RESEARCH

and calcium levels, and alkaline phosphatase activity among people living with and without human immunodeficiency virus and injecting drugs in kenya

Circulating 25-hydroxycholecalciferol

Abel O. Onyango¹, Nathan Shaviya², Valentine Budambula³, George O. Orinda¹, Omu Anzala⁴, Ahmed A. Aabid⁵ and Tom Were^{6*}

Abstract

Background People who inject drugs (PWID) and living with the human immunodeficiency virus (PLHIV) are at higher risk of suffering marked derangements in micronutrient levels, leading to poor disease and treatment outcomes. Consequently, this can be monitored by measuring key biomarkers, such as total circulating (serum) 25-hydroxycholecalciferol ($25(OH)D_3$), calcium, and alkaline phosphatase (ALP) for timely intervention. Therefore, circulating levels of 25(OH)D₃ and calcium, and ALP activity were determined in PWID and are highly active antiretroviral treatment (HAART)-experienced or -naive, along with those without HIV infection.

Methods This cross-sectional study compared serum concentrations of 25(OH)D₃, calcium, and ALP in Kenyan PLHIV and were HAART-naive (n = 30) or -experienced (n = 61), PWID and without HIV (n = 132).

Results Circulating $25(OH)D_2$ levels were significantly different amongst the study groups (P < 0.001), and were significantly lower in the HAART-experienced (median, 17.3; IQR, 18.3 ng/ml; P<0.001) and -naive participants (median, 21.7; IQR, 12.8 ng/ml; P=0.015) relative to uninfected (median, 25.6; IQR, 6.8 ng/ml) PWID. In addition, the proportions of vitamin D deficiency (55.7%, 40.0%, and 17.4%) and insufficiency (31.1%, 53.3%, and 63.6%) compared to sufficiency (13.1%, 6.7%, and 18.9%; P < 0.001) were greater amongst HAART-experienced, -naive, and uninfected study groups, respectively. Likewise, serum total calcium concentrations were lower in the HAART-experienced relative to HIV-negative (P = 0.019) individuals. Serum ALP activity was also lower in the HAART-experienced in contrast to HIV-negative PWID (P = 0.048). Regression analysis indicated that predictors of circulating 25(OH)D₃ were: age ($\beta = 0.287$; $R^2 = 8.0\%$; P = 0.017) and serum ALP ($\beta = 0.283$; $R^2 = 6.4\%$; P = 0.033) in the HAART-experienced PWID, and serum ALP (β = 0.386; R² = 14.5%; P < 0.001) in the HIV-negative PWID.

*Correspondence: Tom Were mugogwe@yahoo.com

Full list of author information is available at the end of the article



© The Author(s) 2024. Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.



Open Access

Conclusion This study suggests that HIV-1 infection and HAART, including injection substance use, decrease circulating $25(OH)D_3$, calcium and ALP activity. In addition, age and ALP activity are associated with low circulating vitamin D levels in HAART-experienced PWID. The results highlight the importance of incorporating vitamin D and calcium supplementation in treatment and rehabilitation protocols for PLHIV.

Keywords 25-hydroxycholecalciferol, Calcium, Alkaline phosphatase, People-who-inject-drugs, People-living-with-HIV

Background

People living with HIV (PLHIV) and those injecting illicit drugs suffer marked micronutrient and macronutrient deficiencies [1-3]. Vitamin D is one of the most important micronutrients altered in PLHIV and those injecting drugs [4–6]. Adequate circulating vitamin D levels in the body are critical in modulating clinical outcomes of HIV infection [7, 8]. Vitamin D is a key regulator of bone homeostasis [9, 10]. In addition, it is involved in regulating immune responses such as the activation of cell-mediated immunity, suppression of leucocyte proliferation, monocyte activation, and cytokine production [11]. Vitamin D deficiency has been reported in PLHIV [12, 13], and this is associated with low intake, as well as the use of efavirenz, nevirapine, tenofovir and ritonavir containing antiretroviral regimens [4, 14]. Nonetheless, supplementation restores vitamin D status, calcium, and alkaline phosphatase (ALP) activity [15]. However, it is not clear how concurrent HIV infection, highly active antiretroviral therapy (HAART) and injecting drug use influence vitamin D status.

Calcium is important in bone mineralisation, but previous studies reported decreased serum calcium levels in PLHIV [15]. HIV infection alters bone metabolism through inflammatory responses [16]. For instance, pro-inflammatory cytokines such as tumour necrosis (TNF)- α function by inhibiting osteoblasts and activating osteoclasts, hence elevating circulating levels of calcium [17]. Although elevated serum calcium independent of low vitamin D levels has been reported in heroin addicts [18], the effect of substance use on circulating calcium levels is not clear.

ALP is an enzyme produced in the liver and osteoblasts that hydrolyses phosphate esters releasing inorganic phosphate, and serum ALP activity is elevated in PLHIV initiated on HAART presenting with severe hepatotoxicity [19]. Likewise, elevated serum ALP activity predicts the degree of hepatic inflammation in chronic hepatitis B infection and marijuana-induced hepatotoxicity, as well as hepatobiliary and bone diseases [20–23]. In addition, previous studies on PLHIV showed that elevated serum ALP activity was associated with immunodeficiency (CD4 count<200 cells/µl), laboratory markers of bone turnover, and non-nucleoside reverse-transcriptase inhibitors (NNRTI; nevirapine and efavirenz) use [24, 25]. Furthermore, use of nucleoside reverse transcriptase inhibitors (NRTIs) such as tenofovir, co-morbidities and demographic factors has also been associated with alterations in serum ALP activity in PLHIV [24]. Nevertheless, no clear mechanisms have been put forth to explain serum ALP elevation in HAART-naive and -experienced PWID living with HIV.

Serum 25-hydroxycholecalciferol (also known as calcifediol or calcidiol and abbreviated as (25(OH)D₃) concentrations are a summation of vitamin D intake and sunlight exposure synthesised vitamin D, and as such is used as a biomarker of the overall vitamin D status because of a longer half-life of 2-3 weeks compared to 4-6 h for 1,25-dihydroxycholecalciferol or calcitriol $(1,25-(OH)_2D_3)$ [26]. Vitamin D from sunlight exposure, diet, and supplements is hydroxylated in the liver to $25(OH)D_3$ and in the kidneys to generate the active form $1,25-(OH)_2D_3$ [27], which promotes calcium and phosphate conservation [28]. Altogether, it appears that the homeostatic balance of these bone mineralisation markers is markedly altered in PWID and living with HIV. However, there are no reports from Kenya on the interrelationships of serum 25(OH)D₃, calcium, and ALP in PWID and are living with HIV. Therefore, it is possible that the increasing population of PWID in Kenya with a high burden of HIV infection suffer marked pathophysiologic derangements which can influence strategies of management. Therefore, this study examined the interrelationship of serum 25(OH)D₃ with calcium and ALP in HAART-experienced or -naive, and HIV-negative PWID.

Methods

Selection and description of participants. This crosssectional study was conducted as part of a larger study investigating the demographic and laboratory factors associated with HIV infection amongst PWID in Mombasa, a coastal city in Kenya. A detailed description of the study site and the population is presented in our previous publications [29–32]. A total sample size of 223 serum specimens from PWID was estimated [33] based on a margin of error of 5%, confidence interval of 95%, response distribution of 82.3%, and a population of 49,167 PWID in Kenya [34]. The sample size was then stratified according to HIV prevalence of 41% in PWID [35], and HAART cover of 0.67% in PWID living with HIV [36]. Thus, the following three groups of PWID were analysed: (1) HIV-negative (n=132); (2) HAART-naive (n=30); and (3) HAART-experienced (n=61). The HAART-naive were individuals newly diagnosed with HIV infection. The HAART-experienced were individuals on HAART, and HIV-negative comprised PWID testing negative for HIV infection. Demographic information, substance use profile, body mass index (BMI), CD4+T cell counts, including HIV screening and viral load determinations, and sample collection procedures were previously described [31, 37].

25-hydroxycholecalciferol, Serum calcium, and alkaline phosphatase activity. About 10 ml of blood samples were collected by venepuncture into plain vacutainer tubes containing a clot activator and used for serum preparation. The serum samples were aliquoted, and stored frozen at -70° C until used for batched analyte measurements. Automated clinical chemistry and immunoassay analyser (ROCHE COBAS® e601 and e501, Lausanne, Switzerland) respectively were used for batched measurements of 25(OH)D₃, and total calcium, while DIRUI CS-4000 (Dirui Industrial Company ltd., Changchui, China) auto-chemistry analyser was used for determining ALP activity. Serum 25(OH)D₃ was used for measuring vitamin D status because of a longer half-life than the $1,25-(OH)_2D_3$ [26]. In the Arsenazo method, total serum calcium is determined at an acidic pH which frees complexed and albumin-bound calcium for specific binding of calcium ions to arsenazo III (2,2'-[1,8-dihydroxy-3,6-disulphonaphthylene-2,7-bisazo]bisbenzenearsonic acid). The intensity of the purple-coloured reaction product is proportional to the concentration of total calcium present in the sample and was quantified by colorimetry. In addition, calcium estimation was based on clinical practice standards [38], as the method measures both bound and free calcium. The ROCHE COBAS® reagents were supplied by Roche diagnostics through a local subsidiary (Sciencescope ltd., Nairobi, Kenya). Quality control assays for 25(OH)D₃ and calcium were performed prior to analysing the samples. All analyses were carried out in accordance with the principles of good clinical laboratory practices.

Statistical analysis. Statistical data analysis was conducted using IBM SPSS Statistics for Windows, Version 25.0. Armonk, NY: IBM Corp. Age, weight, height, BMI, CD4+T cell counts, HIV-1 RNA copies, $25(OH)D_3$, calcium, and ALP were compared across the study groups using Kruskal Wallis U tests followed by Dunn's posthoc corrections. Distributions of gender, BMI, immune, HIV-1 viraemia, $25(OH)D_3$, calcium, and ALP status were compared amongst the study groups using the Pearson's chi-square tests. To determine the prevalence of micronutrient deficiency, the serum concentrations of 25(OH) D_3 were categorised as sufficient (≥ 30 ng/ml), insufficient (21-29 ng/ml) and deficient (< 20 ng/ml) [39]; total calcium was stratified into hypocalcaemia (< 2.2 mmol/L) and hypercalcaemia (>2.6 mmol/L), whereas ALP was categorised into low ALP activity (<53.0 IU/L) as previously established for Kenyan adults [40]. Linear hierarchical regression modelling was performed to determine the predictors of circulating 25(OH)D₃ concentrations. First, the variables were log-transformed towards normality before regression modelling each study group of PWID. In all the study groups, serum concentrations of 25(OH)D₃ were entered as the dependent variable. Calcium plus age, CD4+T cells, HIV-1 RNA copies, and BMI or age, CD4+T cells, and BMI were entered in the models as the predictor variables for the PWID and were HAART-experienced, -naive, and were HIV-negative, respectively. All tests were two-tailed with statistical significance set at P<0.05.

Results

Demographic, drug use, and clinical profiles of the study participants. The demographic and drug use profiles of the study participants are presented in Table 1. The median age distribution was not significantly different (P=0.441) across the study groups. Gender distribution differed significantly across the study groups (P < 0.001), and the HAART-experienced group had more females (n=38; 62.3%) relative to HAART-naive (n=13; 43.3%)and HIV-negative (n=12; 9.1%) PWID. Body height (m) was significantly different in between the study groups (P<0.01). CD4+T cell counts (/µl) were significantly different across groups (P < 0.001) with HAARTexperienced (P < 0.001) and HAART-naive (P < 0.001) participants presenting with lower counts compared to HIV-negative individuals. In addition, proportions of immune suppression (CD4+T cell counts<500.0/µl) were (n=41; 67.2%) in the HAART-experienced, -naive (*n*=15; 50.0%), and HIV-negative (*n*=26; 19.7%; *P*<0.001) PWID. Heroin was the most frequently injected substance in all study groups but the proportion of users varied among the study groups [HAART-experienced (n=42; 68.9%); HAART-naive (n=22; 73.3%) compared to the HIV-negative group (n=120; 90.9%); P<0.001]. Injection cocaine use was reported in less than 30.0% of the HIV-infected groups [HAART-experienced (n=18; 29.5%); HAART-naive (n=7; 23.3%) compared to the HIV-negative group (n=9; 6.8%); P<0.001]. Concomitant injection of cocaine and heroin was reported in less than 3.5% of the study participants [HAART-experienced (n=1; 1.6%); -naive (n=1; 3.3%); and HIV-negative (n=3; 1.6%)2.3%) individuals]. Frequency of drug injection (>twice a day) was higher in HAART-experienced (n=50; 82.0%) and -naive (n=19; 63.3%) individuals compared to the HIV-negative (*n*=72; 54.5%; *P*=0.001] PWID. Finally, the duration of injection (>1 year) was also higher in the HAART-experienced (n=55; 90.2%) and -naive (n=27;

Characteristic	HIV[-]/HAART[-], n=132	HIV[+]/HAART[-], n=30	HIV[+]/HAART[+], n=61	Р
Age, yrs.	32.3 (9.9)	30.7 (7.6)	30.3 (8.7)	0.441
Female, n (%)	12 (9.1)	13 (43.3)	38 (62.3)	< 0.001
Weight, kg	54.5 (8.8)	54.0 (8.8)	53.0 (7.0)	0.051
Height, m	1.7 (0.1)	1.7 (0.1)	1.7 (0.1) ^a	0.005
BMI, kg/m ²	18.7 (2.8)	18.7 (2.6)	18.8 (2.4)	0.984
BMI < 18.5 kg/m ²	73 (44.7)	16 (36.7)	34 (44.3)	0.975
CD4+T cells/µl	937.0 (618.0)	495.0 (369.0) ^c	357.0 (317.5) ^b	< 0.001
<500 CD4+T cells/µl	26 (19.7)	15 (50.0)	41 (67.2)	< 0.001
Log ₁₀ HIV-1 RNA copies/ml		3.5 (2.4)	2.9 (2.4)	0.451
≥1000 Log ₁₀ HIV-1 RNA copies/ml		16 (59.3)	29 (48.3)	0.345
Heroin	120 (90.9)	22 (73.3)	42 (68.9)	< 0.001
Cocaine	9 (6.8)	7 (23.3)	18 (29.5)	< 0.001
Cocaine and heroin	3 (2.3)	1 (3.3)	1 (1.6)	
Frequency of injection > 2/day	72 (54.5)	19 (63.3)	50 (82.0)	0.001
Duration of injection > 1 year.	81 (61.4)	27 (90.0)	55 (90.2)	< 0.001

Table 1 Demographic and clinical profiles of the study participants

Data are presented as number (n) and proportion (%) of participants for categorical variables, and as medians (interquartile range) for age, CD4 T cell counts and log_{10} HIV RNA copies. HIV, human immunodeficiency virus. HAART, highly active antiretroviral treatment. Across group comparisons were performed using the Pearson's chi-square for proportions, and Kruskal Wallis tests for age, weight, height, BMI, and CD4+T cell counts. Post-hoc Dunn's test for multiple comparisons was performed for height and CD4+T cell counts. HIV RNA copies were compared between the HAART-experienced and -naive groups using the Mann-Whitney U test. $a^{p} = 0.004$, $b^{p} < 0.001$, and $c^{p} < 0.001$ vs. uninfected people-who-inject-drugs. Values in bold are significant p-values

 Table 2
 Circulating levels of vitamin D, calcium, and alkaline phosphatase activity

Analyte	HIV(-)/ HAART(-), n=132	HIV(+)/ HAART(-), n = 30	HIV(+)/ HAART(+), n=61	Ρ
25-hydroxycholecal- ciferol, ng/ml	25.6 (6.8)	21.7 (12.8) ^b	17.3 (18.3) ^a	< 0.001
Deficiency	23 (17.4)	12 (40.0)	34 (55.7)	< 0.001
Insufficiency	84 (63.6)	16 (53.3)	19 (31.1)	
Sufficiency	25 (18.9)	2 (6.7)	8 (13.1)	
Calcium, mmol	2.3 (0.2)	2.3 (0.2)	2.2 (0.2) ^c	0.023
Hypocalcaemia	33 (25.0)	9 (30.0)	22 (36.1)	0.532
Hypercalcaemia	7 (5.3)	1 (3.3)	4 (6.6)	
ALP, U/L	61.0 (33.3)	55.0 (26.0)	53.5 (27.5) ^d	0.049
>153 U/L	0 (0.0)	0 (0.0)	0 (0.0)	

Data are presented as medians (interquartile range) for continuous variables and numbers (proportions) of participants for categorical variables. HIV, human immunodeficiency virus. HAART, highly active antiretroviral treatment. ALP, alkaline phosphatase. Deficiency, 25-hydroxycholecalciferol ($25(OH)D_3$) <20 ng/ml; isufficiency, $25(OH)D_3$ >20-29 ng/ml; sufficiency, $25(OH)D_3 \geq 30$ ng/ml [39]. Hypocalcaemia (calcium <2.2 mmol/L) and hypercalcaemia (calcium >2.6 mmol/L). High ALP activity (ALP > 153.0 U/L for males of all ages, and ALP > 130.0 and >170 U/L for females <45 years and ≥ 45 years old) [40]. Data analysis was conducted using the Pearson's chi-square for proportions and Kruskal Wallis tests for $25(OH)D_3$, calcium, and ALP levels across the groups. Post-hoc Dunn's test for multiple comparisons: ${}^aP < 0.001$, ${}^bP = 0.015$, ${}^cP = 0.0194$, and ${}^aP = 0.048$ vs. uninfected people-who-inject-drugs. Values in bold are significant P-values

90.0%) patients relative to the HIV-negative (n=81; 61.4%; P<0.001) individuals.

Serum 25-hydroxycholecalciferol, total calcium and alkaline phosphatase activity. The circulating concentrations of $25(OH)D_3$, calcium and ALP are shown in Table 2. The serum $25(OH)D_3$ levels differed significantly amongst the study groups (P<0.001). Post-hoc analysis indicated that serum $25(OH)D_3$ levels were significantly lower in the HAART-experienced (P<0.001) and -naive (P=0.015) PWID relative to the HIV-negative PWID. Consistent with lower $25(OH)D_3$ levels, the prevalence of vitamin D deficiency was higher in the PLHIV and HAART-experienced (n=34; 55.7%) vs. the -naive (n=12; 40.0%) and HIV-negative (*n*=23; 17.4%) PWID. However, proportions of 25(OH)D₃ insufficiency were lower in the HAART-experienced (n=19; 31.1%) compared to -naive (n=16; 53.3%) and HIV-negative (n=84; 63.6%) PWID. Accordingly, the overall prevalence of 25(OH)D₃ sufficiency was low in the study groups: HAART-experienced (*n*=8; 13.1%), HAART-naive (*n*=2; 6.7%), and HIV-negative (n=25; 18.9%; P<0.001) PWID. Serum total calcium levels were significantly (P=0.023) different across the study groups. Post-hoc analysis showed that serum total calcium concentrations were lower in HAART-experienced PWID in comparison to HIV-negative (P=0.019) participants. Consistent with low calcium concentrations, a higher prevalence of hypocalcaemia was present in HAART-experienced (n=22; 36.1%) vs. HAART-naive (*n*=9; 30.0%) and HIV-negative (*n*=33; 25.0% PWID. As a result, low prevalence of hypercalcaemia was present in all the study groups [HAART-experienced (n=4; 6.6%); -naive (n=1; 3.3%; and HIV-negative (n=7; 5.3%;P=0.532]. Likewise, serum activity of ALP was significantly different amongst the study groups (P=0.049). Post-hoc analyses showed that serum ALP activity was lower in the HAART-experienced PWID relative to the HIV-negative PWID (P=0.048). In addition, no high ALP activity was noted in the study groups [HAART-experienced (*n*=0; 0.0%); HAART-naive (*n*=0; 0.0%); and HIVnegative (*n*=0; 0.0%) PWID].

Predictors of circulating 25-hydroxycholecalciferol concentrations. Hierarchical linear regression modelling

for predictors of circulating 25(OH)D₃ concentrations amongst the HAART-experienced individuals was significant (F (6, 54)=3.661, P=0.004) with the entire set of variables (total calcium plus age, ALP, CD4+T cells, BMI and HIV-1 RNA copies) accounting for 28.9% of the variance in circulating 25(OH)D₃ levels (R=0.538, $R^2=0.289$). In addition, age (β =0.287, *P*=0.017), and ALP (β =0.283, P=0.033) were associated with the 25(OH)D₃ concentrations. Squared semi-partial correlations revealed that the unique 25(OH)D₃ concentrations accounted for by age, and ALP was 8.0%, and 6.4%, respectively. The modelling for the predictors of 25(OH)D₃ levels in the HAARTnaive individuals was, however, not significant (F (6, 23)=1.170, P=0.356; (R=0.484, $R^2=0.234$)). Furthermore, hierarchical regression modelling for $25(OH)D_3$ concentrations in the HIV-negative individuals was significant (F (5, 126)=5.026, P<0.001) with the entire set of variables (calcium plus age, ALP, CD4+T cells, and BMI) accounting for 16.6% of the difference in the circulating $25(OH)D_3$ concentrations (R=0.408, $R^2=0.166$). Besides, ALP activity (β =0.386, *P*<0.001) was significantly associated with the 25(OH)D₃ concentrations. Squared semipartial correlations indicated that the unique quantity of variance in 25(OH)D₃ concentrations accounted for by ALP was 14.5%.

Discussion

The lower levels, including higher proportions of deficiency and insufficiency of serum 25(OH)D₃ in the HAART-naive and -experienced PWID, suggest HIV infection and substance use exacerbation in vitamin D deficiency. These findings are consistent with previous studies showing lower concentrations of vitamin D and high proportions of vitamin D deficiency in PWID living with or without HIV [4-6]. The underlying mechanisms for the low levels of vitamin D status in PWID include HIV infection- and substance-induced chronic inflammation and immunological hyperactivity. This is further emphasised by results indicating that low levels of vitamin D are associated with seropositivity for hepatitis C virus and HIV-infections, both of which are common chronic inflammatory-associated co-morbidities in PWID [4]. The role of inflammation in suppressing the vitamin D status, is also possibly related to a shift in the oxidative and anti-oxidative balance [41] and over-secretion of inflammatory mediators such as TNF-α interfering with production of $25(OH)D_3$ resulting in vitamin D deficiency [42]. Additionally, hepatic injury and altered metabolism can lead to vitamin D deficiency given that antiretroviral drugs such as lopinavir/ritonavir, tenofovir disoproxil fumarate and efavirenz are associated with low vitamin D levels in PLHIV [43, 44]. Consistent with these observations, protease inhibitors, NRTIs and NNRTIs promote hydroxylation of vitamin D and its metabolites to biologically inactive compounds, leading to vitamin D deficiency [45, 46]. Moreover, opioids, antiretroviral drugs, and 25-(OH)D are also metabolised via the cytochrome P450 system [47, 48], resulting in interactions that possibly alter the availability of $1,25-(OH)_2D_3$.

In the present study, age, calcium, ALP, CD4+T cell count and viral load were the key predictors of serum 25-(OH)D concentrations in the study groups. These findings, in part, mirror previous studies in the USA, India, Australia, and Kenya indicating that age, low dietary intake of calcium, CD4+T cells, viral load, opioid dependence and markers of liver injury, such as alanine aminotransferase, ALP and hypoalbuminaemia in PLHIV HAART-experienced non-injecting drug users and PWID [4, 6, 14, 49, 50]. Furthermore, low vitamin D concentrations are common in people of black ethnicity, such as African and black American PLHIV [6, 51]. Therefore, a complex interplay of multiple risk factors influences the development of vitamin D deficiency and insufficiency in PWID.

Hypocalcaemia is common in HAART-naive and -experienced PLHIV [52, 53]. Consistent with previous findings, our study found lower median serum calcium levels in PLHIV HAART-experienced PWID. Previous studies indicated that PWID are largely at a high risk of under-nutrition [54]. This is possibly due to low dietary intake and limited finances, since available resources are primarily used to sustain the drug habit [2, 55]. Consequently, this contributes to the low serum calcium concentrations observed in PWID. Additionally, low calcium levels have been associated with low vitamin D concentrations, since vitamin D enhances absorption of dietary calcium [56]. Likewise, infection with HIV often leads to hypoparathyroidism [56], which is associated with hypocalcaemia. Since a majority of the study participants were using heroin, it is possible to conclude that they would present with low serum calcium levels and subsequently higher proportions of hypocalcaemia.

Although ALP appears to have no clinical utility in PWID and PLHIV, previous studies showed higher serum ALP levels in opioid-dependent individuals [57], and predicted the degree of hepatotoxicity in patients on HAART in Cameroon at one month and six-month follow-ups [19]. However, the current study found that serum ALP activity was reduced in the HAART-experienced PWID, which may be attributed to the polysubstance use in this population. Nonetheless, few studies have examined serum ALP and other hepatic enzyme activities in the context of HAART-experienced HIV infected PWID. It is important to note that our previous studies indicated elevated serum aminotransferases in HAART-experienced people-who-inject-heroin [31], and historical studies over three decades ago found that at least 18% of cocaine users had elevated serum ALP activity [58]. As such, we are proposing that polysubstance use elicits varied hepato-pathophysiological effects in PWID, warranting further investigations.

Altogether, concomitant reduction in vitamin D, calcium and ALP amongst HAART-experienced PWID suggests that substance use, HIV-infection and ARVs directly and/or indirectly alter the delicate balance of vitamin D, calcium and ALP homeostasis in these patients. It appears that mechanistically, the low levels of vitamin D drive the suppression of intestinal, renal, and bone calcium mobilisation [59]. The implications of the dysfunction in these feedback loops include revision of the clinical protocols regarding renal, liver and bone mineral function in PLHIV and injecting drugs. One of the strengths of the present study was the concurrent approach to the measurement and linking of vitamin D status with calcium and phosphate concentrations in HAART-experienced PWID. The limitations of this study are, absence of urine analyses for metabolites of vitamin D and substances used, including the effect of genetic variability in the vitamin D receptors as this would have enabled linking with vitamin D status. Another limitation of this study is the fact that this was a cross-sectional design. A prospective approach would be useful in understanding the dynamics of vitamin D status including, adherence to HAART and injecting drug cessation. However, the current comparisons of HAART-experienced with HAART-naive and HIV-negative PWID provide valuable insights into the complex pathophysiologic mechanisms of HIV infection in PWID. The laboratory analysis for vitamin D status was based on batched automated measurement of 25(OH)D₃ concentrations, and the use of local reference ranges which were consistent in this population. Even though our design was cross-sectional, and possibly limited by confounders from self-reported substance use duration, age, an important predictor of substance use in coastal Kenya [60], was used as a proxy for duration of substance use in the regression analyses. Besides, our study population was drawn from a southern latitude area amongst the native population of the coastal city of Mombasa, Kenya, and hence the findings are not generalisable to northern latitude populations.

Conclusion

The present study therefore suggests that HIV-1 infection, HAART and injection drug use concomitantly reduce vitamin D levels, calcium and ALP in PWID. Additionally, age and serum ALP activity are associated with low circulating vitamin D status in PLHIV injecting drugs and initiated on HAART. The findings of this study highlight the need for policy review on monitoring, supplementation, and rehabilitation of PLHIV injecting drugs. Further research is recommended to evaluate the effects of newer HAART regimens on serum 25(OH) D $_3$ in similar cohorts.

Abbreviations

ALP	Alkaline phosphatase
BMI	Body mass index
CD4	Clusters of differentiation 4
HAART	Highly active antiretroviral therapy
HIV	Human immunodeficiency virus
IBM	International business machines
IQR	Interquartile range
NNRTI	Non-nucleoside reverse-transcriptase inhibitor
PWID	People-who-inject-drugs
PLHIV	People living with HIV
RNA	Ribonucleic acid

SPSS Statistical Package for the Social Sciences

Acknowledgements

We thank the study participants for making this study possible. We are grateful to the management and staff of the Bomu Hospital for their support during the study. We also thank Leonard B.O. Adero and Kenneth Kimengich for technical support in quality assurance and laboratory assays.

Author contributions

TW, VB and AOO conceived and designed the study. TW and VB sourced funding. AOO, AA and VB performed the laboratory experiments. TW and NS performed statistical analyses and interpretation of data. TW and AOO co-drafted the manuscript while GOO, AA and OA critically revised the manuscript. All authors have read and approved the manuscript.

Funding

This study was supported, in part, by the Kenya National Commission for Science, Technology and Innovation [NCST/5/003/065], and Partnership for Innovative Medical Education in Kenya (NIH 1R24TW008889) grants to TW and VB.

Data availability

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

This study was conducted by the guidance of the Helsinki declarations [61]. Ethical approval for this study was obtained from the institutional ethics review committees of Masinde Muliro University of Science and Technology (MMU/COR-403012-V27) and Kenyatta University (PKU019/116/2012). Written informed consent was obtained from the study participants before enrolment into the study. Participants were educated on informed consent and the right to withdraw from the study at any stage, including on harmful effects of substance use. Confidentiality of patient information was ensured throughout the study except when required for clinical care of the patients.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Author details

 ¹Department of Biochemistry, Microbiology, and Biotechnology, Kenyatta University, P. O. Box 43844-00100, Nairobi, Kenya
 ²Department of Medical Laboratory Sciences, Masinde Muliro University of Science and Technology, P. O. Box 190-50100, Kakamega, Kenya
 ³Department of Environment and Health, Technical University of Mombasa, GPO Mombasa, P. O. Box 90420-80100, Mombasa, Kenya
 ⁴Kenya AIDS Vaccine Initiative - Institute of Clinical Research, University of Nairobi, P. O. Box 30197-00100, Nairobi, Kenya
 ⁵Bomu Hospital, P.O. Box 95683-80106, Mombasa, Kenya ⁶Department of Medical Microbiology and Parasitology, Masinde Muliro University of Science and Technology, P. O. Box 190-50100, Kakamega, Kenya

Received: 3 March 2023 / Accepted: 12 July 2024 Published online: 17 July 2024

References

- Migdanis A, Migdanis I, Papadopoulou SK, Hadjivasiliou L, Trifonova N, Villioti M, et al. Assessment of dietary intake and nutritional status of former opioid users undergoing detoxification process. Cureus. 2023;15:e50068.
- Tang AM, Bhatnagar T, Ramachandran R, Dong K, Skinner S, Kumar MS, et al. Malnutrition in a population of HIV-positive and HIV-negative drug users living in Chennai, South India. Drug Alcohol Depend. 2011;118:73–7.
- Yung TK-C, Lau JT-F. Comparing nutrient intake and body weight status amongst adolescent substance users, institutionalised abstainers and never users. Food Nutr Res. 2019;63.
- Lambert AA, Drummond MB, Mehta SH, Brown TT, Lucas GM, Kirk GD, et al. Risk factors for vitamin D deficiency among HIV-infected and uninfected injection drug users. PLoS ONE. 2014;9:e95802.
- Wang Y, Huang X, Wu Y, Li A, Tian Y, Ren M, et al. Increased risk of vitamin D deficiency among HIV-infected individuals: a systematic review and Metaanalysis. Front Nutr. 2021;8:722032.
- Zhang L, Tin A, Brown TT, Margolick JB, Witt MD, Palella FJ, et al. Vitamin D deficiency and metabolism in HIV-infected and HIV-uninfected men in the Multicenter AIDS Cohort Study. AIDS Res Hum Retroviruses. 2017;33:261–70.
- Lee S, Lee JE, Lee SO, Sim YK, Lee SH. Influence of vitamin D deficiency on the development of opportunistic infection in People Living with HIV/AIDS (PWHAs). J Am Coll Nutr. 2021;40:545–50.
- Salawu AA, Oloyede TW, Oke EO, Oladibu OT, Ojedokun SA, Oiwoh SO, et al. Vitamin D level in patients receiving highly active antiretroviral therapy in LAUTECH Teaching Hospital, Ogbomoso. West Afr J Med. 2022;39:70–5.
- 9. Fleet JC. The role of vitamin D in the endocrinology controlling calcium homeostasis. Mol Cell Endocrinol. 2017;453:36–45.
- Lisakovska O, Shymanskyi I, Labudzynskyi D, Mazanova A, Veliky M. Vitamin D auto-/paracrine system is involved in modulation of glucocorticoid-induced changes in angiogenesis/bone remodeling coupling. Int J Endocrinol. 2020;2020:8237610.
- 11. Sirbe C, Rednic S, Grama A, Pop TL. An update on the effects of vitamin D on the immune system and autoimmune diseases. Int J Mol Sci. 2022;23:9784.
- 12. Ashenafi S, Amogne W, Kassa E, Gebreselassie N, Bekele A, Aseffa G, et al. Daily nutritional supplementation with vitamin D_3 and phenylbutyrate to treatment-naïve HIV patients tested in a randomized placebo-controlled trial. Nutrients. 2019;11:133.
- Shivakoti R, Ewald ER, Gupte N, Yang W-T, Kanyama C, Cardoso SW, et al. Effect of baseline micronutrient and inflammation status on CD4 recovery post-cART initiation in the multinational PEARLS trial. Clin Nutr Edinb Scotl. 2019;38:1303–9.
- Deshwal R, Arora S. High prevalence of vitamin D deficiency in HIV infected on antiretroviral therapy in a cohort of Indian patients. J Assoc Physicians India. 2019;67:42–5.
- Noe S, Heldwein S, Pascucchi R, Oldenbüttel C, Wiese C, von Krosigk A, et al. Cholecalciferol 20 000 IU once weekly in HIV-positive patients with low vitamin D levels: result from a cohort study. J Int Assoc Provid AIDS Care. 2017;16:315–20.
- Delpino MV, Quarleri J. Influence of HIV infection and antiretroviral therapy on bone homeostasis. Front Endocrinol. 2020;11:502.
- Wang T, He C. TNF-α and IL-6: the link between immune and bone system. Curr Drug Targets. 2020;21:213–27.
- Teichmann J, Stephan E, Lange U, Discher T, Stracke H, Federlin K. Elevated serum-calcium and parathormone-levels in HIV afflicted female heroin addicts. Eur J Med Res. 1997;2:343–6.
- Abongwa LE, Nyamache AK, Charles F, Torimiro J, Emmanuel N, Domkam I, et al. Risk factors of severe hepatotoxicity among HIV-1 infected individuals initiated on highly active antiretroviral therapy in the Northwest Region of Cameroon. BMC Gastroenterol. 2022;22:286.
- 20. Borini P, Guimarães RC, Borini SB. Possible hepatotoxicity of chronic marijuana usage. Sao Paulo Med J Rev Paul Med. 2004;122:110–6.
- 21. Catalano M, Roviello G, Aprile G, Ramello M, Conca R, Petrioli R, et al. Prognostic value of alkaline phosphatase and gamma-glutamyl transferase

in patients with metastatic pancreatic cancer. Future Oncol Lond Engl. 2023;19:937–46.

- 22. Huang G, Li W, Zhong Y, Liao W, Zhang Z. Mendelian randomization to evaluate the causal relationship between liver enzymes and the risk of six specific bone and joint-related diseases. Front Immunol. 2023;14:1195553.
- 23. Yang K, Pan Y, Yan B, Yu F-R, Chen J, Liu P et al. Serum sCD14 as a biomarker for significant liver inflammation in chronic hepatitis B patients with normal or mildly elevated ALT. Clin Lab. 2021;67.
- 24. Alemu BT, Troy SB, Beydoun HA, Akpinar-Elci M, Cunningham TD. Determinants of elevated alkaline phosphatase in patients infected with HIV. South Med J. 2016;109:487–91.
- Yazie TS. Derangement of liver enzymes, hyperglycemia, anemia, and associated factors among HIV-infected patients treated with tenofovir disoproxil fumarate-based regimen in Ethiopia: a prospective cohort study. BioMed Res Int. 2021;2021:6613519.
- Holick MF, Vitamin D, Status. Measurement, interpretation and clinical application. Ann Epidemiol. 2009;19:73–8.
- 27. Bikle DD, Vitamin D. Production, metabolism and mechanisms of action. In: Feingold KR, Anawalt B, Blackman MR, Boyce A, Chrousos G, Corpas E, et al. editors. Endotext. South Dartmouth (MA). MDText.com, Inc; 2000.
- 28. Peacock M. Phosphate metabolism in Health and Disease. Calcif Tissue Int. 2021;108:3–15.
- Budambula V, Matoka C, Ouma J, Ahmed AA, Otieno MF, Were T. Sociodemographic and sexual practices associated with HIV infection in Kenyan injection and non-injection drug users. BMC Public Health. 2018;18:193.
- Ndombi EM, Budambula V, Webale MK, Musumba FO, Wesongah JO, Mibei E, et al. Serum adiponectin in HIV-1 and Hepatitis C virus mono- and coinfected Kenyan injection drug users. Endocr Connect. 2015;4:223–32.
- Were T, Wesongah JO, Munde E, Ouma C, Kahiga TM, Ongecha-Owuor F, et al. Clinical chemistry profiles in injection heroin users from Coastal Region, Kenya. BMC Clin Pathol. 2014;14:32.
- Webale MK, Budambula V, Lihana R, Musumba FO, Nyamache AK, Budambula NLM, et al. Hepatitis B virus sero-profiles and genotypes in HIV-1 infected and uninfected injection and non-injection drug users from coastal Kenya. BMC Infect Dis. 2015;15:299.
- http://www.raosoft.com/samplesize.html. Sample Size Calculator by Raosoft, Inc. http://www.raosoft.com/samplesize.html. Accessed 22 Oct 2023.
- UNODC. World Drug Report. 2022. United Nations: Office on Drugs and Crime. 2022. //www.unodc.org/unodc/en/data-and-analysis/world-drugreport-2022.html. Accessed 31 Mar 2023.
- Mathers BM, Degenhardt L, Phillips B, Wiessing L, Hickman M, Strathdee SA, et al. Global epidemiology of injecting drug use and HIV among people who inject drugs: a systematic review. Lancet Lond Engl. 2008;372:1733–45.
- 36. UNAIDS, Kenya HIV, Estimates AIDS. 2023. 2023. https://www.unaids.org/en/ regionscountries/countries/kenya. Accessed 6 Jul 2023.
- Budambula V, Musumba FO, Webale MK, Kahiga TM, Ongecha-Owuor F, Kiarie JN, et al. HIV-1 protease inhibitor drug resistance in Kenyan antiretroviral treatment-naive and -experienced injection drug users and non-drug users. AIDS Res Ther. 2015;12:27.
- Kenny CM, Murphy CE, Boyce DS, Ashley DM, Jahanmir J. Things we do for no Reason[™]: calculating a corrected calcium level. J Hosp Med. 2021;16:499–501.
- Institute of Medicine (IOM). Dietary reference intakes for calcium and Vitamin D. Washington, DC: National Academies; 2011.
- 40. Omuse G, Ichihara K, Maina D, Hoffman M, Kagotho E, Kanyua A, et al. Determination of reference intervals for common chemistry and immunoassay tests for Kenyan adults based on an internationally harmonized protocol and up-to-date statistical methods. PLoS ONE. 2020;15:e0235234.
- Baser H, Can U, Baser S, Hidayetoglu BT, Aslan U, Buyuktorun I, et al. Serum total oxidant/anti-oxidant status, ischemia-modified albumin and oxidizedlow density lipoprotein levels in patients with vitamin D deficiency. Arch Endocrinol Metab. 2015;59:318–24.
- Haug CJ, Aukrust P, Haug E, Mørkrid L, Müller F, Frøland SS. Severe deficiency of 1,25-dihydroxyvitamin D3 in human immunodeficiency virus infection: association with immunological hyperactivity and only minor changes in calcium homeostasis. J Clin Endocrinol Metab. 1998;83:3832–8.
- Calza L, di Pietro G, Colangeli V, Borderi M, Zaghi I, Malosso P, et al. Factors associated with vitamin D deficiency in HIV-1 infected patients on combination antiretroviral therapy: a case-control study. New Microbiol. 2019;42:145–9.
- Piloya TW, Bakeera-Kitaka S, Kisitu GP, Idro R, Cusick SE. Vitamin D status and associated factors among HIV-infected children and adolescents on antiretroviral therapy in Kampala, Uganda. PLoS ONE. 2021;16:e0253689.

- Klassen KM, Kimlin MG, Fairley CK, Emery S, Anderson PH, Ebeling PR, et al. Associations between vitamin D metabolites, antiretroviral therapy and bone mineral density in people with HIV. Osteoporos Int J Establ Result Coop Eur Found Osteoporos Natl Osteoporos Found USA. 2016;27:1737–45.
- 46. Maggiolo F, Rizzardini G, Raffi F, Pulido F, Mateo-Garcia MG, Molina J-M, et al. Bone mineral density in virologically suppressed people aged 60 years or older with HIV-1 switching from a regimen containing tenofovir disoproxil fumarate to an elvitegravir, cobicistat, emtricitabine, and tenofovir alafenamide single-tablet regimen: a multicentre, open-label, phase 3b, randomised trial. Lancet HIV. 2019;6:e655–66.
- Campbell SD, Gadel S, Friedel C, Crafford A, Regina KJ, Kharasch ED. Influence of HIV antiretrovirals on methadone N-demethylation and transport. Biochem Pharmacol. 2015;95:115–25.
- Ratsma DMA, Muller M, Koedam M, Zillikens MC, van der Eerden BCJ. In vitro regulation of fibroblast growth factor 23 by 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D synthesized by osteocyte-like MC3T3-E1 cells. Eur J Endocrinol. 2023;189:448–59.
- Barasa E, Shaviya N, Budambula V, Were T. rs1445776009 variants in the human ALB gene: Association with serum albumin and clinical outcomes in HIV-infected Kenyan injection substance users. Int J Health Sci. 2021;15:3–11.
- Reece AS, Hulse GK. What are the characteristics of vitamin D metabolism in opioid dependence? An exploratory longitudinal study in Australian primary care. BMJ Open. 2018;8:e016806.
- Belete AM, Tefera AA, Getnet M, Asefa A, Aynalem YA, Shiferaw WS. Prevalence and predictors of hypovitaminosis D in Ethiopian HIV-Infected adults. HIVAIDS Auckl NZ. 2021;13:1101–9.
- Moges B, Amare B, Yabutani T, Kassu A. HIV associated hypocalcaemia among diarrheic patients in northwest Ethiopia: a cross sectional study. BMC Public Health. 2014;14:679.
- Noe S, Heldwein S, Wiese C, Pascucci R, von Krosigk A, Schabaz F, et al. Tenofovir Disoproxil Fumarate is Associated with a set-point variation in the calcium-parathyroid hormone-vitamin D Axis: results from a German cohort. Adv Pharmacol Sci. 2018;2018:6069131.

- Negessie A, Jara D, Taddele M, Burrowes S. Determinants of undernutrition among adult patients receiving antiretroviral therapy at Debre Markos referral hospital, Northwest Ethiopia: a case-control study design. BMC Nutr. 2019;5:20.
- Strike C, Rudzinski K, Patterson J, Millson M. Frequent food insecurity among injection drug users: correlates and concerns. BMC Public Health. 2012;12:1058.
- Sandhu S, Desai A, Batra M, Girdhar R, Chatterjee K, Kemp EH, et al. Severe symptomatic hypocalcemia from HIV related hypoparathyroidism. Case Rep Endocrinol. 2018;2018:8270936.
- Balodimos S, Nikolaou K, Njau S, Karamouzis M, Kovatsi L. The effect of opioid dependence on conventional and novel biochemical parameters of bone metabolism. Am J Drug Alcohol Abuse. 2015;41:535–40.
- Kothur R, Marsh F, Posner G. Liver function tests in nonparenteral cocaine users. Arch Intern Med. 1991;151:1126–8.
- 59. Khammissa RAG, Fourie J, Motswaledi MH, Ballyram R, Lemmer J, Feller L. The Biological activities of vitamin D and its receptor in relation to Calcium and Bone Homeostasis, Cancer, Immune and Cardiovascular systems, skin Biology, and oral health. BioMed Res Int. 2018;2018:9276380.
- Oguya FO, Kenya PR, Ongecha F, Mureithi P, Musyoka H, Muraguri N, et al. Rapid situational assessment of people who inject drugs (PWID) in Nairobi and coastal regions of Kenya: a respondent driven sampling survey. BMC Public Health. 2021;21:1549.
- WMA. World Medical Association declaration of Helsinki ethical principles for medical research involving human subjects. J Am Med Assoc. 2013;310:2191–4.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.