A systematic review on antibiotic therapy of cutaneous bacillary angiomatosis not related to major immunocompromising conditions: from pathogenesis to treatment

Salvatore Rotundo¹, Maria Teresa Tassone¹, Nadia Marascio², Helen Linda Morrone¹, Simona Gigliotti², Angela Quirino², Alessandro Russo^{1,3}, Giovanni Matera², Enrico Maria Trecarichi^{1,3} and Carlo Torti^{4,5*}

Abstract

Background Cutaneous bacillary angiomatosis (cBA) is a vascular proliferative disorder due to *Bartonella* spp. that mostly affects people living with HIV (PLWH), transplanted patients and those taking immunosuppressive drugs. Since cBA is mostly related to these major immunocompromising conditions (i.e., T-cell count impairment), it is considered rare in relatively immunocompetent patients and could be underdiagnosed in them. Moreover, antimicrobial treatment in this population has not been previously investigated.

Methods We searched the databases PubMed, Google Scholar, Scopus, OpenAIRE and ScienceDirect by screening articles whose title included the keywords "bacillary" AND "angiomatosis" and included case reports about patients not suffering from major immunocompromising conditions to provide insights about antibiotic treatments and their duration.

Results Twenty-two cases of cBA not related to major immunocompromising conditions were retrieved. Antibiotic treatment duration was shorter in patients with single cBA lesion than in patients with multiple lesions, including in most cases macrolides and tetracyclines.

Conclusions cBA is an emerging manifestation of *Bartonella* spp. infection in people not suffering from major immunocompromising conditions. Until evidence-based guidelines are available, molecular tests together with severity and extension of the disease can be useful to personalize the type of treatment and its duration.

Keywords Bartonella, Antibiotic, Bacillary angiomatosis, Emerging disease, PCR, One health

*Correspondence: Carlo Torti carlo.torti@unicatt.it ¹Dipartimento di Scienze Mediche e Chirurgiche, Università "Magna Graecia", Catanzaro, Italy ²Dipartimento di Scienze della Vita, Unità Operativa Complessa di

Microbiologica Clinica, Università "Magna Graecia", Catanzaro, Italy

© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Common Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

³Unità Operativa Complessa di Malattie Infettive e Tropicali, Azienda Ospedaliero-Universitaria "R. Dulbecco", Catanzaro, Italy
⁴Dipartimento di Scienze di Laboratorio e Infettivologiche, Fondazione Policlinico Universitario Agostino Gemelli IRCCS, Rome, Italy
⁵Dipartimento di Sicurezza e Bioetica, Università Cattolica del Sacro Cuore, Rome, Italy



BMC Infectious Diseases



Open Access



Background

Bartonella spp. includes fastidious and arthropod-borne Gram-negative bacilli infecting both vector insects and mammalian hosts [1]. Several animals act as a reservoir since they are often asymptomatic, while fleas and other blood-sucking arthropods are the vectors of bartonellosis [2, 3]. Although many *Bartonella* spp. have been detected in both wild animals and pets, few data have been published regarding transmission of these bacteria in pets and wild animals and the possible pathogenicity for these hosts [4–7]. The most studied species in human pathology are B. henselae, B. bacilliformis and B. quintana [8]. B. bacilliformis is responsible of Carrion's disease, which stands among the array of neglected tropical diseases, often overlooked despite its significant impact on affected communities. The initial acute phase of Carrion's disease manifests with fever and haemolytic anaemia, presenting a formidable mortality rate ranging from 44 to 88% in untreated individuals [9]. Following this acute stage, a subsequent phase ensues, which may emerge weeks to months later, often with or without a history of antecedent illness. This phase is characterized by the eruption of clusters of skin lesions, categorized as nodular lesions, called verruga peruviana [9, 10] since this disease primarily affects regions within the Andean cordillera spanning across Peru, Ecuador, and Colombia. This geographical confinement is largely attributed to the behavior of its suspected primary vector, Lutzomyia verrucarum, characterized by its limited, hopping flight capabilities and sensitivity to extreme temperatures [9]. The pathological manifestations of *Bartonella* spp. infections are widely heterogeneous, including asymptomatic bacteremia, neurological disorders, myocarditis, retinitis, chronic lymphadenopathies, endocarditis, sepsis, and vascular proliferative disorders such as bacillary peliosis and cutaneous bacillary angiomatosis (cBA) [11, 12], which are caused by *B. henselae* or *B. quintana* [11, 13–22]. However, cBA and verruga peruana are difficult to distinguish since those are nearly identical presenting as angiomatous lesions [23]. Homelessness, low socioeconomic status and being infested with lice are the most common risk factors associated with cBA caused by B. quintana, while owning a cat bearing fleas or cat bites and scratches are associated with cBA due to B. hense*lae* [11, 13]. Regarding the host, cBA is mostly described in severely immunocompromised patients with T-cell response impairment such as transplant recipients, those taking immunosuppressive drugs and people living with HIV (PLWH). Only for the latter cathegory treatment guidelines are available, issued by the National Institute of Health (NIH) that recommends using doxycycline or erythromycin for at least three months [24]. However, cBA in patients not suffering from major immunocompromising conditions is considered rare [14] and there is no consensus about treatment, since no studies enrolled enough of these patients to validate type and duration of antimicrobic treatment. Therefore, we focused on patients not suffering from immunocompromising conditions and reviewed current literature to provide insights about antibiotic treatment and its duration.

Methods

Two independent reviewers searched the database PubMed, Google Scholar, Scopus, OpenAIRE and ScienceDirect by screening articles whose title included the keywords "bacillary" and "angiomatosis". Only case reports regarding cBA published until November 15th, 2023, written in English language and published in peerreviewed journals dealing with patients without major immunocompromising conditions, either ongoing or in past medical history (i.e., PLWH, organ transplant recipient, affected by hematological malignancy and/or taking immunosuppressive drugs) were included. Since we were interested in correlating the patient clinical healing with the treatment provided, only cases with available type and duration of antibiotic therapy were included. References of each article were checked to include as many cases as possible regarding cBA not related to major immunocompromising conditions. The outcome was the clinical cure (i.e., healing of skin lesions).

The systematic review protocol was registered on PROSPERO on June 30th, 2023 (registration number: CRD42023437976). The protocol and article follow the PRISMA checklist for the reporting of systematic reviews [25] (Fig. 1).

Results

Up to November 15th, 2023, 24 cases of cBA are reported in not severely immunocompromised hosts [8, 11, 14– 22, 26–36]. In one case antibiotic treatment was not prescribed [31] and in another case therapy was not reported [35]. Therefore, 22 cases were included in this analysis whose characteristics are summarized in Table 1.

Among these patients, seven were reported in the United States of America, six in Turkey, and four in India. Only one case for each country was reported in Spain, Brazil, Iran, Romania and Italy. Nine were females and ten were males, in three cases gender was not reported [22]. Six patients were children. The most common risk factor for cBA was having had a contact with a cat (7/22). Other possible risk factors were trauma (4/22), arthropod bite (3/22), burn (1/22) and having contact with a parakeet (1/22). Three patients reported multiple risk factors [22] while in nine cases no risk factors were identified. Nine patients had at least a lesion involving their head or neck and nine patients had their upper limbs involved. Nine cases showed multiple cBA lesions, while in thirteen cases patients had a single cBA lesion. In those



Fig. 1 PRISMA flowchart of study selection process

instances reporting single cBA lesions, notable heterogeneity in size was observed, ranging from as small as 5 mm to as large as 10 cm. Diagnosis was mainly made by histopathology (17/22), polymerase chain reaction (PCR) was positive in four cases on biopsy and in one case [36] was reported to be positive even on blood sample. However, in many of the other cases it was not stated whether PCR was performed or not. Culture was reported to be positive in only one case [27].

In multiple cBA lesion, lesion removal was performed for diagnostic purposes. In single cBA lesions, complete removal was reported in five cases before antibiotic treatment [11, 19, 20, 29, 36]. Only in one case [30] it was specified that the lesion was biopsied without complete removal. In the remaining cases, biopsies were performed without specifying whether the lesion was completely removed or not [8, 32–34].

Macrolides were used in most cases (17/22), being erythromycin the most frequently prescribed antibiotic (12/22) and only in two patients it was switched due to intolerance [22, 33]. Clarithromycin alone was used in three cases and in one case in association with rifampin [19]. One patient received azithromycin [11]. Tetracyclines were prescribed in seven cases: doxycycline was prescribed in six patients, including the two mentioned above who did not tolerate macrolides [22, 33]; in one patient doxycycline was used in a combined regimen and later switched due to side effect onset [36]. One patient was treated with minocycline [22]. Only one patient received a quinolone (ciprofloxacin) as monotherapy [30] (Fig. 2).

Antibiotic treatment duration was significantly (p=0.0038, Fig. 3) shorter in patients with a single cBA lesion (median 42 days, interquartile range [IQR]: 14–45 days) than in those with multiple cBA lesions (median 90 days, IQR: 51–112 days).

Discussion

In most cases included in the review (17/22), cBA affected body surfaces more susceptible to vector insect exposure which are not protected by clothing (such as head, neck and upper limbs). Indeed, despite

Table 1 Characteristics of the 22 cases included in this analysis. M: male; NR: not reported; F: female; PCR: polymerase chain reaction

Author, year of publication	Country	Patient age and sex	Risk factor(s)	Site of lesion(s)	Diagnostic method	Lesion(s)	Antibiotic (duration)
Cockerell CJ et al., 1990 [27]	United States of America	37 M	Parakeet	Forearm	Culture	Multiple	Erythromycin (3 months and 2 weeks)
Tappero JW et al., 1993 [22]	United States of America	41 NR	Cat, fleas, chiggers	Nose	Histopatholology	Single	Minocycline (4 to 6 weeks)
		74 NR	Cat, fleas, mites	Neck	PCR	Single	Erythromycin/ doxycycline* (4 to 6 weeks)
		42 NR	Cat, fleas, fire ants	Genital	PCR	Single	Erythromycin (4 to 6 weeks)
Paul MA et al., 1994 [29]	United States of America	6 F	Cat	Neck	Histopatholology	Single	Erythromycin (6 weeks)
Karakaş M et al., 2000 [16]	Turkey	21 F	Burn	Face	Histopatholology	Multiple	Erythromycin (2 months)
Gangopadhya AK et al., 2001 [26]	India	65 M	None	Forearm	Histopatholology	Multiple	Erythromycin (2 weeks)
Asharaf M et al., 2002 [18]	India	5 M	Trauma	Lips, knees, buttocks, ankles, elbows	Histopatholology	Multiple	Erythromycin (3 months)
Kayaselçuk F et al., 2002 [30]	Turkey	67 F	None	Scalp	Histopatholology	Single	Ciprofloxacin (10 days)
Karakas M et al., 2003 [17]	Turkey	32 M	None	Leg	Histopatholology	Multiple	Erythromycin (2 months)
Turgut M et al., 2004 [19]	Turkey	6 M	Trauma	Forehead	Histopatholology	Single	Clarithromycin plus rifampin (7 weeks)
Bernabeu-Wittel J et al., 2010 [8]	Spain	59 F	None	Ankle	PCR	Single	Doxycycline (2 months)
Kacar N et al., 2010 [32]	Turkey	10 M	Trauma	Leg	Histopatholology	Single	Erythromycin (1 week)
Bellissimo-Ro- drigues F et al., 2010 [34]	Brazil	32 F	None	Thumb	Histopatholology	Single	Erythromycin (4 week)
Zarraga M et al., 2011 [11]	United States of America	10 F	Cat	Chest	Histopatholology	Single	Azithromycin (14 days)
Albayrak A et al., 2011 [20]	Turkey	5 M	None	Arm	Histopatholology	Single	Erythromycin (2 months and 2 weeks)
Blattner C et al., 2014 [33]	United States of America	76 F	None	Upper lip	Histopatholology	Single	Erythromycin/ doxycycline* (2 weeks)
lraji F et al., 2015 [28]	Iran	26 F	None	Arm, fingers	Histopatholology	Multiple	Clarithromycin (3 months)
Nikam BP et al., 2018 [15]	India	45 F	Cat	Arm, fore- arm, ankle	Histopatholology	Multiple	Doxycycline (4 months)
Balaban M et al., 2019 [21]	Romania	43 M	Trauma	Face	Histopatholology	Multiple	Clarithromycin (6 weeks)
Agrawal S et al., 2022 [14]	India	45 M	None	Hands, forearm	Histopatholology	Multiple	Doxycycline (4 months)
Rotundo S et al., 2023 [<mark>36</mark>]	Italy	67 M	Cat	Forearm	PCR	Single	Doxycycline/ clarithromycin [§] plus levofloxacin (1 month)

*Erythromycin was switched to doxycycline because the patient became intolerant

[§]Doxycycline was switched to clarithromycin due to side effect



Duration of antibiotic in single cBA lesion

Duration of antibiotic treatment in multiple cBA lesions



Fig. 2 Type and duration of antibiotic treatment in single (a) and in multiple (b) cutaneous bacillary angiomatosis (cBA) lesions

Bartonella spp. is usually inoculated into the derma by a vector insect that feeds on blood [37], it was previously identified in non-blood-sucking arthropods such as *Dermatophagoides* spp [38]. and *Demodex* spp [39].. Interestingly, in 5/22 cases included in this review, burn or trauma were the only risk factors reported that could be associated with cBA. Therefore, we may hypothesize that these mites could cause both the transmission and the persistence of *Bartonella* spp. in cases in which skin is already damaged, since a bite from a blood-sucking arthropod may not necessarily be the only way of transmission of *Bartonella* spp. infection [38, 39] although no currently available data suggest that these arthropods are associated with *Bartonella* spp. infection in humans.

Once *B. henselae* or *B. quintana* reaches the skin, it eludes phagocytosis with mechanisms that are still being clarified [36, 40, 41]. For instance, *B. quintana* developed a low pathogenic lipopolysaccharide (LPS) which suppresses the classical activation of toll-like receptor (TLR)-4 preventing the production of acute phase



Fig. 3 Comparison of duration of antibiotic therapy between single and multiple cutaneous bacillary angiomatosis. Statistical analysis was performed by GraphPad Prism 9.0 Version 9.3.1 (GraphPad Software, San Diego, CA 92,108), and the data are expressed as median \pm interquartile range. Mann-Whitney test was applied to analyze duration of antibiotic therapy in the two groups. Exact duration of antibiotic therapy in patients reported by Tappero JW et al. [22] was not specified and ranged from four to six weeks. The maximum duration of antibiotic therapy (six weeks) was considered for this analysis

proteins by peripheral blood mononuclear cells [42]. Moreover, B. henselae and B. quintana hide in mesenchymal staminal and endothelial cells and upregulate several angiogenic factors with both direct mitogenic and antiapoptotic properties (including chemokines, interleukins and the vascular endothelial growth factor) [21, 43, 44] which have a key role in developing angioproliferative lesions [37] and in escaping the immune system. Growth of these angioproliferative lesions depends on the persistence of bacteria in blood vessels and could be reverted by Bartonella spp. eradication [45]; indeed, some antibiotics demonstrated not only activity against the bacteria but also direct modulation of endothelial cell proliferation [46] Notably, B. bacilliformis, B. henselae, and B. quintana share similarities in producing angiogenic factors, suggesting a common mechanism underlying endothelial proliferation [10]. The main host factor involved in hampering cBA appears to be the T-cell response [47]. Indeed, T-cells play a pivotal role in controlling Bartonella spp. infection since these cells support a Th-1 response and activate macrophages in infections due to *Bartonella* spp [10].. Moreover, since the first description reported 40 years ago in a black man with T CD4+cell count below 200/µL [48], immunocompromised patients with T-cell response impairment (i.e., PLWH [49–58], organ transplant recipients [59-69], those on immunosuppressive medication [70-72]) were found to be most at risk of developing cBA due to B. henselae or B. quintana. Despite in some cases retrieved for this analysis T CD4+cell count was reported to be normal [8, 22, 28, 31, 36], we can speculate that in these patients a functional T-cell impairment could explain some of these rare

occurrences. For instance, the patient reported by Kaçar N. et al. was affected by chronic HBV infection [32] and it has been shown that HBV leads to a T-cell functional impairment characterized by compromised cytokine production and upregulation of multiple inhibitory receptors [73]. Moreover, immunosenescence (i.e., agerelated changes that affect T-cell capacity to respond to infections [74]) could impair T-cell functions in elderly patients included in this review [22, 26, 30, 33, 36]. Therefore, although bartonellosis is classically considered as a disease affecting immunocompromised hosts, patients not suffering from major immunocompromising conditions may also be affected by cBA, as emerges from this review.

The complex and not completely understood interactions between host T-cell responses and Bartonella spp. virulence factors mentioned above lead to the red to violaceous lesions which are clinically seen even in not severely immunocompromised patients with cBA and the typical granulomatous and angioproliferative lesions of cBA which can be observed on microscopic examination [8, 11, 14-22, 26-33, 36]. In such cases, diagnosis could be very tricky for several reasons. First, physicians do not include cBA in differential diagnosis since it is considered an infectious disease affecting only severely immunocompromising patients. Second, cBA lesions can be mistaken for similar ones due to both infective (e.g., Mycobacterium spp., Nocardia spp., Sporothrix spp., Histoplasma spp.) or other (e.g., neoplasm, trauma) causes [8, 11, 14, 15]. Third, Bartonella spp. are seldom isolated from cutaneous specimens since they are difficult to culture [8]. Fourth, PCR is not widely available in all settings. Indeed, among the cases we identified, it was performed only in three patients to diagnose cBA [8, 22]. For all these reasons, cBA could be underdiagnosed [3, 14] thus it is likely that its prevalence is underestimated, although an increase in incidence should be expected in the next years [75].

Antibiotic therapy is the mainstay treatment for cBA and different schemes were used. Several drugs such as macrolides, aminoglycosides, rifampin, ciprofloxacin and β -lactam antibiotics are active against *Bartonella* spp [12, 21].. Erythromycin is considered the first choice in PLWH and, even if more studies are needed to support this indication, in this analysis its effectiveness is likewise confirmed in patients without major immunocompromising conditions. Moreover, it has been suggested that this drug may modulate the pathological angiogenesis mediated by Bartonella spp [76].. Doxycycline should also be considered as a valid treatment option since there were no differences in relapses between the use of erythromycin and this drug [24]. However, this finding refers to PLWH and no data are available in patients without HIV infection or not suffering from the other major

immunocompromising conditions listed above. By contrast, several failures and relapses were reported after treatment with aminoglycosides, trimethoprim/sulfamethoxazole and β-lactam antibiotics in angioproliferative lesions due to Bartonella spp [27].. As emerged from this analysis including patients not suffering from major immunocompromising conditions, a single-drug regimen administered for 42 (IQR: 14-47) days could be effective in treating a single cBA lesion, while multiple cBA lesions could require longer courses. This finding suggests that both host immune status and clinical features of cutaneous lesions (i.e., single or multiple) should be taken into consideration when prescribing treatment of cBA concerning type and duration. Indeed, longer antibiotic course and/or combined antimicrobial treatment is the standard of care in PLWH affected by cBA [24] as well as in other cutaneous infectious diseases [77]. However, this hint must be validated by further studies as for cBA concern since we were not able to conclude which is the best available treatment or whether the shorter therapy is more effective in these patients. Moreover, in cases with only a single lesion it could be interesting to know whether the treatment was motivated to prevent future dissemination of the disease, or by a persistent, remaining lesion. Unfortunately, we do not have enough data to address this intriguing question, as it was not answered by the literature that we have retrieved for the purpose of this review. However we suspect that the rationale for treatment may have been both to prevent future dissemination and treat persistent/remaining lesions.

Lastly, global warming is becoming a driver of major health problems in Europe, where the average air temperature has recently risen by one degree Celsius more than in the other continents [78]. In fact, several arthropodborne infectious diseases such as Chikungunya, Dengue, West Nile and Zika are associated with global warming [78] and also Bartonella spp. incidence could be influenced by climate changes [79]. In particular, Bartonella spp. is susceptible to human intervention on environment and a selective pressure on vectors makes bartonellosis a possible re-emerging disease [2]. For instance, most studied vectors such as Ctenocephalides felis prefer warm climates (optimal temperature between 27 °C and 32 °C) [80]. Moreover, pets living in close contact with humans such as dogs and cats from warm countries have both a higher number of potential vectors and levels of bacteremia [46, 81, 82]. Therefore, the wide variety of Bartonella spp [75]., of pets as reservoirs [46, 81, 82] and of vectors [3, 38, 39] involved in human pathology as well as the recent climate changes make the prevalence of cBA highly dynamic and complex. Indeed, our review shows that cBA was mostly reported in warmer countries (India, Turkey, United States of America), while in Europe only three cases of cBA in patients not suffering from major immunocompromising conditions were reported [8, 21, 36]. In these countries, recent climate change could explain the emergence of cBA since it was not previously reported, although it cannot be excluded that the recent increase was due to more frequent diagnoses subsequent to the greater awareness of the problem and to a more widespread use of molecular biology techniques [12, 75]. To this regard, since PCR method allows to perform diagnosis using several samples, (such as frozen, paraffin embedded and lymph node tissues [83]), researchers should implement high-throughput sensitive techniques to identify Bartonella spp. DNA in line with what has been developed for other arthropod-born infections, by combining a broad-range PCR amplification of highly-conserved DNA regions (i.e., gene encoding the 16 S rRNA) with temporal temperature gradient gel electrophoresis [84]. However, despite preanalytical factors can hinder assay yield since formalin-fixed and paraffinembedded tissues are less suitable than fresh-frozen ones for molecular diagnostic purpose [75], sequencing of specie-specific genes (i.e. 16-23 S rRNA internal transcribed spacer and citrate synthase regions) followed by molecular analysis can improve the routine data and identify serotype/genotype of Bartonella strains [85, 86]. This technique will likely be applied in high-income countries to help clinicians in diagnosis and follow-up of Bartonella spp. infections.

In summary, this review confirmes that cBA appears to be very rare in patients not suffering from major immunocompromising conditions but it also probably remains underdiagnosed due to the limited availability of molecular tests. However, in the era of next-generation sequencing, an active surveillance of re-emerging pathogens needs to be improved using molecular testing [87, 88]. Moreover, it is not possible to conclude which is the best available treatment or whether the shorter therapy is more effective in these patients. Indeed, the patients retrieved were very heterogeneous in terms of age and in several cases comorbidities were not reported, implying that further studies should be put in place to obtain more representative cohorts. Finally, in most of the cases reported the diagnosis of cBA was made by histological examination, while culture or PCR was performed in a minority of cases [8, 22, 27]. In this regard, despite cBA and verruga peruana are angioproliferative lesions that are not clinically or histopathologically distinguishable [23], only one case retrieved from the literature was reported in a Country where verruga peruviana is known to be endemic [34]. This observation further underscores the crucial role of species-specific PCR in facilitating species diagnosis. This method should be standardized and widely adopted for both diagnosis and treatment monitoring, potentially elevating the accuracy and efficiency of patient care across a variety of clinical settings [87, 88]..

Conclusions

Since both B. quintana and B. henselae are responsible for a wide variety of cBA lesions which can be mistaken for similar ones due to others causes, *Bartonella* spp. are difficult to culture and PCR is not widely available in all settings, diagnosis is challenging and the burden of cBA in non severely immunocompromised patients could be overlooked. Therefore, the implementation of molecular testing is a necessary high-sensitivity test that could enable to treat and uncover the real burden in such cases. In conclusion, clinicians should consider cBA as a possible clinical manifestation of B. quintana or B. henselae infection even in patients not suffering from major immunocompromising conditions. Erythromycin should be considered the first choice while doxycycline and clarithromycin are valid alternatives. Rifampin may be useful in combination in some difficult to treat cases. A median antibiotic course of 42 and 90 days could be effective in single and multiple cBA lesions, respectively, but studies including more patients are needed to assess which is the most appropriate therapy for cBA. This is another piece of evidence that more attention should be given to a one health approach for prevention of infectious diseases in the current World.

Abbreviations

- cBA cutaneous bacillary angiomatosis
- HIV human immunodeficiency virus
- PLWH people living with HIV
- NIH National Institute of Health
- PCR polymerase chain reaction TLR toll-like receptor

Acknowledgements

The authors thank all the clinical staff members who take care of patients everyday at the "Renato Dulbecco" Teaching Hospital of Calabria Region, Italy.

Author contributions

SR was responsible for the idea and study design under the supervision of CT. SR and MTT reviewed literature. SR, MTT, HLM, AR, EMT and CT handled the clinical section of the review. NM, SG, AQ, GM handled the microbiological section of the review. SR performed statistical analysis. HLM reviewed English language. All the authors drafted and performed a critical revision of the manuscript and provided important intellectual contents.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval

Since the review included only case reports previously published, both the need for approval by ethical committee of the Calabria Region (Italy) and the consent for publication were waived.

Conflict of interest

The authors declare that they have no competing interests.

Competing interests

The authors declare no competing interests.

Received: 19 November 2023 / Accepted: 25 March 2024 Published online: 08 April 2024

References

- Adal KA, Cockerell CJ, Petri WAJNEJM Jr. Cat scratch disease, bacillary angiomatosis, and other infections due to Rochalimaea. 1994, 330(21):1509–15.
- Breitschwerdt EB. Bartonellosis, One Health and all creatures great and small. Vet Dermatol. 2017;28(1):96–e21.
- Mullins K, Canal E, Ouch P, Prasetyo D, Tagoe J, Attram N, Yeboah C, Kumordjie S, Fox A, Letizia AG, et al. Bartonella Species in Cambodia, Ghana, Laos, and Peru: results from Vector and Serosurveys. Vector Borne Zoonotic Dis. 2023;23(1):9–17.
- Koehler JE, Cederberg L. Intra-abdominal mass associated with gastrointestinal hemorrhage: a new manifestation of bacillary angiomatosis. Gastroenterology. 1995;109(6):2011–4.
- Billeter SA, Levy MG, Chomel BB, Breitschwerdt EB. Vector transmission of Bartonella species with emphasis on the potential for tick transmission. Med Vet Entomol. 2008;22(1):1–15.
- Regnery R, Tappero J. Unraveling mysteries associated with cat-scratch disease, bacillary angiomatosis, and related syndromes. Emerg Infect Dis. 1995;1(1):16–21.
- Reis C, Cote M, Le Rhun D, Lecuelle B, Levin ML, Vayssier-Taussat M, Bonnet SI. Vector competence of the tick Ixodes ricinus for transmission of Bartonella birtlesii. PLoS Negl Trop Dis. 2011;5(5):e1186.
- Bernabeu-Wittel J, Luque R, Corbi R, Mantrana-Bermejo M, Navarrete M, Vallejo A, Bernabeu-Wittel M. Bacillary angiomatosis with atypical clinical presentation in an immunocompetent patient. Indian J Dermatol Venereol Leprol. 2010;76(6):682–5.
- Sanchez Clemente N, Ugarte-Gil CA, Solorzano N, Maguina C, Pachas P, Blazes D, Bailey R, Mabey D, Moore D. Bartonella bacilliformis: a systematic review of the literature to guide the research agenda for elimination. PLoS Negl Trop Dis. 2012;6(10):e1819.
- Huarcaya E, Best I, Rodriguez-Tafur J, Maguina C, Solorzano N, Menacho J, Lopez De Guimaraes D, Chauca J, Ventosilla P. Cytokines and T-Lymphocute count in patients in the acute and chronic phases of Bartonella bacilliformis infection in an endemic area in Peru: a pilot study. Rev Inst Med Trop Sao Paulo. 2011;53(3):149–54.
- Zarraga M, Rosen L, Herschthal D. Bacillary angiomatosis in an immunocompetent child: a case report and review of the literature. Am J Dermatopathol. 2011;33(5):513–5.
- Mogollon-Pasapera E, Otvos L Jr., Giordano A, Cassone M. Bartonella: emerging pathogen or emerging awareness? Int J Infect Dis. 2009;13(1):3–8.
- Koehler JE, Sanchez MA, Garrido CS, Whitfeld MJ, Chen FM, Berger TG, Rodriguez-Barradas MC, LeBoit PE, Tappero JW. Molecular epidemiology of bartonella infections in patients with bacillary angiomatosis-peliosis. N Engl J Med. 1997;337(26):1876–83.
- Agrawal S, Singal A, Arora VK. Bacillary Angiomatosis in an Immunocompetent patient: an unusual occurrence. Indian Dermatol Online J. 2022;13(4):527–9.
- Nikam BP, Vijayendran N, Jamale V, Kale M. Bacillary Angiomatosis in an Immunocompetent Individual. Indian Dermatol Online J. 2018;9(3):205–6.
- Karakas M, Baba M, Aksungur VL, Homan S, Memisoglu HR, Uguz A. Bacillary angiomatosis on a region of burned skin in a immunocompetent patient. Br J Dermatol. 2000;143(3):609–11.
- Karakas M, Baba M, Homan S, Akman A, Acar MA, Memisoglu HR, Gumurdulu D. A case of bacillary angiomatosis presenting as leg ulcers. J Eur Acad Dermatol Venereol. 2003;17(1):65–7.
- Asharaf M, Letha S. Cutaneous bacillary angiomatosis. Indian J Pediatr. 2002;69(11):1003–5.
- Turgut M, Alabaz D, Karakas M, Kavak M, Aksaray N, Alhan E, Cevlik F, Tuncer I. Bacillary angiomatosis in an immunocompetent child with a grafted traumatic wound. J Dermatol. 2004;31(10):844–7.

- Albayrak A, Albayrak Y, Unal D, Atasoy M, Uyanik MH. A case of bacillary angiomatosis developed at a burn site. Indian J Dermatol Venereol Leprol. 2012;78(1):121.
- Balaban M, Ioana Nedelcu R, Balmes G, Adela Todorovic T, Brinzea A, Nichita L, Gabriela Popp C, Theodor Andrei R, Andrada Zurac S, Adriana Ion D, et al. Bacillary angiomatosis triggered by severe trauma in a healthy caucasian patient: a case report. Exp Ther Med. 2020;20(1):56–60.
- Tappero JW, Koehler JE, Berger TG, Cockerell CJ, Lee TH, Busch MP, Stites DP, Mohle-Boetani J, Reingold AL, LeBoit PE. Bacillary angiomatosis and bacillary splenitis in immunocompetent adults. Ann Intern Med. 1993;118(5):363–5.
- 23. Jimenez-Lucho V. Images in clinical medicine. Verruga peruana. N Engl J Med. 1998;339(7):450.
- 24. Panel on Opportunistic Infections in Adults and Adolescents with HIV. Guidelines for the prevention and treatment of opportunistic infections in adults and adolescents with HIV: recommendations from the Centers for Disease Control and Prevention, the National Institutes of Health, and the HIV Medicine Association of the Infectious Diseases Society of America. http:// aidsinfo.nih.gov/contentfiles/lvguidelines/adult_oi.pdf. Accessed October 9, 2023.
- Moher D, Shamseer L, Clarke M, Ghersi D, Liberati A, Petticrew M, Shekelle P, Stewart LA. Group P-P: Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. Syst Rev. 2015;4(1):1.
- 26. Gangopadhyay AK, Sharma PK. Bacillary angiomatosis in an immune-competent patient. Indian J Dermatol Venereol Leprol. 2001;67(1):37–8.
- Cockerell CJ, Bergstresser PR, Myrie-Williams C, Tierno PM. Bacillary epithelioid angiomatosis occurring in an immunocompetent individual. Arch Dermatol. 1990;126(6):787–90.
- Iraji F, Pourazizi M, Abtahi-Naeini B, Meidani M, Rajabi P. Bacillary Angiomatosis in Immunocompetent patient with atypical manifestations. Indian J Dermatol. 2015;60(5):523.
- Paul MA, Fleischer AB Jr., Wieselthier JS, White WL. Bacillary angiomatosis in an immunocompetent child: the first reported case. Pediatr Dermatol. 1994;11(4):338–41.
- Kayaselcuk F, Ceken I, Bircan S, Tuncer I. Bacillary angiomatosis of the scalp in a human immunodeficiency virus-negative patient. J Eur Acad Dermatol Venereol. 2002;16(6):612–4.
- 31. Smith KJ, Skelton HG, Tuur S, Larson PL, Angritt P. Bacillary angiomatosis in an immunocompetent child. Am J Dermatopathol. 1996;18(6):597–600.
- 32. Kacar N, Tasli L, Demirkan N, Ergin C, Ergin S. HIV-negative case of bacillary angiomatosis with chronic hepatitis B. J Dermatol. 2010;37(8):722–5.
- Blattner C, Jacobson-Dunlop E, Miller JH, Elston DM. A case of bacillary angiomatosis in a patient with pancreatic adenocarcinoma. J Cutan Pathol. 2014;41(3):277–80.
- 34. Bellissimo-Rodrigues F, da Fonseca BA, Martinez R. Bacillary angiomatosis in a pregnant woman. Int J Gynaecol Obstet. 2010;111(1):85–6.
- 35. Omidian M, Ranjbari N, Omidian EJIJD. Bacillary angiomatosis in an immune competent patient: a case report from Iran. 2014, 17(1):39–41.
- Rotundo S, Bono F, Mazzitelli M, Scaglione V, Lamberti AG, Giancotti A, Tucci L, Costa C, Tassone MT, Morrone HL et al. An autochthonous case of cutaneous bacillary angiomatosis not related to major immunosuppression: an emerging or overlooked disease? Int J Infect Dis 2023.
- Harms A, Dehio C. Intruders below the radar: molecular pathogenesis of Bartonella Spp. Clin Microbiol Rev. 2012;25(1):42–78.
- Valerio CR, Murray P, Arlian LG, Slater JE. Bacterial 16S ribosomal DNA in house dust mite cultures. J Allergy Clin Immunol. 2005;116(6):1296–300.
- Murillo N, Mediannikov O, Aubert J, Raoult D. Bartonella quintana detection in Demodex from erythematotelangiectatic rosacea patients. Int J Infect Dis. 2014;29:176–7.
- Jin X, Gou Y, Xin Y, Li J, Sun J, Li T, Feng J. Advancements in understanding the molecular and immune mechanisms of Bartonella pathogenicity. Front Microbiol. 2023;14:1196700.
- Lionello FCP, Rotundo S, Bruno G, Marino G, Morrone HL, Fusco P, Costa C, Russo A, Trecarichi EM, Beltrame A et al. Touching Base with Some Mediterranean Diseases of Interest from Paradigmatic Cases at the Magna Graecia University Unit of Infectious Diseases: A Didascalic Review. *Diagnostics (Basel)* 2023, 13(17).
- 42. Malgorzata-Miller G, Heinbockel L, Brandenburg K, van der Meer JW, Netea MG, Joosten LA. Bartonella quintana lipopolysaccharide (LPS): structure and characteristics of a potent TLR4 antagonist for in-vitro and in-vivo applications. Sci Rep. 2016;6:34221.
- 43. Scutera S, Mitola S, Sparti R, Salvi V, Grillo E, Piersigilli G, Bugatti M, Alotto D, Schioppa T, Sozzani S, et al. Bartonella henselae persistence

within mesenchymal stromal cells enhances endothelial cell activation and infectibility that amplifies the angiogenic process. Infect Immun. 2021;89(8):e0014121.

- Kempf VA, Volkmann B, Schaller M, Sander CA, Alitalo K, Riess T, Autenrieth IB. Evidence of a leading role for VEGF in Bartonella henselae-induced endothelial cell proliferations. Cell Microbiol. 2001;3(9):623–32.
- 45. Angelakis E, Raoult D. Pathogenicity and treatment of Bartonella infections. Int J Antimicrob Agents. 2014;44(1):16–25.
- 46. Brenner EC, Chomel BB, Singhasivanon OU, Namekata DY, Kasten RW, Kass PH, Cortes-Vecino JA, Gennari SM, Rajapakse RP, Huong LT, et al. Bartonella infection in urban and rural dogs from the tropics: Brazil, Colombia, Sri Lanka and Vietnam. Epidemiol Infect. 2013;141(1):54–61.
- 47. Resto-Ruiz S, Burgess A, Anderson BE. The role of the host immune response in pathogenesis of Bartonella henselae. DNA Cell Biol. 2003;22(6):431–40.
- Waldo E, Sidhu GS, Stahl R, Zolla-Pazner S. Histiocytoid hemangioma with features of angiolymphoid hyperplasia and Kaposi's sarcoma. A study by light microscopy, electron microscopy, and immunologic techniques. Am J Dermatopathol. 1983;5(6):525–38.
- Baron AL, Steinbach LS, LeBoit PE, Mills CM, Gee JH, Berger TG. Osteolytic lesions and bacillary angiomatosis in HIV infection: radiologic differentiation from AIDS-related Kaposi sarcoma. Radiology. 1990;177(1):77–81.
- Fagan WA, DeCamp NC, Kraus EW, Pulitzer DR. Widespread cutaneous bacillary angiomatosis and a large fungating mass in an HIV-positive man. J Am Acad Dermatol. 1996;35(2 II):285–7.
- Fagan WA, Skinner SM, Ondo A, Williams JT, Anthony K, DeVillez RL, Pulitzer DR. Bacillary angiomatosis of the skin and bone marrow in a patient with HIV infection. J Am Acad Dermatol. 1995;32(3):510–2.
- Hettmannsperger U, Soehnchen R, Gollnick H, Detmar M, Orfanos CE. Bacillary epithelioid angiomatosis in advanced HIV infection. Hautarzt. 1993;44(12):803–7.
- Husain S, Singh N. Pyomyositis associated with bacillary angiomatosis in a patient with HIV infection. Infection. 2002;30(1):50–3.
- Mateen FJ, Newstead JC, McClean KL. Bacillary angiomatosis in an HIVpositive man with multiple risk factors: a clinical and epidemiological puzzle. Can J Infect Dis Med Microbiol. 2005;16(4):249–52.
- Monteil RA, Michiels JF, Hofman P, Saint-Paul MC, Hitzig C, Perrin C, Santini J. Histological and ultrastructural study of one case of oral bacillary angiomatosis in HIV disease and review of the literature. Eur J Cancer Part B: Oral Oncol. 1994;30(1):65–71.
- Murillo O, Mimbrera D, Petit A, Gil H, Anda P, Carrera M, Podzamczer D. Fatal bacillary angiomatosis mimicking an infiltrative vascular tumour in the immune restoration phase of an HIV-infected patient. Antivir Ther. 2012;17(2):405–7.
- Plettenberg A, Lorenzen T, Burtsche BT, Rasokat H, Kaliebe T, Albrecht H, Mertenskotter T, Bogner JR, Stoehr A, Schofer H. Bacillary angiomatosis in HIV-infected patients–an epidemiological and clinical study. Dermatology. 2000;201(4):326–31.
- Rodriguez O, Campbell LR, Bacha JM, Kovarik CL. Successful treatment of bacillary angiomatosis with oral doxycycline in an HIV-infected child with skin lesions mimicking Kaposi sarcoma. JAAD Case Rep. 2016;2(1):77–9.
- Brzewski P, Kwiecińska M, Sułowicz J, Podolec K, Obtułowicz A, Dyduch G, Wojas-Pelc A. Bacillary Angiomatosis in Renal Transplant recipient: a Case Report. Transpl Proc. 2020;52(8):2524–6.
- 60. Cline MS, Cummings OW, Goldman M, Filo RS, Pescovitz MD. Bacillary angiomatosis in a renal transplant recipient. Transplantation. 1999;67(2):296–8.
- Dardenne S, Coche E, Weynand B, Poncelet A, Zech F, De Meyer M. High suspicion of Bacillary Angiomatosis in a kidney transplant recipient: a difficult way to diagnose-Case Report. Transpl Proc. 2007;39(1):311–3.
- Eid R, Assayag M, Lefevre E, Escaut L, Laifi M, Brodin-Sartorius A, Zaidan M, Snanoudj R. Invasive bacillary angiomatosis in a kidney transplant recipient: a challenging case on belatacept immunosuppression. Int J Infect Dis. 2023;133:43–5.
- 63. Grabas M, Darrieux L, Potier J, Safa G. Hemophagocytic syndrome as the presenting manifestation of bacillary angiomatosis in a renal transplant recipient. J Am Acad Dermatol. 2012;67(5):e236–237.
- 64. Helleberg M. Bacillary angiomatosis in a solid organ transplant recipient. IDCases. 2019;18:e00649.
- 65. Juskevicius R, Vnencak-Jones C. Pathologic quiz case: a 17-year-old renal transplant patient with persistent fever, pancytopenia, and axillary lymphadenopathy. Bacillary angiomatosis of the lymph node in the renal transplant recipient. Arch Pathol Lab Med. 2004;128(1):e12–14.

- Mehrmal S, Mhlaba JM, Zhou XA. Cutaneous bacillary angiomatosis in a renal transplant patient. Skinmed. 2021;19(2):150–4.
- Morillas JA, Hassanein M, Syed B, Liaqat A, Bergfeld W, Sardiña LA, Fatica R, Lum J. Early post-transplant cutaneous bacillary angiomatosis in a kidney recipient: case report and review of the literature. Transpl Infect Disease 2021, 23(4).
- Orsag J, Flodr P, Melter O, Tkadlec J, Sternbersky J, Hruby M, Klicova A, Zamboch K, Krejci K, Zadrazil J. Cutaneous bacillary angiomatosis due to Bartonella quintana in a renal transplant recipient. Transpl Int. 2015;28(5):626–31.
- 69. Vetos D, Rickstrew J, Siscos S, Wang T. A case of bacillary angiomatosis following solid organ transplant. Int J Dermatol. 2022;61(7):e257–9.
- Aydoğan I, Parlak AH, Alper M, Aksoy KA. Bacillary angiomatosis in an HIV seronegative patient. Turkderm Deri Hastaliklari Ve Frengi Arsivi. 2004;38(1):71–4.
- Schwartz RA, Gallardo MA, Kapila R, Gascon P, Herscu J, Siegel I, Lambert WC. Bacillary angiomatosis in an HIV seronegative patient on systemic steroid therapy. Br J Dermatol. 1996;135(6):982–7.
- 72. Kreitzer T, Saoud A. Bacillary angiomatosis following the use of long-term methotrexate therapy: a case report. West Va Med J. 2006;102(1):317–8.
- Ye B, Liu X, Li X, Kong H, Tian L, Chen Y. T-cell exhaustion in chronic hepatitis B infection: current knowledge and clinical significance. Cell Death Dis. 2015;6(3):e1694.
- Rodriguez IJ, Lalinde Ruiz N, Llano Leon M, Martinez Enriquez L, Montilla Velasquez MDP, Ortiz Aguirre JP, Rodriguez Bohorquez OM, Velandia Vargas EA, Hernandez ED. Parra Lopez CA: Immunosenescence Study of T cells: a systematic review. Front Immunol. 2020;11:604591.
- McCormick DW, Rassoulian-Barrett SL, Hoogestraat DR, Salipante SJ, Sen-Gupta D, Dietrich EA, Cookson BT, Marx GE, Lieberman JA. Bartonella Spp. Infections identified by molecular methods, United States. Emerg Infect Dis. 2023;29(3):467–76.
- Meghari S, Rolain JM, Grau GE, Platt E, Barrassi L, Mege JL, Raoult D. Antiangiogenic effect of erythromycin: an in vitro model of Bartonella quintana infection. J Infect Dis. 2006;193(3):380–6.
- 77. Aronson N, Herwaldt BL, Libman M, Pearson R, Lopez-Velez R, Weina P, Carvalho EM, Ephros M, Jeronimo S, Magill A. Diagnosis and treatment of Leishmaniasis: clinical practice guidelines by the Infectious Diseases Society of America (IDSA) and the American Society of Tropical Medicine and Hygiene (ASTMH). Clin Infect Dis. 2016;63(12):e202–64.
- van Daalen KR, Romanello M, Rocklov J, Semenza JC, Tonne C, Markandya A, Dasandi N, Jankin S, Achebak H, Ballester J, et al. The 2022 Europe report of the Lancet countdown on health and climate change: towards a climate resilient future. Lancet Public Health. 2022;7(11):e942–65.

- 79. Chinga-Alayo E, Huarcaya E, Nasarre C, del Aguila R, Llanos-Cuentas A. The influence of climate on the epidemiology of bartonellosis in Ancash, Peru. Trans R Soc Trop Med Hyg. 2004;98(2):116–24.
- Silverman J, Rust MK, Reierson DA. Influence of temperature and humidity on survival and development of the cat flea, Ctenocephalides felis (Siphonaptera: Pulicidae). J Med Entomol. 1981;18(1):78–83.
- Kokkinaki KCG, Saridomichelakis MN, Skampardonis V, Mataragka A, Ikonomopoulos J, Leontides L, Mylonakis ME, Steiner JM, Suchodolski JS, Xenoulis PG. Prevalence and risk factors for Bartonella spp. and Haemoplasma Infections in cats from Greece. Vet Sci 2022, 9(7).
- Sepulveda-Garcia P, Alabi A, Alvarez K, Rojas L, Mella A, Goncalves LR, Andre MR, Machado RZ, Muller A, Monti G. Bartonella spp. in households with cats: risk factors for infection in cats and human exposure. One Health. 2023;16:100545.
- Mitchell BM, Font RL. Molecular detection of bartonella henselae for the diagnosis of cat scratch disease and bacillary angiomatosis of the conjunctiva. Cornea. 2011;30(7):807–14.
- Halos L, Mavris M, Vourc'h G, Maillard R, Barnouin J, Boulouis HJ, Vayssier-Taussat M. Broad-range PCR-TTGE for the first-line detection of bacterial pathogen DNA in ticks. Vet Res. 2006;37(2):245–53.
- La Scola B, Liang Z, Zeaiter Z, Houpikian P, Grimont PA, Raoult D. Genotypic characteristics of two serotypes of Bartonella henselae. J Clin Microbiol. 2002;40(6):2002–8.
- 86. Sala M, Font B, Sanfeliu I, Quesada M, Ponts I, Segura F. Bacillary angiomatosis caused by Bartonella quintana. Ann N Y Acad Sci. 2005;1063:302–7.
- Mazzitelli M, Lamberti AG, Quirino A, Marascio N, Barreca GS, Costa C, Pisani V, Strazzulla A, Greco G, Liberto MC, et al. Utility of molecular identification and Quantitation of Bartonella Species with species-specific real-time PCR for monitoring treatment response: a Case Series. Open Microbiol J. 2018;12:148–53.
- Liberto MC, Lamberti AG, Marascio N, Matera G, Quirino A, Barreca GS, Baudi F, Foca A. Molecular identification of Bartonella quintana infection using species-specific real-time PCR targeting transcriptional regulatory protein (bqtR) gene. Mol Cell Probes. 2011;25(5–6):238–42.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.