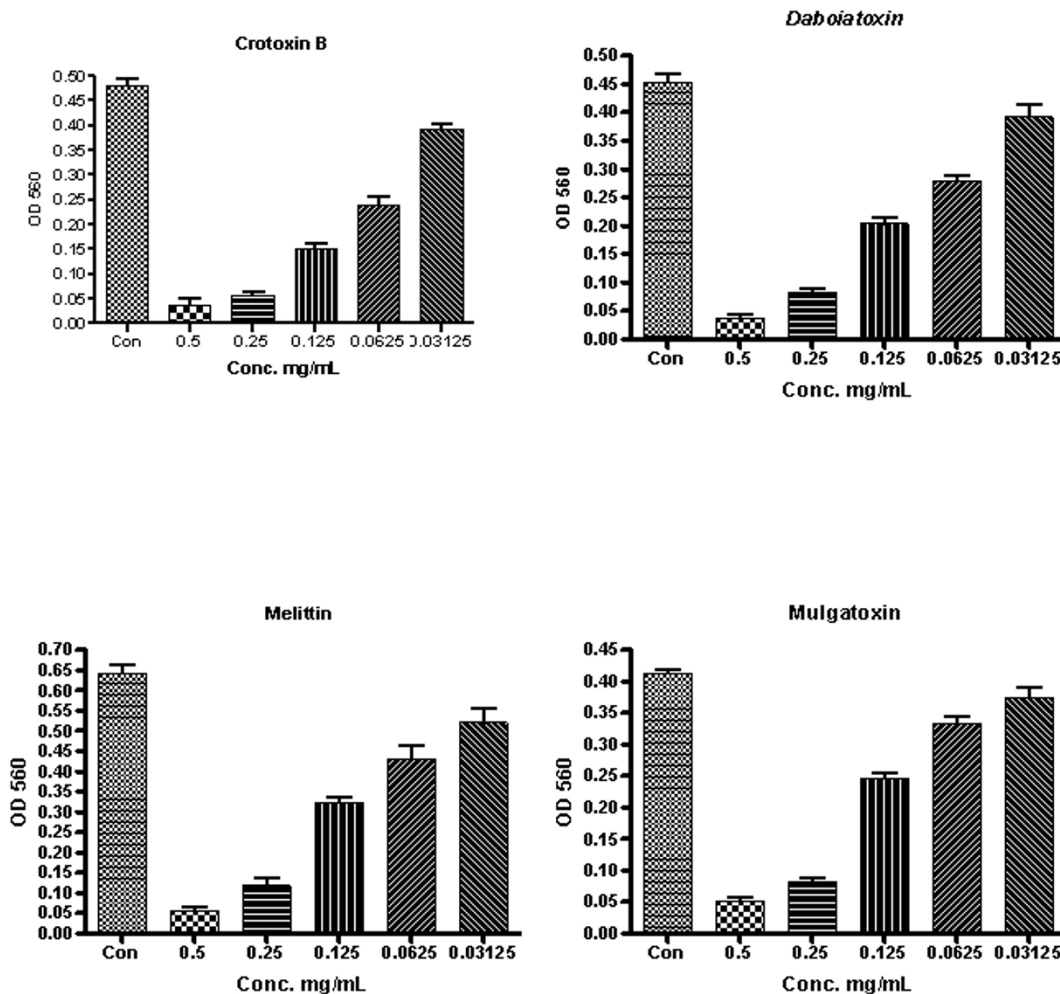


Figure 3
Phospholipase A₂ activity against pathogen from patients with TES. Micro-dilution technique was used to test the MICs of PLA₂s as compared to that of the antibiotics. The values are the optical density read at 560 nm (means ± S.D.) from a single experiment performed in triplicates.

gosaccharide is needed for resistance to three structurally unrelated antimicrobial peptides for in vivo survival in a rat model of chronic lung infection [44].

When PLA₂ activity was examined, the highest activity was found in the Australian elapid venoms (*Oxyuranus scutellatus*>*Pseudonaja inframagula*) followed by *Pseudechis australis*, *Pseudechis porphyriacus*, *Naja kaouthia*, *Naja naja naja*, *Acanthophis praelongus* and *Acanthophis pyrrhus* respectively. In contrast, the remaining venoms of the Apiidae and Scorpionidae showed relatively less phos-

pholipase A₂ activity than the viperidae venoms (Table 1).

The survival bar resulting from the XTT assay shows that all five venoms (*C. adamanteus*, *B. gabonica*, *P. australis*, *D. russelli russelli* and *A. halsy*) do not have cytotoxic effects on the proliferation of cells up to 0.5 mg/mL concentration (Fig. 4a–e). However, the higher concentrations (1, 5 and 10 mg/mL) showed severe morphological changes of the cell lines such as membrane disruption and significant cell lyses when compared with control and the positive

Table 3: MIC breakpoints for ceftazidime and chloramphenicol when compared to that of purified PLA₂s enzymes.

Phospholipase A ₂ enzymes (PLA ₂ s)	MIC mg/mL	<i>B. pseudomallei</i> (strain KHW)					MIC mg/mL	<i>B. pseudomallei</i> (strain TES)				
		Ctrl	0.5	0.25	0.125	0.0625		0.03125	Ctrl	0.5	0.25*	0.125 ^a
Crotoxin B (CB)	0.65	0.012 (64%)	0.04 (61%)	0.33 (32%)	0.43 (32%)	0.56 (9%)	0.73	0.05 (73%)	0.06 (69%)	0.33 (40%)	0.43 (30%)	0.52 (21%)
Daboiatoxin (DbTx)	0.65	0.26 (39%)	0.09 (56%)	0.37 (28%)	0.49 (16%)	0.55 (10%)	0.68	0.03 (65%)	0.05 (63%)	0.24 (20%)	0.36 (8%)	0.43 (1%)
Bee venom PLA ₂	0.48	0.038 (38%)	0.056 (42%)	0.15 (33%)	0.24 (24%)	0.39 (9%)	0.44	0.03 (45%)	0.06 (38%)	0.27 (17%)	0.36 (8%)	0.41 (3%)
Mulgatoxin	0.46	0.07 (39%)	0.083 (37%)	0.21 (25%)	0.28 (18%)	0.39 (7%)	0.43	0.04 (40%)	0.07 (37%)	0.26 (18%)	0.32 (12%)	0.37 (7%)
Chloramphenicol	0.67	0.01 (68%)	0.03 (64%)	0.07 (60%)	0.42 (10%)	0.51 (1%)	0.77	0.026 (74%)	0.04 (73%)	0.07 (70%)	0.34 (41%)	0.37 (38%)
Ceftazidime	0.76	0.08 (58%)	0.05 (61%)	0.04 (62%)	0.24 (24%)	0.44 (22%)	0.89	0.01 (88%)	0.03 (86%)	0.09 (80%)	0.35 (47%)	0.49 (33%)

* MIC values are given as mean of five replicates (n = 5), the bacterial inoculums per plate contained 3.2×10^8 cfu/mL forming units/well, Control (bacterial inoculums); ^aThe ceftazidime breaking points (MICs 0.125 mg/mL) against TES, *The PLA₂ toxin breaking points (MICs 0.250 mg/mL) against both the strains of *B. pseudomallei* after 24 h incubation.

control (Fig. 5). In contrast, the purified PLA₂s, crotoxin B and daboiatoxin, did not change the viability (Fig. 4f–g) of cells up to 0.05 – 10 µg/mL concentrations as compared to the control. The cell viability and morphology of macrophage cells were shown (Fig. 6) after exposure to crotoxin B in a dose- and time-dependent manner.

A comparison of the N-terminal sequences of crotoxin B with other snake venom PLA₂s shows (Fig. 7) that most amino acid residues are conserved in Ca²⁺ binding and catalytic network regions. The C-terminal segment of crotoxin B, on the other hand, shows a modest difference in the amino acid residues as compared with the C-terminal sequences of other venom PLA₂s. Moreover, comparison of the hydrophobic profiles of different snake venom PLA₂s reveals that the C-terminal segment of crotoxin B is relatively more hydrophobic than those of other PLA₂s (Fig. 8). This cationic hydrophobic nature of crotoxin B phospholipase A₂ enzyme may most likely be responsible for the strong antimicrobial action seen on *B. pseudomallei*.

Conclusion

The present study indicates that the purified PLA₂ enzymes (crotoxin B, daboiatoxin) possess strong antibacterial activity against *B. pseudomallei*. The fact that viperidae (*Crotalus adamanteus*, *Daboia russelli russelli*) and elapidae (*Pseudechis australis*) venoms display more potency than other venoms may be due to the PLA₂ enzymes present. However, this antibacterial profile of snake venoms reported herein will be useful in the search

for potential antibacterial agents against drug resistant microorganisms like *B. pseudomallei*.

Abbreviations

LAAO – L-amino acid oxidase

PLA₂s – Phospholipase A₂ enzymes

MICs – Minimum Inhibitory Concentrations

XTT – Tetrazolium salts

KHW, TES – different isolates of *Burkholderia pseudomallei*

RPMI – Roswell Park Memorial Institute (RPMI) medium

FBS – Fetal Bovine Serum

Competing interests

The author(s) declare that they have no competing interests.

Authors' contributions

RPS – Study design, performance of the entire parameters, data analysis and manuscript drafting.

AP – Cell proliferation study using XTT based cytotoxic assay.

PG – Supervision, laboratory support, critical analysis and manuscript edition.

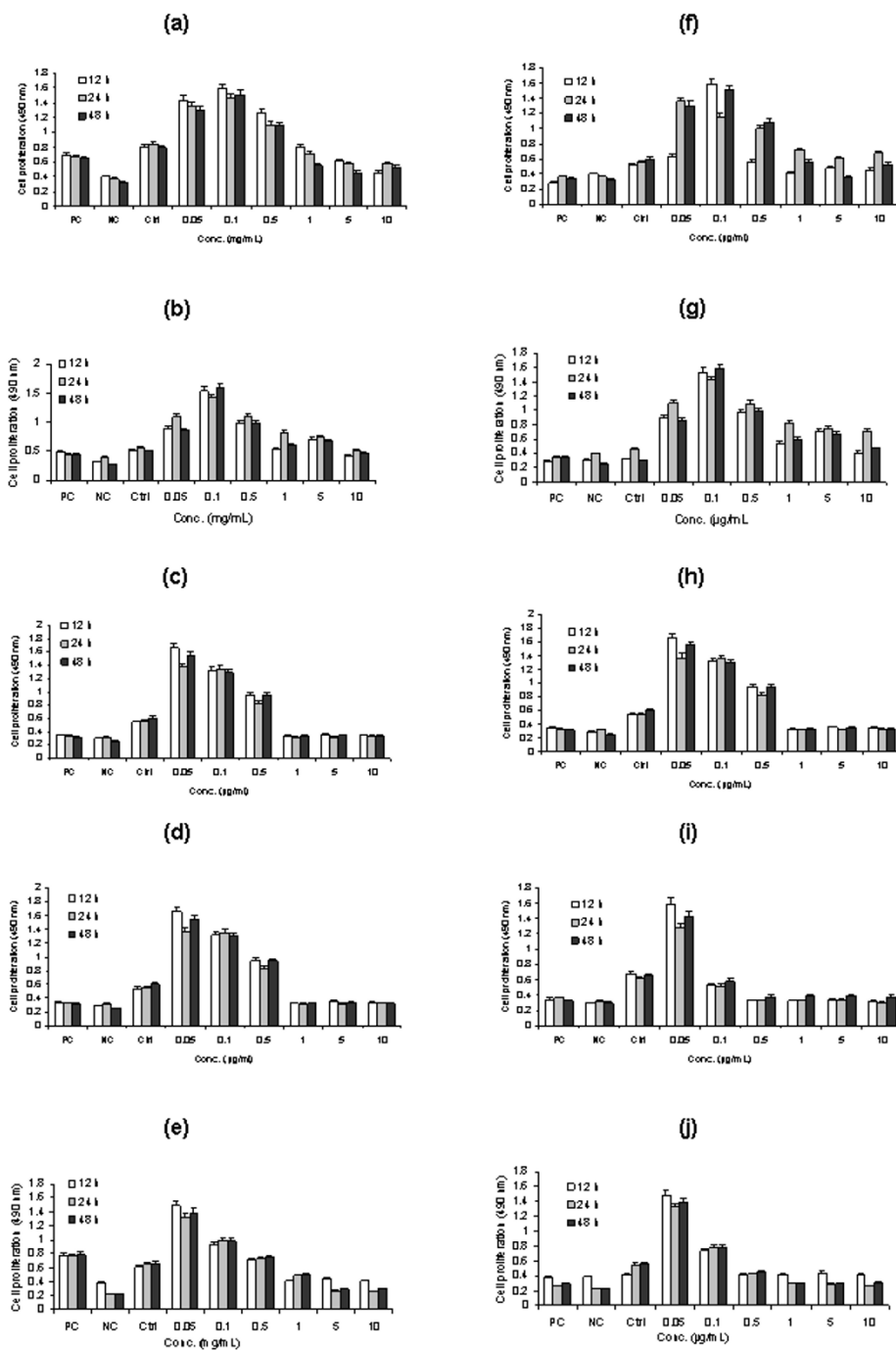
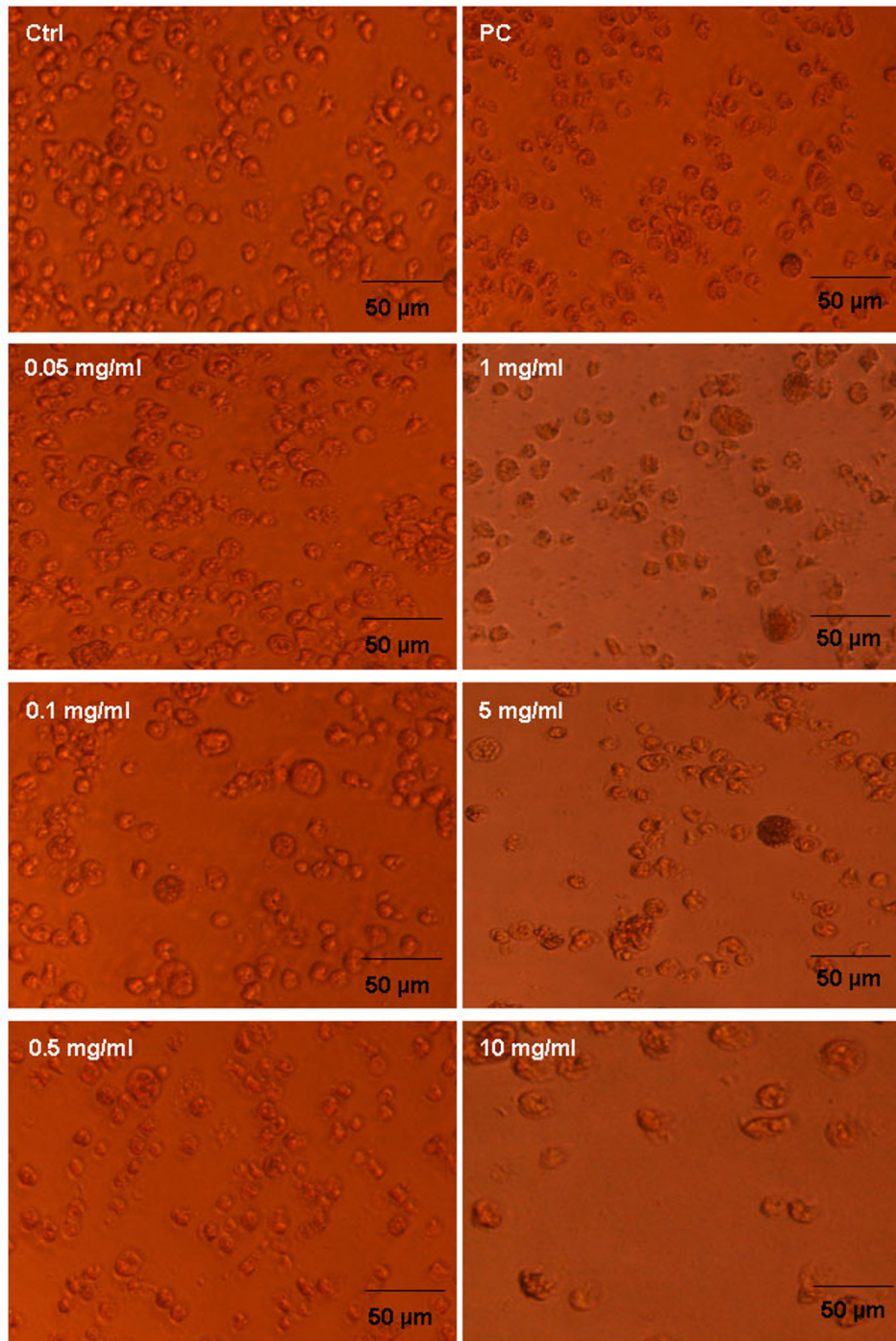


Figure 4

Cell proliferation (U-937 Human macrophage cell line) was determined by XTT assay to evaluate the cytotoxic effect of different venoms (a – e) and PLA₂s (f – j) exposed at different time intervals. (a) South American rattlesnake (*Crotalus adamanteus*), (b) Russells viper (*Daboia russelli russelli*), (c) King brown (*Pseudechis australis*), (d) Pallas (*Agkistrodon halys*) (e) Speckled brown (*Pseudechis guttata*) (f) Crotoxin B (*Crotalus durissus terrificus*), (g) Daboiatoxin (*Daboia russelli russelli*), (h) Melittin (*Apis mellifera*), (i) Mulgatoxin (*Pseudechis australis*) (j) Crotoxin A (*Crotalus durissus terrificus*). Macrophages were incubated with varying concentrations of venoms (0.05 – 10 mg/ml) and PLA₂s (0.05 – 10 µg/ml).

**Figure 5**

Morphological changes of U-937 Human macrophage cell line after exposure to *Crotalus adamanteus* venom at different concentrations. (Ctrl) macrophage supplemented with medium without any treatment served as control; (PC), cells exposed with ceftazidime as a positive control. Macrophages were incubated with venom at different (0.05–10 mg/mL) concentrations (Other photos not shown).

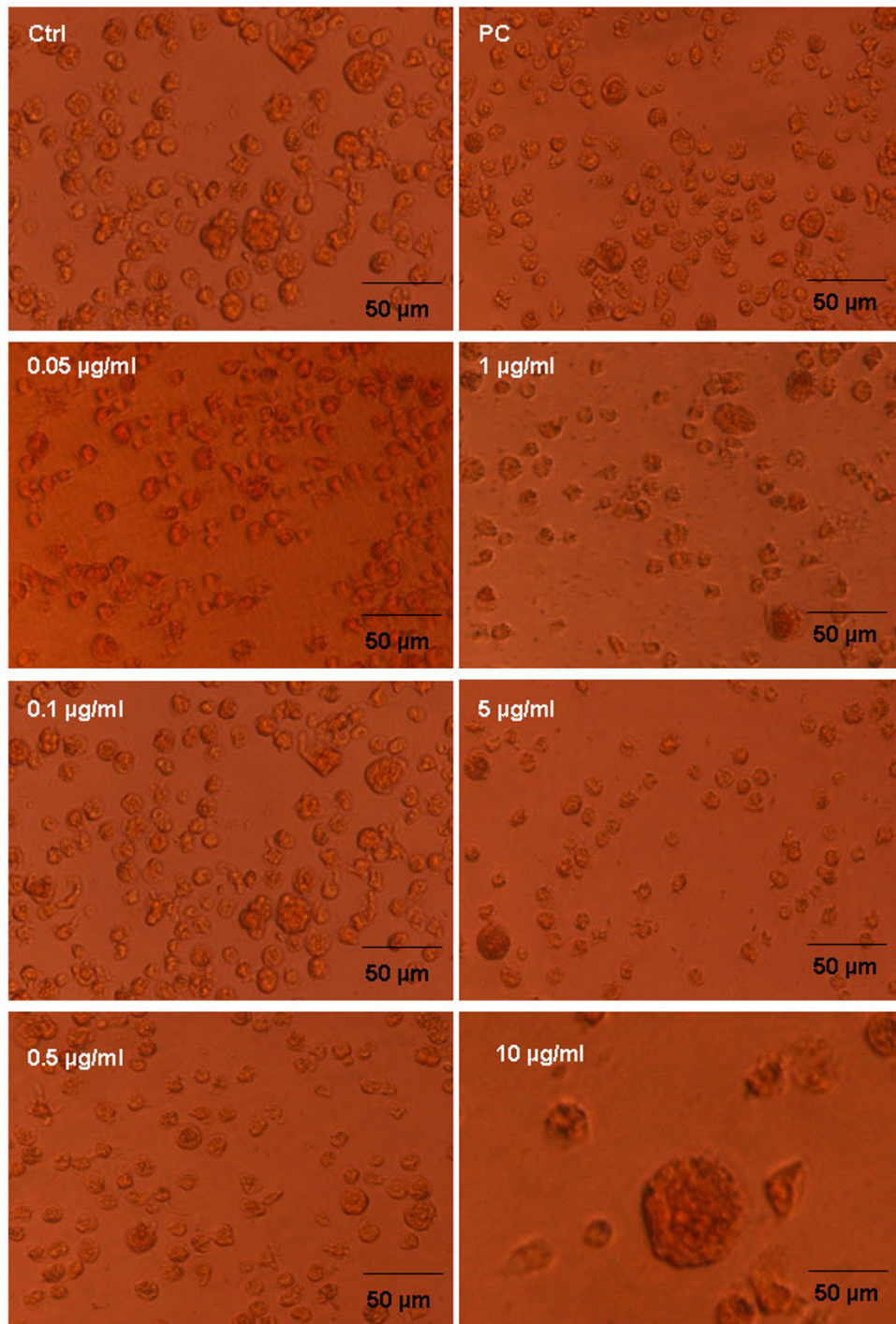


Figure 6
Morphological changes of U-937 Human macrophage cell line after exposure to PLA₂s at varying concentrations (0.05–10 µg/mL). (Ctrl), control macrophages supplemented with medium without any treatment; (PC), cells exposed with ceftazidime as a positive control.

Accession No	sPLA ₂ s	Amino acid sequence					Residues
		1	10	20	30	40	
P62022	CB1	HLLQFNKHKIKFETRKN	-RIPFYAFYGCYCGWGGGRPKD	ATDRCCFVHDCCYGKLA	---		55
P62023	Mtx-b	HLLQFNKHKIKFETRKN	-RIPFYAFYGCYCGWGGGRPKD	ATDRCCFVHDCCYGKLA	---		55
P24027	CB2	SLLQFNKHKIKFETRKN	-RVPFYAFYGCYCGWGGGRPKD	ATDRCCFVHDCCYGKLA	---		55
P14421	ATX	NLLQFNKHKIKETGKN	-RIPFYAFYGCYCGGGGQKPKD	GTDRCCFVHDCCYGRLV	---		55
P59071	VRV-VIIIa	-LLEFGKMLEETGKL	-RIPSYSYGCYCGWGGKGTPKD	ATDRCCFVHDCCYGNLP	---		54
P20474	Myot I	-LIEFAKMLEETKRL	-RPFYTYTYGCYCGWGGGQKPKD	ATDRCCFVHDCCYGKLS	---		54
P00626	Ammody A	SLLFPGHMLGETGKN	-PLTSYSYGCYCGWGGKGTPKD	ATDRCCFVHDCCYGNLP	---		55
P81458	RVV-VD	NLQFPAEMIVKMTGKN	-PLSSYSYGCYCGWGGKGTQD	ATDRCCFVHDCCYEKVK	---		55
Q02471	RV-4 pre	NLQFARMINGKLGAF	-SVMWYISYGCYCGWGGQKPKD	ATDRCCFVHDCCYGKLV	---		55
P24605	Myot II	-LLEFGKMLEETGKN	-PAKSYGAYGCNCGULGRGKPKD	ATDRCCYVHKCCYKGLT	---		54
Q90249	BthTX-I	-LLEFGKMLEETGKN	-PAKSHGAYGCNCGULGRGKPKD	ATDRCCYVHKCCYKGLT	---		54
P48650	ECHCA	-VVELGKMIQETGKS	-RPFSTYSYGCYCGGGGERGPLD	ATDRCCCLAHSCCYDTLP	---		53
P08878	CA	SLVEFETLMMKIAGRS	-GISYYSYGCYCGAGGQKPKD	ASDRCCFEHDCCYAKLT	---		55
Q90251	β-bunga	NLYQFKEMIRYIP	CEKTWGEVADYGCYCGAGGS	GTIDALDRCCYVHDCCYGDAAKIR			59
P00614	Taipoxin	NLLQFGRMIRCAMR	SRPVMWYMDYGCYCGKGGSGT	PVDDLRCCQVHDCCYGEAVRRF			59
		:: :: ::	:	*** **	** * ****	* **	

sPLA ₂ s	Amino acid sequence						Residues	Homology
	60	70	80	90	100	110		
CB1	KCNTKWDIYPYSLKSGYITCG	-KGTWCEEQICECDRVAAE	CLRRSLSTYKYGYHFPD	SRCRGPSETC			122	100*
Mtx-b	KCNTKWDIYPYSLKSGYITCG	-KGTWCEEQICECDRVAAE	CLRRSLSTYKYGYHFPD	SRCRGPSETC			122	90*
CB2	KCNTKWDIYRYSLKSGYITCG	-KGTWCEEQICECDRVAAE	CLRRSLSTYKNEYHFPD	SRCRGPSETC			122	84*
ATX	KCNTKSDIYRYSLKSGYITCG	-KGTWCEEQICECDRVAAE	CFRRNLDTYNGYHFPD	SKCTETSEEC			122	72*
VRV-VIIIa	DCNPKSDRYKRYKRVNGAIVCG	-KGTSCENRICECDKAAAIC	FRQNLMTYSKCYMLY	PDFLCKG----			116	55*
Myot I	NCKPKTDRYSYSRKSGVVICG	-EGTPEKQICECDKAAAIC	FRNLRTYKRYMAYPDL	LCKKPAEK			121	55*
Ammody A	DCSPKTDRYKRYHRENGAIVCG	-KGTSCENRICECDRVAAE	CFRNLKTYNYIYRNY	PDFLCKESEK			121	53*
RVV-VD	SCKPKLSLYSYFQNGGIVCG	-DNHSCRAVCECDRVAAT	CFRDLNLTYSKRYHFP	PSQCTG-TEQC			122	45*
RV-4 pre	GCPKLAISYSYFQNGGIVCG	-RNNGCLRTICECDRVAANC	FRQNLMTYKRYKFL	SSSKCRQRSEQC			122	45*
Myot II	GCPKLDRYSYSWKDKTIVCG	-ENNSCLKELICECDKAAV	ICLRENLTYNKRYRYL	KPLCK-----			121	44*
BthTX-I	GCDPKLDRYSYSWKDKTIVCG	-ENNPCLKELICECDKAAV	ICLRENLTYNKRYRYL	KPLCK-----			121	41*
ECHCA	DCSPKTDRYKRYKRENGEIVCG	-NSTSCKRCICECDKAAV	CLRNLNTYNKRYTY	YVNFVCKGDIEK			122	41*
CA	GCDPTDVTYRQEDGEIVCG	-EDDPGCTQICECDKAAAIC	FRNSMDTYKYLQF	SPENCQGESQPC			122	39*
β-bunga	GCDPKTQSYSYKLTQRTIVCG	-AAGTCARVWCD	CDRTAALCFGNS	-----			103	37*
Taipoxin	GCAPYWTLYSWKCYGKAPT	CN-TKTRCQRVCRCD	AKAAECFARS	-----			103	34*
	* . * :	* . * :	* . * :	* . * :	* . * :	* . * :		

Figure 7
 Comparison of the amino acid sequences of Crotoxin basic chain I, CBI [*Crotalus durissus terrificus*] phospholipase A₂ enzymes (Phosphatidylcholine 2-acylhydrolase) with other PLA₂s of Mojave toxin basic chain, Mtx-b [*Crotalus scutulatus scutulatus*], Crotoxin basic chain 2, CB2 [*Crotalus durissus terrificus*], Agkistrotoxin, ATX [*Agkistrodon halys*], VRV-PL-VIIIa [*Daboia russelli pulchella*], BOTAS (Myotoxin I) *Bothrops asper* (Terciopelo), ATXA, ammodytin A precursor [*Vipera ammodytes ammodytes*]; RVV-VD [*Daboia russelli russelli*]; RV-4 precursor [*Daboia russelli siamensis*], BOTAS (Myotoxin II) [*Bothrops asper*], BOTJR (BthTX-I) [*Bothrops jararacussu*], ECHCA (Ecarpholin S) [*Echis carinatus* (Saw-scaled viper)], Crotoxin acid chain precursor (CA) [*Crotalus durissus terrificus*], β-bungarotoxin A6 chain precursor [*Bungarus multicinctus* (Many-banded krait)] and OXYSC taipoxin alpha chain [*Oxyuranus scutellatus scutellatus*] and completely conserved residues in all sequences are bolded and marked by asterisks. The gaps are inserted in the sequences in order to attain maximum homology.

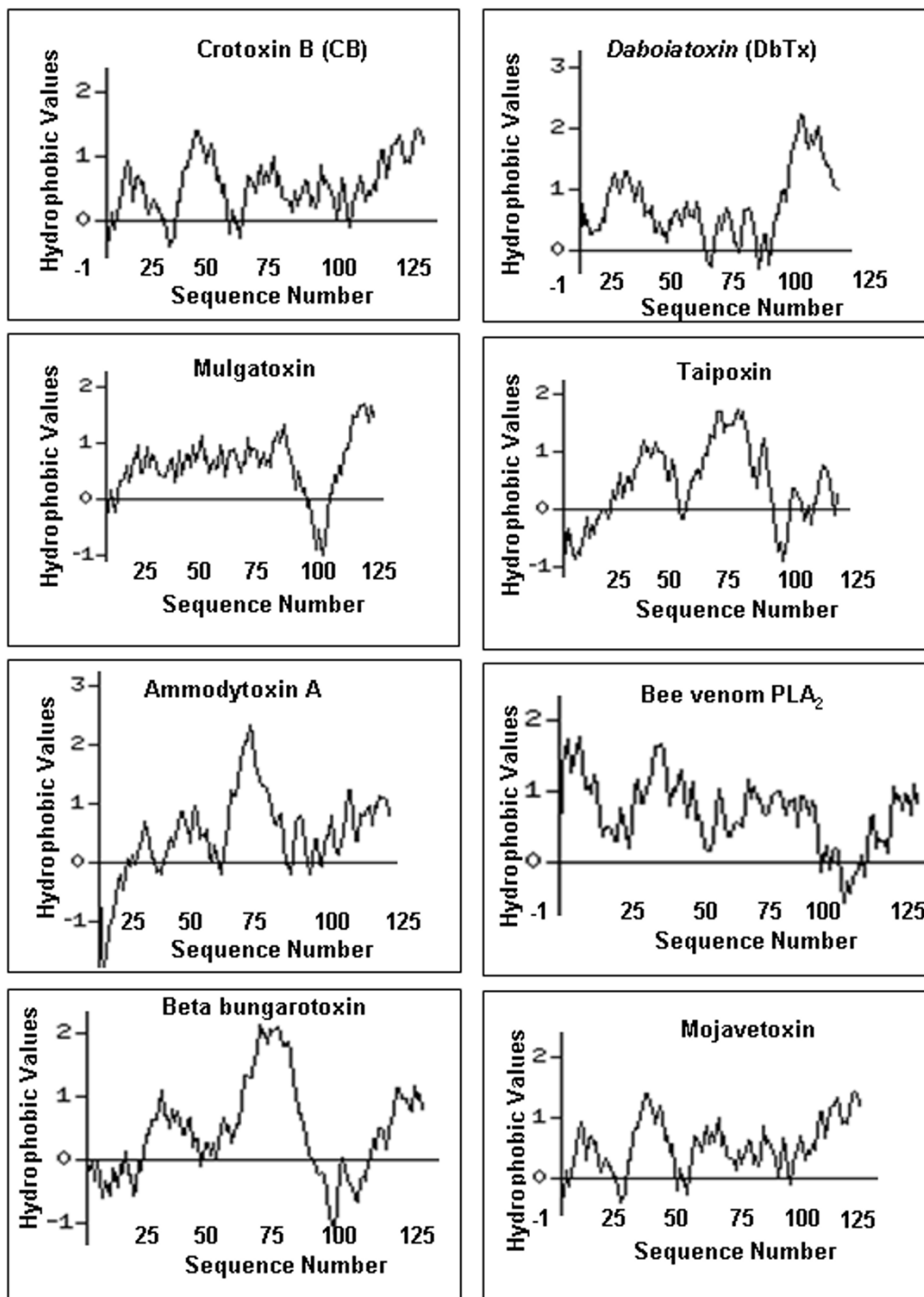


Figure 8
 Hydrophobic profiles of phospholipase A₂ enzyme such as crotoxin b, daboia toxin, mulgatoxin, taipoxin, ammodytoxin A, bee venom PLA₂, beta-bungarotoxin and mojavetoxin were calculated by using the kyte-doolittle method.

MMT – Manuscript evaluation and critical analysis.

YHH, VTKC, HB – Co supervision of the antimicrobial susceptibility testing, provision of bacterial cultures and laboratory support.

JTW – Assisted in technical support for antimicrobial testing and culturing of bacteria.

Acknowledgements

This work was supported by the Research Grant from the Defence Science and Technology Agency (DSTA), Singapore (Grant No R -181 000 063 422). We thank the Department of Microbiology, Yong Loo Lin School of Medicine, National University of Singapore, for their support in Lab facility and the gift of bacterial cultures used in this investigation.

References

1. Chaowagul W, White NJ, Dance DAB, Wattanagoon Y, Naigowit P, Davis TME, Looareesuwan S, Pitakwatchara N: **Melioidosis: a major cause of community acquired septicemia in north-eastern Thailand.** *J Infect Dis* 1989, **159**:890-899.
2. Yabuuchi E, Arakawa M: **Burkholderia pseudomallei and melioidosis: be aware in temperate area.** *Microbiol Immunol* 1993, **37**:823-836.
3. Puthuchery SD, Parasakthi N, Lee MK: **Septicaemic melioidosis: a review of 50 cases from Malaysia.** *Trans Roy Soc Trop Med Hyg* 1992, **86**:683-685.
4. White NJ: **Melioidosis.** *Lancet* 2003, **361**:1715-1722.
5. Currie BJ, Fisher DA, Howard DM: **Endemic melioidosis in tropical northern Australia a 10 year perspective study and review of the literature.** *Clin Infect Dis* 2000, **31**:981-986.
6. Hsueh PR, Teng LJ, Lee LN: **Melioidosis: an emerging infection in Taiwan?** *Emerging Infect Dis* 2001, **7**:428-433.
7. Yang S: **Melioidosis research in China.** *Acta Tropica* 2000, **77**:157-165.
8. Chaowagul W, Suputtamongkol Y, Dance DAB, Rajchanuvong A, Pattara-arechai J, White NJ: **Relapse in melioidosis: incidence and risk factors.** *J Infect Dis* 1993, **168**:1181-1185.
9. Simpson AJH, Smith MD, Weverling GJ: **Prognostic value of cytokine concentrations (tumor necrosis factor, interleukin 6 and interleukin 10) and clinical parameter in severe melioidosis.** *J Infect Dis* 2000, **181**:621-625.
10. Nuntayanuwat S, Dharakul T, Chaowagul W, Songsivilai S: **Polymorphism in the promoter region of tumor necrosis factor-alpha gene is associated with severe melioidosis.** *Hum Immunol* 1999, **60**:979-983.
11. Bonomo RA: **Multiple antibiotic-resistant bacteria in long-term-care facilities: an emerging problem in the practice of infectious diseases.** *Clin Infect Dis* 2000, **31**:1414-1422.
12. Dance DAB: **Melioidosis.** In *Manson's tropical diseases* 20th edition. Edited by: Cook GC. London, England: W B. Saunders Co. Ltd; 1996:925-930.
13. Leelarasamee A: **Burkholderia pseudomallei: the unbeatable foe?** *Southeast Asian J Trop Med Public Health* 1998, **29**:410-415.
14. Eickhoff TC, Bennett JV, Hayes PS, Feeley J: **Pseudomonas pseudomallei: susceptibility to chemotherapeutic agents.** *J Infect Dis* 1970, **121**:95-102.
15. Jones AL, Beveridge TJ, Woods DE: **Intracellular survival of Burkholderia pseudomallei.** *Infect Immun* 1996, **64**:782-790.
16. Ganz T, Lehrer RI: **Antimicrobial peptides of leukocytes.** *Current Opinion in Haematology* 1997, **4**:53-58.
17. Paramo L, Lomonte B, Pizarro-Cerda J, Bengoechea JA, Gorvel JP, Moreno E: **Bactericidal activity of Lys49 and Asp49 myotoxic phospholipase A₂ from Bothrops asper snake venoms-synthetic Lys49 myotoxin II- (115-129)-peptide identifies its bactericidal region.** *Euro J Biochem* 1998, **253**:452-461.
18. Veridiana Rodrigues M, Silvana Marcussi J, Rafael Cambráira S, Ana de Araújo L, Natael Malta-Neto R, Amélia Hamaguchi, Eloísa Ferro AV, Maria Homs-Brandeburgo I, José Giglio R, Andreimar Soares M: **Bactericidal and neurotoxic activities of two myotoxic phospholipases A₂ from Bothrops neuwiedi pauloensis snake venom.** *Toxicon* 2004, **44**:305-314.
19. Stiles BG, Sexton FW, Weinstein SA: **Antibacterial effects of different snake venoms: purification and characterisation of antibacterial proteins from Pseudechis australis (Australian king brown or Mulga snake) venom.** *Toxicon* 1991, **29**:1129-1141.
20. Corzo G, Escoubas P, Villegas E, Barnham KJ, Weilan HE, Norton RS, Nakajima T: **Characterization of unique amphipathic antibacterial peptides from venom of the scorpion Pandinus imperator.** *J Biochem* 2001, **359**:35-45.
21. Possani LD, Zurita M, Delepierre M, Hernandez FH, Rodriguez MH: **From noxiustoxin to Shiva-3, a peptide toxic to the sporogonic development of Plasmodium berghei.** *Toxicon* 1998, **36**:1683-1692.
22. Budnik BA, Olsen JV, Egorov TA, Anisimova VE, Galkina TG, Musolyamov AK, Grishin EV, Zubarev RA: **De novo sequencing of antimicrobial peptides isolated from the venom glands of the wolf spider Lycosa Singoriensis.** *J Mass Spectrometry* 2004, **39**:193-201.
23. Haeblerli S, Kuhn-Nentwig L, Schaller J, Nentwig W: **Characterisation of antibacterial activity of peptides isolated from the venom of the spider Cupiennius salei (Araneae: Ctenidae).** *Toxicon* 2000, **38**:373-380.
24. Fennel JE, Shipman WH, Cole LJ: **Antibacterial action of melittin, a polypeptide from the venom.** *Proc Soc Exp Biol Med* 1968, **127**:707-710.
25. Stocker JF, Traynor JR: **The action of various venoms on Escherichia coli.** *J Applied Bacteriol* 1986, **61**:383-88.
26. Hancock REW: **Peptide antibiotics.** *Lancet* 1997, **349**:418-422.
27. Hancock RE, Scott MG: **The role of antimicrobial peptides in animal defenses.** *Proc Natl Acad Sci USA* 2000, **97**:8856-8861.
28. Silphaduang U, Noga EJ: **Antimicrobials-peptide antibiotics in mast cells of fish.** *Nature* 2001, **414**:268-269.
29. National Committee for Clinical Laboratory Standards: **Performance Standards for Antimicrobial Disk Susceptibility Tests: Approved Standard. M2-A6. National Committee for Clinical Standards, Wayne Pennsylvania.** 1997.
30. Thwin MM, Gopalakrishnakone P, Yuen R, Tan CH: **A major lethal factor of the venom of Burmese Reussell's viper (Daboia russelli siamensis): Isolation, N-terminal sequencing and biological activities of daboitoxin.** *Toxicon* 1995, **33**:63-76.
31. National Committee for Clinical Laboratory Standards: **Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria that Grow Aerobically. Approved Standard M7-A4. National Committee for Clinical Standards, Wayne Pennsylvania.** 1997.
32. Amsterdam D: **Susceptibility testing of antimicrobials in liquid media.** In *Antibiotics in laboratory medicine* 4th edition. Edited by: Lorian V. Baltimore, MD: Williams and Wilkins; 1996:52-111.
33. Reynolds LJ, Hughes LL, Dennis EA: **Analysis of human synovial fluid phospholipase A₂ on short chain phosphatidylcholine-mixed micelles: Development of a spectrophotometric assay suitable for a microtiterplate reader.** *Ann Biochem* 1992, **204**:190-197.
34. Bradford MM: **A rapid and sensitive method for quantification of microgram quantities of protein utilizing the principle of protein dye binding.** *Ann Biochem* 1976, **72**:248-254.
35. Pachiappan A, Thwin MM, Manikandan J, Gopalakrishnakone P: **Glial inflammation and neurodegeneration induced by candoxin, a novel neurotoxin from Bungarus candidus venom: global gene expression analysis using microarray.** *Toxicon* 2005, **46**:883-899.
36. Talan DA, Citron DM, Overturf GD, Singer B, Froman P, Goldstein EJ: **Antibacterial activity of crotalid venoms against oral snake flora and other clinical bacteria.** *J Infect Dis* 1991, **164**:195-198.
37. Blaylock RSM: **Antibacterial properties of KwaZulu Natal snake venoms.** *Toxicon* 2000, **38**:1529-1534.
38. Oliveira DG, Toyama MH, Novello JC, Beriam LOS, Marangoni S: **Structural and functional characterization of basic PLA₂ isolated from Crotalus durissus terrificus venom.** *J Protein Chem* 2002, **21**:161-168.
39. Rajchanuvong A, Chaowagul W, Suputtamongkol Y, Smith MD, Dance DAB, White NJ: **A perspective comparison of co-amoxiclav and the combination of chloramphenicol, doxycycline and**

- co-trimoxazole for the oral maintenance treatment of melioidosis.** *Br J Clin Pharmacology* 2000, **49**:184-191.
40. White NJ, Dance DA, Chaowagul W, Wattanagoon Y, Wuthiekanun V, Pitakwatchara N: **Halving of mortality of severe melioidosis by ceftazidime.** *Lancet* 1989, **2**:697-701.
 41. Bouchier C, Boulain JC, Bon C, Menez A: **Analysis of cDNAs encoding the two subunits of crotoxin, a phospholipase A₂ neurotoxin from rattlesnake venom: the acidic non enzymatic subunit derives from a phospholipase A₂-like precursor.** *Biochem Biophys Acta* 1991, **1088**:401-408.
 42. Saiman L, Tabibi S, Starner TD, San Gabriel P, Winokur PL, Jia HP, McCray PB Jr, Tack BF: **Cathelicidin peptides inhibit multiply antibiotic-resistant pathogens from patients with cystic fibrosis.** *Antimicrob Agents Chemother* 2001, **45**:2838-2844.
 43. Sahly H, Schubert S, Harder J, Rautenberg P, Ullmann U, Schröder J, Podschun R: **Burkholderia is highly resistant to human Beta-defensin 3.** *Antimicrob Agents Chemother* 2003, **47**:1739-1741.
 44. Loutet SA, Flannagan RS, Kooi C, Sokol PA, Valvano I MA: **A complete lipopolysaccharide inner core oligosaccharide is required for resistance of Burkholderia cenocepacia to antimicrobial peptides and bacterial survival in vivo.** *J Bacteriol* 2006, **18**:2073-2080.

Pre-publication history

The pre-publication history for this paper can be accessed here:

<http://www.biomedcentral.com/1471-2334/6/100/prepub>

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:

http://www.biomedcentral.com/info/publishing_adv.asp

