

CORRECTION

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Correction to: Rapid and simple colorimetric detection of multiple influenza viruses infecting humans using a reverse transcriptional loopmediated isothermal amplification (RT-LAMP) diagnostic platform

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Correction to: BMC Infect Dis 19, 676 (2019)
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Following publication of the original article [1], the authors noticed that in Table 1, two primer sequences (B-LF and H5-LF) should have been reverse complemented.

The sequences to correct are below marked:

The correct sequences should be indicated as below:

No.	Needs Correction:	Corrected version
1.	Table 1/ B (NA gene)/B-LF Sequence: GATGTCCTGTAAGATACCAA	CTACAGGC ACATTCTATG GTT

Comment: Based on the reference in designing the forward loop (LF) primer: **LF 5'-AACCATAGAATGTGCCTGTAG-3'** (Figure 1, page 4), this identified sequence should have been reverse complemented however inadvertently the submitted sequence for the mentioned B-LF primer was only reversed.

(Continued)

2.	Table 1/ A/H5 (HA gene)/ H5-LF Sequence:	CTACCAACCATACCCATGG	GGTACCCA TACCAACCAT C
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Comment: Based on the reference in designing the forward loop (LF) primer: **LF 5'-GATGGTTGGTATGGGTACC-3'** (Figure 1, page 4), this identified sequence should have been reverse complemented however inadvertently the submitted sequence for the mentioned H5-LF primer was only reversed.

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Table 1 Reverse transcriptional loop-mediated isothermal amplification (RT-LAMP) primers for detection of influenza subtypes

Target	Gene	Primer name	Sequence(5'-3')	Primer final concentration	Gene position	Length (mer)
B	NA	B-F3	CAGGAAGAGTAAAACATACTGAGGA	5 μ M	856–881	25
		B-B3	GATTCGCAAGGCCCTGTT	5 μ M	1052–1069	18
		B-FIP	AGGGTCTTTTTGCTGTGTAAGTGT-GCACATGCCGATTTGCCAG	40 μ M	933–955 + 883–901	44
		B-BIP	GTGGAGACTGATACAGCGGAA-TGCTTCCATCATTGGTCTGG	40 μ M	972–992 + 1030–1050	42
		B-LF	CTACAGGCACATTCTATGGTT	10 μ M	908–928	21
		B-LB	ATAAGATTGATGTGCACA	10 μ M	993–1010	18
A/H1	HA	H1-F3	AGCAAGAAGTTCAAGCCG	5 μ M	619–639	18
		H1-B3	CGTGAAGTGGTGTATCTGAA	5 μ M	801–820	20
		H1-FIP	GGCTCTACTAGTGTCCAGTAATAGT-AAATAGCAATAAGACCCAAAGTG	80 μ M	734–758 + 689–711	48
		H1-BIP	ATAACATTGCAAGCAACTGGAAATC-TGATAATACCAGATCCAGCATT	80 μ M	718–742 + 778–799	47
		H1-LF	TCTCCCTTCTTGATCCC	10 μ M	713–729	17
		H1-LB	TAGTGGTACCCGAGATATGCA	10 μ M	794–813	20
A/H3	HA	H3-F3	GGGGTTACTTCAAATACAAG	5 μ M	841–859	19
		H3-B3	GTTGCCAATTTGAGAGTGTT	5 μ M	1011–1028	18
		H3-FIP	GAGTGATGCATTCAGAATTGCATT-TGGGAAAAGCTCAATAATGAGA	40 μ M	903–927 + 863–884	47
		H3-BIP	AATGGAAGCATTCCCAATGACA-GCTTAACATATCTGGGACAGG	40 μ M	930–951 + 988–1008	43
		H3-LF	CCAATGGGTGCATCTGA	10 μ M	885–901	17
		H3-LB	AACCATTCCAAAATGTAAAC	10 μ M	952–971	20
A/H5	HA	H5-F3	GCTATAGCAGGTTTTATAGAGG	5 μ M	1048–1069	22
		H5-B3	GCCTCAAAGTGTGTTTCAT	5 μ M	1210–1229	20
		H5-FIP	ACTCCCTGCTCATTGCTAT-GGATGGCAGGGAATGGTA	80 μ M	1112–1131 + 1072–1089	38
		H5-BIP	GGTACGCTGCAGACAAAGAAT-TGAGTTGACCTTATTGGTGAC	80 μ M	1133–1153 + 1177–1197	42
		H5-LF	GGTACCATACCAACCATC	5 μ M	1093–1114	19
		H5-LB	CCACTCAAAGGCAATAGATGGA	5 μ M	1154–1176	23
A/H7	HA	H7-F3	GCGGGTTTCATTGAAAATGG	5 μ M	1036–1055	20
		H7-B3	CTACCTCATTGAATCATTGTCT	5 μ M	1215–1237	23
		H7-FIP	TCCCTCTCCCTGTGCATTCT-ATGGGAAGGCCTAATTGATG	80 μ M	1097–1116 + 1056–1075	40
		H7-BIP	ACTGCTGCAGATTACAAAAGCAC-TGGTTGGTTTTTCTATAAGCC	80 μ M	1117–1139 + 1178–1199	45
		H7-LF	TCTGAAACCATACCAAC	5 μ M	1076–1092	17
		H7-LB	TCAATCGGCAATTGATCAAATA	5 μ M	1140–1161	22