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# Endotoxin tolerance and cross-tolerance in mast cells involves TLR4, TLR2 and FcεR1 interactions and SOCS expression: perspectives on immunomodulation in infectious and allergic diseases

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## Abstract

**Background:** The study of the endotoxin tolerance phenomenon in light of the recently defined roles of mast cells and toll-like receptors as essential components of the innate immune response and as orchestrators of acquired immunity may reveal potentially useful mechanisms of immunomodulation of infectious and allergic inflammatory responses, such as sepsis or asthma. Here we evaluated the phenomenon of direct tolerance of endotoxins, as well as the induction of cross-tolerance and synergism by stimulation with toll-like receptor-2 (TLR2) and FcεR1 agonists, in murine mast cells prestimulated with lipopolysaccharide (LPS). Additionally, we evaluated some stimulatory and inhibitory signaling molecules potentially involved in these phenomena.

**Methods:** MC/9 cells and primary bone marrow-derived mast cells obtained from C57BL/6 and TLR4<sup>-/-</sup> knock-out mice were sensitized to DNP-HSA (antigen) by incubation with DNP-IgE and were prestimulated with LPS for 18 hr prior to stimulation. Cultures were stimulated with LPS or Pam3Cys-Ser-(Lys)<sub>4</sub> 3HCl (P3C), a TLR2 agonist, individually or in combination with antigen. The production of IL-6 and TNFα, the phosphorylation of NFκB and p38 MAPK, and the expression of TLR4 and SOCS-1 and -3 were analyzed.

**Results:** We found that production of TNFα and IL-6 in murine mast cells that have been pretreated with LPS and challenged with TLR4 (LPS) or -2 (P3C) agonists was reduced, phenomena described as endotoxin tolerance (LPS) and cross-tolerance (P3C), respectively. The expression of TLR4 was not affected by LPS pretreatment. Our results show that the FcεR1 agonist DNP-HSA (antigen) interacts synergistically with LPS or P3C to markedly enhance production of cytokines (TNFα and IL-6). This synergistic effect with LPS and P3C was also attenuated by LPS pretreatment and was mediated by TLR4. These results may be attributed to the reduction in phosphorylation of the mitogen-activated protein kinase (MAPK), p38, and the transcription factor NFκB, as well as to an increase in the expression of the suppressors of cytokine signaling (SOCS)-1 and -3 proteins in LPS-pretreated mast cells.

**Conclusions:** These findings can be explored with respect to the modulation of inflammatory responses associated with infectious and allergic processes in future studies.

## Background

Currently, treatment of sepsis is ineffective and few therapeutic innovations have been developed to improve

it [1,2]. Since 1972, stemming from the quasi-intuitive ideas of Thomas Lewis [3], the concept that our immune response to infection is the sustaining foundation for sepsis has been widely accepted. An increased understanding of sepsis pathophysiology has provided stronger support for this hypothesis [4]. During the second half of the 1990 s, two coinciding facts led to new

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perspectives on a different approach to sepsis. First, the discovery of toll-like receptors (TLR) [5,6] initiated our present comprehension of how the host recognizes pathogen molecular patterns, how the innate response is initiated, and how the acquired immune response is organized [7]. Second, in 1996, Bernd Echtenacher et al. determined the essential role of mast cells in the innate immune response using a model of peritoneal sepsis in mast cell-deficient mice [8]. Additional work from other groups (reviewed in 9) elevated these cells, previously considered mere effectors, to the category of sentinels of the innate immune system and organizers of the adaptive immune response. In fact, their permanent location in sites likely to suffer invasion by pathogens - skin, paranasal sinuses, lungs, and intestinal mucosa - places them in a privileged position in terms of detection and subsequent organization of the immune response. Innumerable possibilities for modulation of the inflammatory response appeared after a better understanding of the role played by TLRs was gained, having as targets their stimulatory and inhibitory signaling pathways [10]. The accumulation of evidence defining mast cells as fundamental in the immune response to sepsis [11-13] opens up new perspectives on the mechanisms of immunomodulation by this type of cell [14].

Interaction between responses to infectious and allergic stimuli has been suggested on several levels, raising the possibility of a common pathway. Genetics-based studies have revealed the association of polymorphisms in the myosin light chain kinase (MYLK) gene with increased risk of sepsis and acute pulmonary injury. This same gene is involved in other inflammatory pathologies, including bronchial asthma [15]. Epidemiological data have demonstrated the effect of exposure to TLR agonists on the incidence of allergic phenomena [16]. Experimental studies in human mast cell cultures have shown an interaction between Fc $\epsilon$ R1 receptors and TLR2 [17]. In bone marrow derived-mast cells (BMMC), we have previously demonstrated the synergic action of co-stimulation of Fc $\epsilon$ R1 and TLR in the production of inflammatory cytokines [18].

The endotoxin tolerance phenomenon was described more than 60 years ago [19], and it is characterized by hyporesponsiveness to endotoxin exposure, induced by prior exposure. Cross-tolerance, which is defined by tolerance of a different stimulus induced by endotoxin, was described later. In the context of TLR signaling, the tolerance phenomenon can be used as a tool for identification of signaling molecules that can attenuate the inflammatory response, revealing potential participants in immunomodulation. Mast cells are some of the first cells to have contact with invading pathogens; when activated, they release immunoregulatory cytokines that organize the inflammatory response. The release of

TNF $\alpha$  and the recruitment of circulating leukocytes are essential elements of the immune response that are attributed to mast cells, and characteristically, mast cells are the only type of cell that can store pre-formed TNF $\alpha$  and release it when activated [20]. Few studies have addressed endotoxin tolerance in mast cells, likely due to the fact that its primary role in response to infection has been defined so recently. In other cell types, where endotoxin tolerance have been more thoroughly evaluated, the proteins suppressors of cytokine signaling (SOCS) are involved in tolerance phenomena as negative regulators of the pro-inflammatory response, via the TLR4-NF $\kappa$ B pathway [21].

We tested the hypothesis that mast cells display the phenomenon of direct tolerance to endotoxins after prestimulation of BMMC with LPS, and that this could induce cross-tolerance for stimulation with Fc $\epsilon$ R1 and TLR2 agonists. The release of TNF $\alpha$  and IL-6 was measured, and the same experiment was conducted in BMMC obtained from TLR4<sup>-/-</sup> knock-out (KO) mice to determine the role of TLR4 in the induction of cross-tolerance and in the potentiation of this response by co-stimulation. Additionally, we evaluated the effects of LPS prestimulation on phosphorylation of the mitogen-activated protein kinase, p38, and the transcription factor NF $\kappa$ B, as well as the expression of the SOCS-1 and -3 proteins, signaling molecules involved in cytokine production.

## Methods

### Materials

Reagents were obtained from the following sources: culture medium and reagents from Invitrogen/GIBCO (Carlsbad, CA); recombinant mouse IL-3 from Pepro Tech (Rocky Hill, NJ); IgE monoclonal anti-DNP antibody and its antigen, dinitrophenylated human serum albumin (DNP-HSA), and highly purified lipopolysaccharide (LPS, *Escherichia coli* 055:B5) from Sigma (St. Louis, MO); Pam3Cys-Ser-(Lys)<sub>4</sub> 3HCl (P3C) from EMC Microcollections GmbH (Tuebingen, Germany); and polyclonal antibodies that detect phospho-p38 MAPkinase (Thr180/Tyr182), phospho-NF $\kappa$ B (Ser536), and the proteins themselves from Cell Signaling Technology (Danvers, MA).

### Mast cell culture

Bone-marrow derived mast cells (BMMC) were cultivated in suspension in 75 cubic milliliter flasks (300,000 cell/ml) in RPMI-1640 supplemented with 5% fetal bovine serum, 30 ng/ml of IL-3, 2 mM glutamine, 100  $\mu$ M non-essential amino acids, 10  $\mu$ M 2-mercaptoethanol, and 1 mM sodium pyruvate. The MC/9 mast cell line was cultured in the same culture medium without IL-3. BMMC were obtained from wild-type (C57BL/6)

and TLR4<sup>-/-</sup> KO mice, under approval of the Ethics in Animal Experimentation Committee [Comitê de Ética em Experimentação Animal] of the Federal University of Minas Gerais (CETEA-UFMG), using previously described procedures [22], and the cells were cultivated in a CO<sub>2</sub> incubator at 37 degrees Celsius. BMMC were utilized in experiments after 4 weeks in culture and showed 99% mast cells when stained with toluidine blue.

#### Experimental protocols

BMMC were sensitized to DNP-HSA (antigen) with 100 ng/ml DNP-IgE and were pretreated with LPS (10 ng/ml or LPS 100 ng/ml) for 18 hours prior to stimulation. After 18 hours, cells were washed 3 times with cell culture medium, resuspended in complete medium and stimulation was performed with antigen (20 ng/ml), LPS (1 µg/ml), or P3C (1 µg/ml) alone or in combinations (antigen+LPS or antigen+P3C) for 24 hours for analyzing production of cytokines or 30 min for measuring degranulation. For signaling studies, BMMC were washed 3 times with HEPES buffer and stimulated in the same buffer for 30 minutes with antigen (20 ng/ml), LPS (1 µg/ml), or a combination of antigen+LPS.

#### Assay for cytokine measurement

TNFα and IL-6 were measured in the supernatant after 24 hours of stimulation using the enzyme-linked immunosorbent assay (ELISA) method (BioSource kits, Camarillo, CA).

#### Measurement of degranulation

For these measurements, cultures were washed and the medium was substituted with HEPES-buffered saline medium before stimulation. Degranulation was determined by the measurement of β-hexosaminidase marker release by a colorimetric assay in which the release of p-nitro phenol (coming from p-nitro phenyl-N-acetyl-β-D-glucosaminide) is measured. Values are expressed as the percentage of β-hexosaminidase that is released into the medium.

#### Immunoblotting

Cells were lysed in a protease/phosphatase-inhibiting buffer by the addition of 100 µl of this solution to the cell suspension. This buffer consisted of Complete Protease Inhibitor Cocktail (Roche Molecular Biochemicals, Indianapolis, IN), Sigma protease inhibitor cocktail (Sigma, St Louis, MO), 3-4 dichloroisocoumarin (Roche Molecular Biochemicals), and benzamidine, as well as the phosphatase inhibitors sodium orthovanadate, sodium pyrophosphate, and sodium fluoride (Sigma). Samples were boiled for 4 minutes, and any existing debris was removed by centrifugation at 14,000 rpm for 5 minutes prior to loading the gels. Proteins were

separated by NuPAGE BisTris gels (Invitrogen) and then transferred to nitrocellulose membranes for immunoblotting with the primary antibodies indicated. Visualization was by chemiluminescence, and quantification was by densitometry (Kodak Image Station 4000R).

#### Statistical analysis

The results are provided as mean ± SEM and were analyzed using the Student t-test with the confidence interval established at 95%. Values were considered significant if  $p < 0.05$ . Data were stored in the Graphpad Prism 4 statistical program.

#### Results

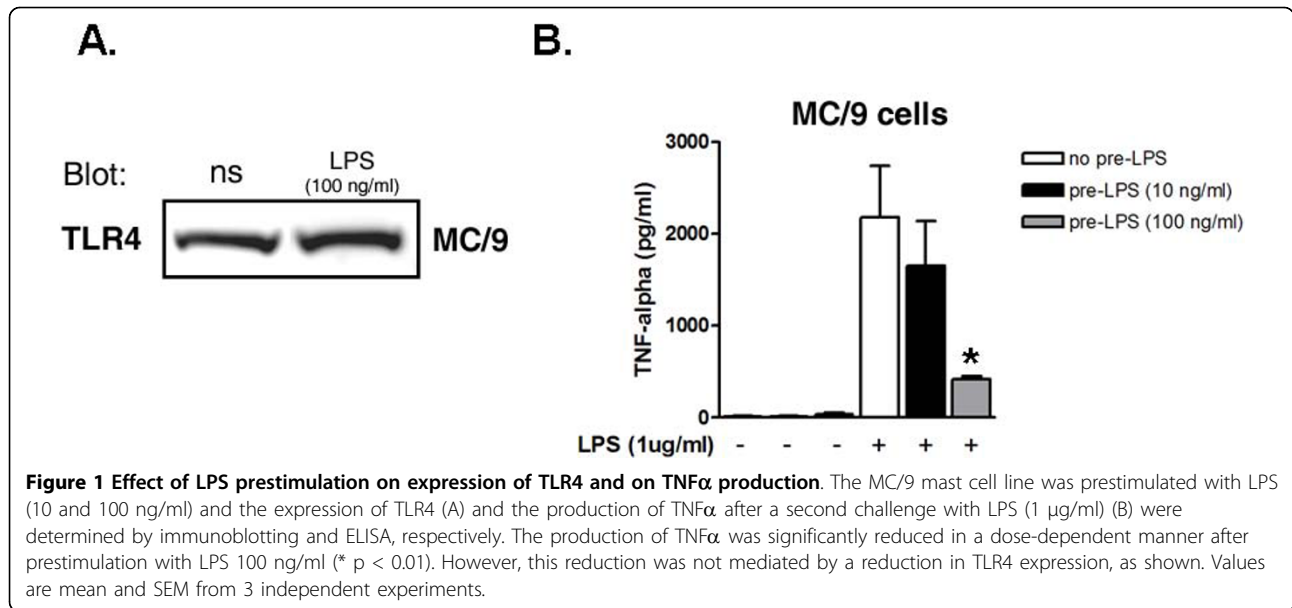
##### **Mast cells express endotoxin tolerance not related to the reduction of toll-like receptor-4 expression**

Pretreatment with 100 ng/ml of LPS for 18 hours did not affect expression of TLR4 in mast cells (Figure 1A). However, mast cells pretreated with LPS (100 ng/ml) showed lower production of TNFα upon a second challenge with LPS (1 µg/ml) compared with cells that not receive prestimulation ( $p < 0.05$ ) (Figure 1B).

##### **LPS acts in synergy with antigens through TLR4 signaling to augment production of cytokines; this potentiation is reduced by preconditioning with LPS**

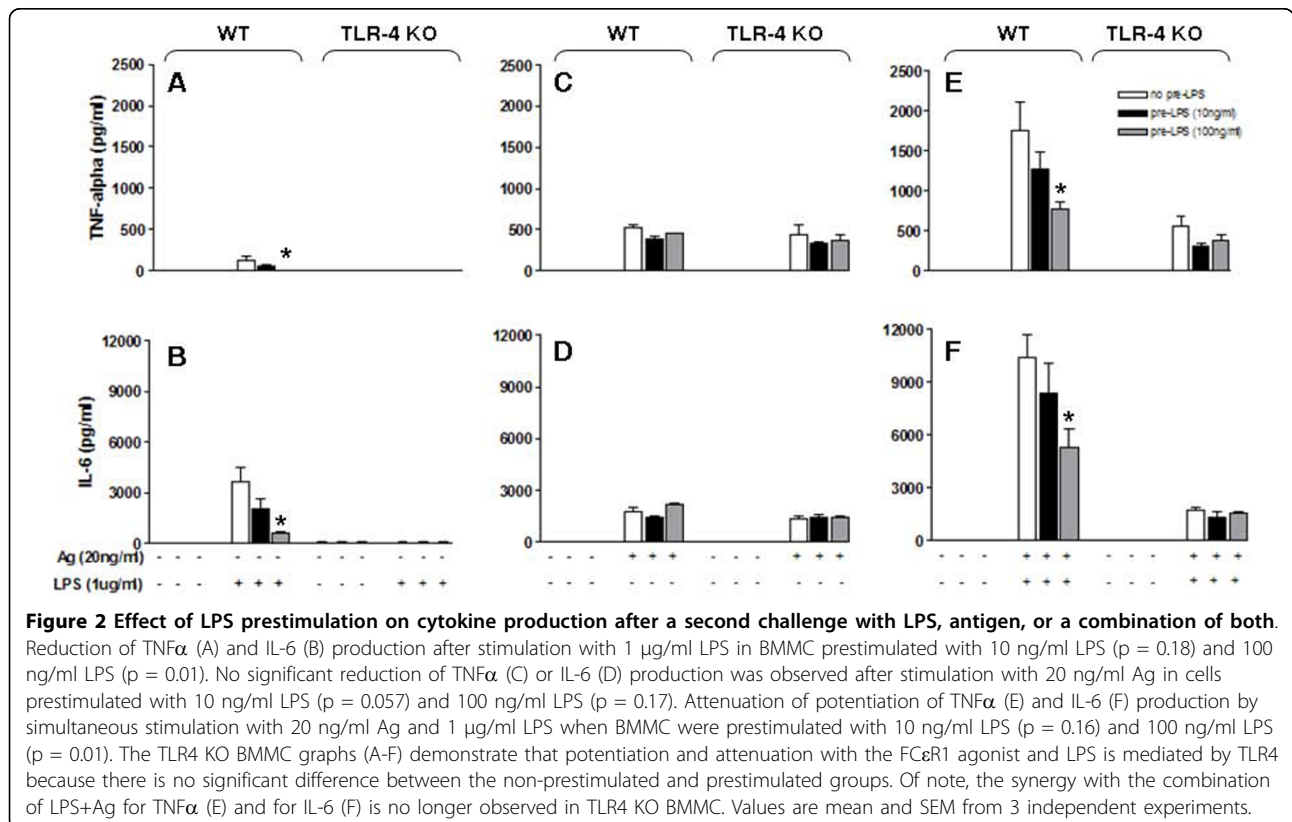
Similar to the results seen with the MC/9 cell line, primary BMMC pretreated with LPS (100 ng/ml) showed lower production of TNFα upon a second challenge with LPS (1 µg/ml), ( $p = 0.01$ ). Prestimulation with LPS (10 ng/ml) was non-significant ( $p = 0.18$ ) (Figure 2A, WT). As for IL-6, the same was observed with lower production upon prestimulation with LPS 100 ng/ml ( $p = 0.01$ ) and non-significant with 10 ng/ml ( $p = 0.17$ ) (Figure 2B, WT). In comparison with BMMC that did not receive prestimulation, cells prestimulated with LPS (100 ng/ml) displayed significantly reduced production of TNFα and IL-6 upon a second challenge with LPS (1 µg/ml). These results illustrate that prior exposure to endotoxins induces a significant reduction in the production of these proinflammatory cytokines upon a second stimulation with LPS (Figures 2A and 2B, WT). In BMMC from TLR4 knockout mice, we did not detect production of TNFα or IL-6 in response to a second challenge with LPS, in cells that were either pretreated or not treated with LPS (Figure 2A and 2B, TLR4 KO).

To test the hypothesis that a common pathway may modulate responses to stimulation of the TLR4 and FcεR1 receptors, BMMC were incubated with DNP-IgE (50 ng/ml), prestimulated with LPS (10 ng/ml or 100 ng/ml) for 18 hours, and then stimulated with antigen (20 ng/ml). Prestimulation with 10 ng/ml LPS ( $p = 0.057$ ) and 100 ng/ml LPS ( $p = 0.17$ ) did not reduce



production of TNF $\alpha$  when the cells were stimulated with antigen (Figure 2C, WT). As for IL-6, there was no reduction with prestimulation with LPS 10 ng/ml ( $p = 0.1$ ) and 100 ng/ml ( $p = 0.1$ ) (Figure 2D, WT). The lack of TLR4 did not affect the production of cytokines in response to antigen (Figures 2C and 2D, TLR4 KO).

As we have previously demonstrated [18], simultaneous stimulation of the Fc $\epsilon$ R1 and TLR4 receptors results in greater production of cytokines (TNF $\alpha$  and IL-6) when compared to the individual stimulation of each. In support of the common pathway hypothesis, we demonstrate two new observations in this sequence of



events. First, we show the attenuation of synergy in TNF $\alpha$  ( $p = 0.01$ ) (Figure 2E) and IL-6 ( $p = 0.03$ ) (Figure 2F) production between LPS and antigen after preconditioning with LPS (100 ng/ml). Second, this synergism is dependent on TLR4, as will be demonstrated further on. Therefore, prestimulation with LPS (100 ng/ml) results in attenuation of the synergistic effect of co-stimulation with LPS and antigen.

To determine the role of TLR4, we conducted experiments using the same protocol of prestimulation with LPS, but stimulated BMMC that do not express TLR4, which were obtained from TLR4-knockout mice. Neither tolerance nor a tendency to lower production of proinflammatory cytokines were evident when analyzing the production of TNF $\alpha$  and IL-6 by mast cells obtained from TLR4 KO mice, whether prestimulation was performed with LPS at 10 or 100 ng/ml. The synergistic effect of simultaneous stimulation is thus mediated by TLR4 because this phenomenon was not observed in mast cells that do not express TLR4 (Figures 2E and 2F, TLR KO).

#### Preconditioning with LPS did not affect the intensity of degranulation

For analysis of the effects of prestimulation with LPS (10 ng/ml or 100 ng/ml) on degranulation, we measured the percentage of  $\beta$ -hexosaminidase released after stimulation with antigen (20 ng/ml), LPS (1  $\mu$ g/ml) or a combination of the two. There was no significant change in the percentage of degranulation in response to any of the stimuli (data not shown).

#### Preconditioning with LPS determines cross-tolerance to the toll-like receptor 2 agonist P3C

After analyzing the influence of TLR4 expression on tolerance phenomena in mast cells, we tried to determine

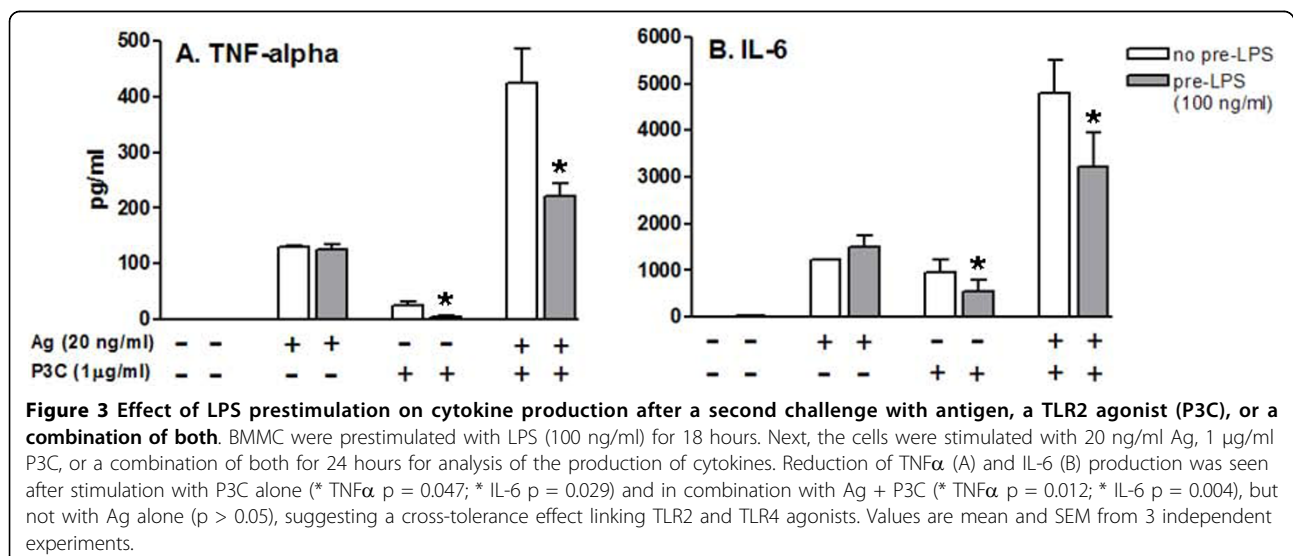
a possible common pathway linking TLR4 and TLR2 responses in endotoxin tolerance. BMMC were prestimulated with LPS (100 ng/ml) for 18 hours. After 18 hours, the cells were stimulated with antigen (20 ng/ml), P3C (1  $\mu$ g/ml), or a combination of both for 24 hours for analysis of the production of cytokines. Reduction of TNF $\alpha$  and IL-6 production after stimulation with P3C alone (TNF $\alpha$   $p = 0.047$ ; IL-6  $p = 0.029$ ) or with the combination of antigen + P3C (TNF $\alpha$   $p = 0.012$ ; IL-6  $p = 0.004$ ) was observed, but this reduction was not seen with antigen alone (Figure 3A and 3B).

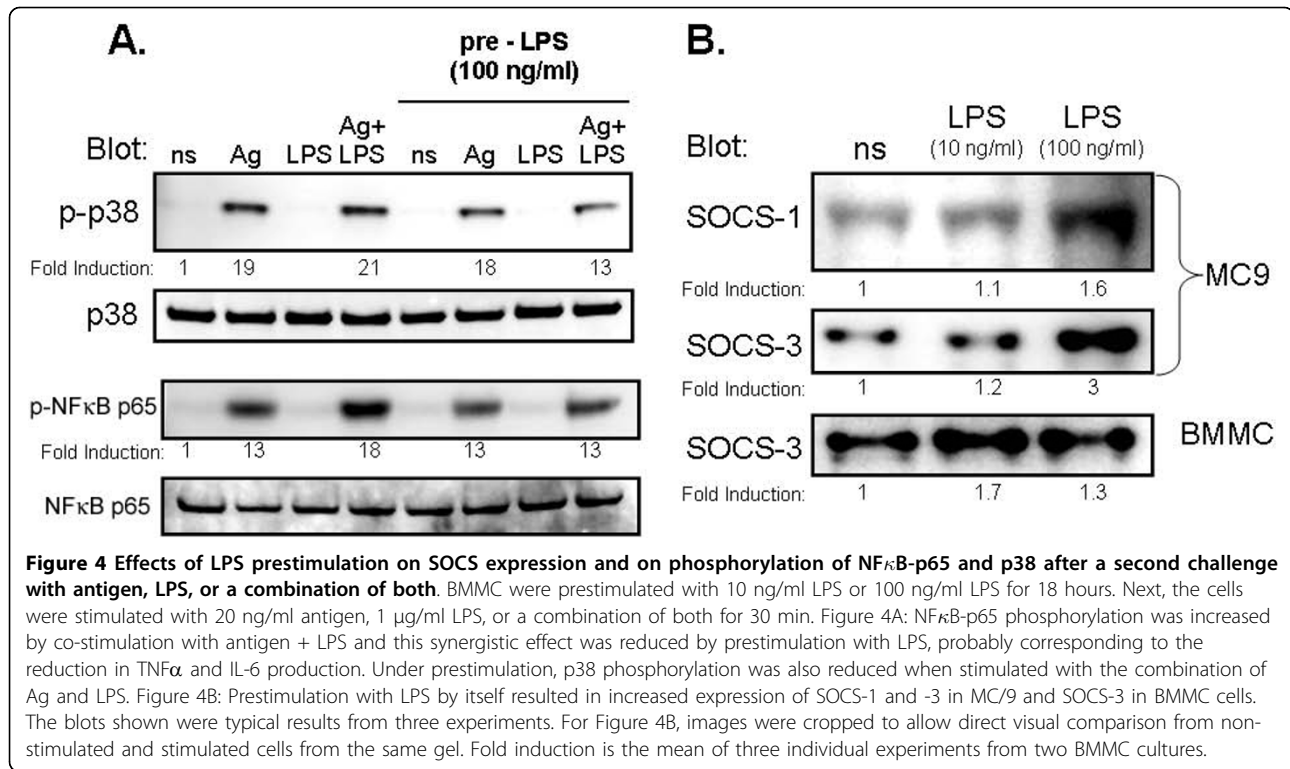
#### Prestimulation with LPS leads to an increase in SOCS expression and in attenuation of phosphorylation of NF $\kappa$ B and p38 MAPkinase after co-stimulation with antigen and LPS

We also conducted experiments with BMMC that have been prestimulated with LPS (100 ng/ml) and with non-prestimulated cells as a control group. After 18 hours, pre- and non-prestimulated BMMC were stimulated with 1  $\mu$ g/ml LPS, 20 ng/ml antigen, or a combination of both for 30 min. NF $\kappa$ B phosphorylation was increased by co-stimulation with antigen and LPS, and this synergistic effect was reduced by prestimulation with LPS. After prestimulation with LPS, p38 phosphorylation was also reduced when stimulated with the combination of antigen and LPS (Figure 4A). Prestimulation with LPS alone resulted in increased expression of SOCS-1 and -3 in MC/9 cells and increased SOCS-3 in BMMC cells (Figure 4B). We could not detect expression of SOCS-1 in BMMC cells.

#### Discussion

With the increased understanding of the immune response-regulating mechanisms introduced during the





second half of the 1990 s, models of tolerance to endotoxins have been seen as a tool to identify mediators and opportunities in modulation of the inflammatory response, particularly in sepsis [23]. Our results elucidate the phenomenon of direct endotoxin tolerance mediated by TLR4-NFκB pathway and cross-tolerance of TLR2 agonists in mast cells. Furthermore, we show synergism between TLR4 and FcεR1 agonists and attenuation of this response after prestimulation with LPS. These findings may result at least in part from a reduction in the phosphorylation of the mitogen-activated protein kinase, p38, and the transcription factor NFκB, as well as the increase in expression of SOCS-1 and -3 in mast cells.

Membranes of mast cells express Toll-like receptors that, under stimulation by components of pathogens, lead these cells to release cytokines, particularly TNFα and other effector molecules. These cytokines then control the behavior of other cells, promoting inflammation and afflux to the infection site [24]. The reprogramming induced by LPS in mast cells that we have shown here offers the possibility of evaluating the mechanism of modulation of the inflammatory response at its roots, both in response to first contact with the pathogen and at the level of consequent signaling events. We show that this reprogramming is not determined by reduction of TLR4 expression but is mediated by TLR4-NFκB signaling pathway. Reports of the possibility that negative

regulation of the immune response might be mediated by TLR [25] may be extrapolated to address mast cells.

The synergistic interaction between TLR and FcεR1 receptors was demonstrated by our prior results [18]. The results herein confirmed this synergy and added information regarding interaction between these receptors, showing the attenuation of synergy by pre-conditioning through TLR4 stimulation with LPS. We also show the fundamental role of TLR4 in potentiation after co-stimulation, which is absent in BMMC from TLR4 KO mice. The functional correspondence to human mast cells with respect to TLR4 and FcεR1 responses was demonstrated in terms of immune response modulation, including the analysis of genetic expression [26]. Regarding sepsis, common genetic foundations define the connecting link between the risk of sepsis with acute pulmonary injury and allergic phenomena such as asthma [14]. Inflammatory response-regulating mechanisms associated with the tolerance phenomenon identified in this context can possibly be common to an infectious or allergic response.

Of note is the identification of a cross-tolerance effect linking TLR4 and TLR2 agonists, which is mediated by a factor that can influence the transduction mechanisms of both receptors in mast cells. This suggests a common signaling pathway or protein that can be a target of immunomodulation of these receptors. As these receptors are responsible for recognizing components of

gram-negative (TLR4) and gram-positive (TLR2) bacteria, they have become a major target to blunt the pro-inflammatory response in sepsis. The phenomena of cross-tolerance has been previously described [27], and our observations regarding mast cells are new.

We showed attenuation of phosphorylation induced by reprogramming of mast cells after prestimulation with LPS, particularly in the TLR4-NF $\kappa$ B pathway (probably corresponding to the reduction in TNF $\alpha$  and IL-6 production observed in our experiments). These results are vital data in the search for mediators that may be used in an attempt to obtain a more balanced inflammatory response. Several negative regulators have been described as being involved in the endotoxin tolerance process, such as SH2-containing inositol phosphatase (SHIP) [28]. BMMC from SHIP knockout mice do not display endotoxin tolerance, probably acting in concert with IRAK-M and SOCS-1 to downregulate the TLR signaling pathway. In SHIP<sup>+/+</sup> cells LPS tolerance is associated with inhibition of NF- $\kappa$ B activation, one of the principal endpoints of LPS signal transduction [28]. In a recent report, SOCS-1 and -3 were described as suppressor factors involved in a coordinated negative regulation of innate and adaptive immune responses in macrophages, dendritic cells and T-lymphocytes [29]. In our experiments, we show that increases in expression of SOCS-1 and -3 were associated with tolerance and cross-tolerance in mast cells. SOCS mediates a negative feedback mechanism in LPS induced TLR4 transduction, possibly acting downstream in MyD88 dependent pathway, at IRAK and p50/p65 levels, in order to reduce the production of proinflammatory cytokines [30]. The negative regulation at p50/p65 level may be involved in Fc $\epsilon$ R1/TLR4 attenuation of the synergistic activation of these receptors. Moreover, SOCS 3 inhibit ubiquitination of TRAF6, preventing association and activation of TAK1 in the TLR pathway [30]. As TRAF6 contributes to Fc $\epsilon$ R1 mediated cytokine production in mast cells [31], this mechanism is potentially relevant in further studies addressing the interaction between TLR and Fc $\epsilon$ R1 receptors. Accordingly, in mast cells, evidence has accumulated on their immunomodulating properties, presenting new opportunities to explore these cells in inflammatory response imbalance processes [32], such as sepsis or bronchial asthma.

## Conclusions

Mast cells express endotoxin tolerance, and our data demonstrate dependence on TLR4 for this phenomenon to occur in response to co-stimulation with LPS and antigen. These facts strengthen the evidence of interaction between inflammatory responses caused by infection or allergy through TLR4 and Fc $\epsilon$ R1 receptors. Moreover, evidence in the literature on the role of mast

cells in innate immunity and the results presented here regarding cross-tolerance between TLR4 and TLR2 highlight the relevance of these findings with respect to sepsis. The association of endotoxin tolerance, cross-tolerance involving TLR4, TLR2, Fc $\epsilon$ R1 receptors, and SOCS expression implicate these proteins as potential modulators of innate and adaptive responses. In mast cells, recently defined as coordinators of innate and adaptive immunity, use of these potential modulators likely produces a more balanced inflammatory response in processes in which imbalance determines damage, such as in sepsis and allergic diseases.

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## Authors' contributions

SFS was responsible for study design, acquisition of data, analysis and interpretation of data, and drafting of the manuscript. ROP performed the western blotting. JRCM analyzed and interpreted the data. MVA was responsible for study design, acquisition of data, the analysis and interpretation of data and drafting of the manuscript. All authors have read and approved the final manuscript.

## Competing interests

The authors declare that they have no competing interests.

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## References

1. Martin GS, Mannino DM, Eaton S, Moss M: **The Epidemiology of sepsis in the United States from 1979 through 2000.** *N Engl J Med* 2003, **348**:1546-1554.
2. Dellinger RP, Levy MM, Carlet JM, Bion J, Parker MM, Jaeschke R, Reinhart K, Angus DC, Brun-Buisson C, Beale R, Calandra T, Dhainaut J, Gerlach H, Harvey M, Marini JJ, Marshall J, Ranieri M, Ramsay G, Sevransky J, Thompson BT, Townsend S, Vender JS, Zimmerman JL, Vincent J, for the International Surviving Sepsis Campaign Guidelines Committee: **Surviving Sepsis Campaign: International guidelines for management of severe sepsis and septic shock: 2008.** *Crit Care Med* 2008, **36**:296-327.
3. Thomas L: **Germes.** *N Engl J Med* 1972, **287**:553-555.
4. Cohen J: **The immunopathogenesis of sepsis.** *Nature* 2002, **420**:885-891.
5. Lemaître B, Nicolas E, Michaut L, Reichhart JM, Hoffmann JA: **The dorsoventral regulatory gene cassette spätzle/Toll/cactus controls the potent antifungal response in Drosophila adults.** *Cell* 1996, **6**:973-983.
6. Poltorak A, He X, Smirnova I, Liu MY, Van Huffel C, Du X, Birdwell D, Alejos E, Silva M, Galanos C, Freudenberg M, Ricciardi-Castagnoli P, Layton B, Beutler B: **Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: mutations in Tlr4 gene.** *Science* 1998, **282**:2085-2088.
7. Beutler B: **Inferences, questions and possibilities in Toll-like receptor signaling.** *Nature* 2004, **430**:257-263.
8. Echtenacher B, Männel DN, Hültner L: **Critical protective role of mast cells in a model of acute septic peritonitis.** *Nature* 1996, **381**:75-77.
9. Galli SJ, Nakae S, Tsai M: **Mast cells in the development of adaptive immune responses.** *Nat Immunol* 2005, **6**:135-142.
10. Saturnino SF, Andrade MV: **Toll-Like Receptors, New Horizons in Sepsis.** *Curr Mol Med* 2007, **7**:522-531.

11. Nakano N, Nishiyama C, Kanada S, Niwa Y, Shimokawa N, Ushio H, Nishiyama M, Okumura K, Ogawa H: **Survival from polymicrobial infections involvement of mast cells in IL-12/23 p40 production is essential for survival from polymicrobial infections.** *Blood* 2007, **109**:4846-4855.
12. Maurer M, Wedemeyer J, Metz M, Piliponsky AM, Weller K, Chatterjea D, Clouthier DE, Yanagisawa MM, Tsai M, Galli SJ: **Mast cells promote homeostasis by limiting endothelin-1-induced toxicity.** *Nature* 2004, **432**:512-516.
13. Mullen-St Clair J, Pham CT, Villalta SA, Caughey GH, Wolters PJ: **Mast cell dipeptidyl peptidase I mediates survival from sepsis.** *J Clin Invest* 2004, **113**:628-634.
14. Supajatura V, Ushio H, Nakao A, Okumura K, Ra C, Ogawa H: **Protective roles of mast cells against enterobacterial infection are mediated by Toll-like receptor 4.** *J Immunol* 2001, **167**:2250-2256.
15. Gao L, Grant A, Halder I, Brower R, Sevransky J, Maloney JP, Moss M, Shanholtz C, Yates CR, Meduri GU, Shriver MD, Ingersoll R, Scott AF, Beaty TH, Moitra J, Ma SF, Ye SQ, Barnes KC, Garcia JG: **Novel polymorphisms in the myosin light chain kinase gene confer risk for acute lung injury.** *Am J Respir Cell Mol Biol* 2006, **34**:487-495.
16. Braun-Fahrlander C, Riedler J, Herz U, Eder W, Waser M, Grize L, Maisch S, Carr D, Gerlach F, Bufe A, Lauener RP, Schierl R, Renz H, Nowak D, von Mutius E, Allergy and Endotoxin Study Team: **Environmental exposure to endotoxin and its relation to asthma in school-age children.** *N Engl J Med* 2002, **347**:869-877.
17. Yoshioka M, Fukuishi N, Iriguchi S, Ohsaki K, Yamanobe H, Inukai A, Kurihara D, Imajo N, Yasui Y, Matsui N, Tsujita T, Ishii A, Seya T, Takahama M, Akagi M: **Lipoteichoic acid downregulates FcεRI expression on human mast cells through Toll-like receptor 2.** *J Allergy Clin Immunol* 2007, **120**:452-461.
18. Qiao H, Andrade MV, Lisboa FA, Morgan K, Beaven MA: **FcεRI and toll-like receptors mediate synergistic signals to markedly augment production of inflammatory cytokines in murine mast cells.** *Blood* 2006, **107**:610-618.
19. Paul B: **Beeson and With the Technical Assistance of Elizabeth Roberts. Tolerance to bacterial pyrogens: I. Factors influencing its development.** *J Exp Med* 1947, **86**:29-38.
20. Tkaczyk C, Beaven MA, Brachman SM, Metcalfe DD, Gilfillan AM: **The phospholipase Cγ1-dependent pathway of FcεRI-mediated mast cells activation is regulated independently of phosphatidylinositol-3-Kinase.** *J Biol Chem* 2003, **278**:48474-48484.
21. Baetz A, Frey M, Heeg K, Dalpke AH: **Suppressor of cytokine signaling (SOCS) proteins indirectly regulate toll-like receptor signaling in innate immune cells.** *J Biol Chem* 2004, **279**:54708-15.
22. Gordon JR, Galli SJ: **Release of both preformed and newly synthesized tumor necrosis factor alpha (TNF-alpha)/cachectin by mouse mast cells stimulated via the Fc epsilon RI. A mechanism for the sustained action of mast cell-derived TNF-alpha during IgE-dependent biological responses.** *J Exp Med* 1991, **174**:103-107.
23. Cavaillon JM, Adib-Conquy M: **Bench-to-bedside review: endotoxin tolerance as a model of leukocyte reprogramming in sepsis.** *Crit Care* 2006, **10**:233.
24. Dawicki W, Marshall JS: **New and emerging roles for mast cells in host defence.** *Curr Opin Immunol* 2007, **19**:31-38.
25. Liew FY, Xu D, Brint EK, O'Neill LA: **Negative regulation of toll-like receptor-mediated immune responses.** *Nat Rev Immunol* 2005, **5**:446-458.
26. Okumura S, Kashiwakura J, Tomita H, Matsumoto K, Nakajima T, Saito H, Okayama Y: **Identification of specific gene expression profiles in human mast cells mediated by Toll-like receptor 4 and FcεRI.** *Blood* 2003, **102**:2547-2554.
27. de Vos AF, Pater JM, van den Pangaart PS, van't Veer C, van der Poll T: **In vivo lipopolysaccharide exposure of human blood leukocytes induces cross-tolerance to multiple TLR ligands.** *J Immunol* 2009, **183**:533-542.
28. Sly LM, Rauh MJ, Kalesnikoff J, Song CH, Kristal G: **LPS-induced upregulation of SHIP is essential for endotoxin tolerance.** *Immunity* 2004, **21**:227-239.
29. Dimitriou ID, Clemenza L, Scotter AJ, Chen G, Guerra FM, Rottapel R: **Putting out the fire: coordinated suppression of the innate and adaptive immune systems by SOCS1 and SOCS3 proteins.** *Immunol Rev* 2008, **224**:265-283.
30. Frobøse H, Rønn SG, Heding PE, Mendoza H, Cohen P, Mandrup-Poulsen T, Billestrup N: **Suppressor of cytokine signaling-3 inhibits interleukin-1 signaling by targeting the TRAF-6/TAK1 complex.** *Mol Endocrinol* 2006, **20**:1587-1596.
31. Yang YJ, Chen W, Carrigan SO, Chen WM, Roth K, Akiyama T, Inoue J, Marshall JS, Berman JN, Lin TJ: **TRAF6 specifically contributes to FcεRI-mediated cytokine production but not mast cells degranulation.** *J Biol Chem* 2008, **283**:32110-32118.
32. Galli SJ, Grimbaldston M, Tsai M: **Immunomodulatory mast cells: negative, as well as positive, regulators of immunity.** *Nat Rev Immunol* 2008, **8**:478-86.

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